

STUDIES ON BUD DORMANCY OF
WOODY SPECIES

A thesis
submitted in partial fulfilment of
the requirements for the degree
of
Doctor of Philosophy in Plant Physiology
in the
University of Canterbury

by
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PART II

(Figures and Plates)

Figure 1. Outline of the generalised procedure used for the purification of free and bound ABA from methanolic extracts of *Alnus viridis* tissue.

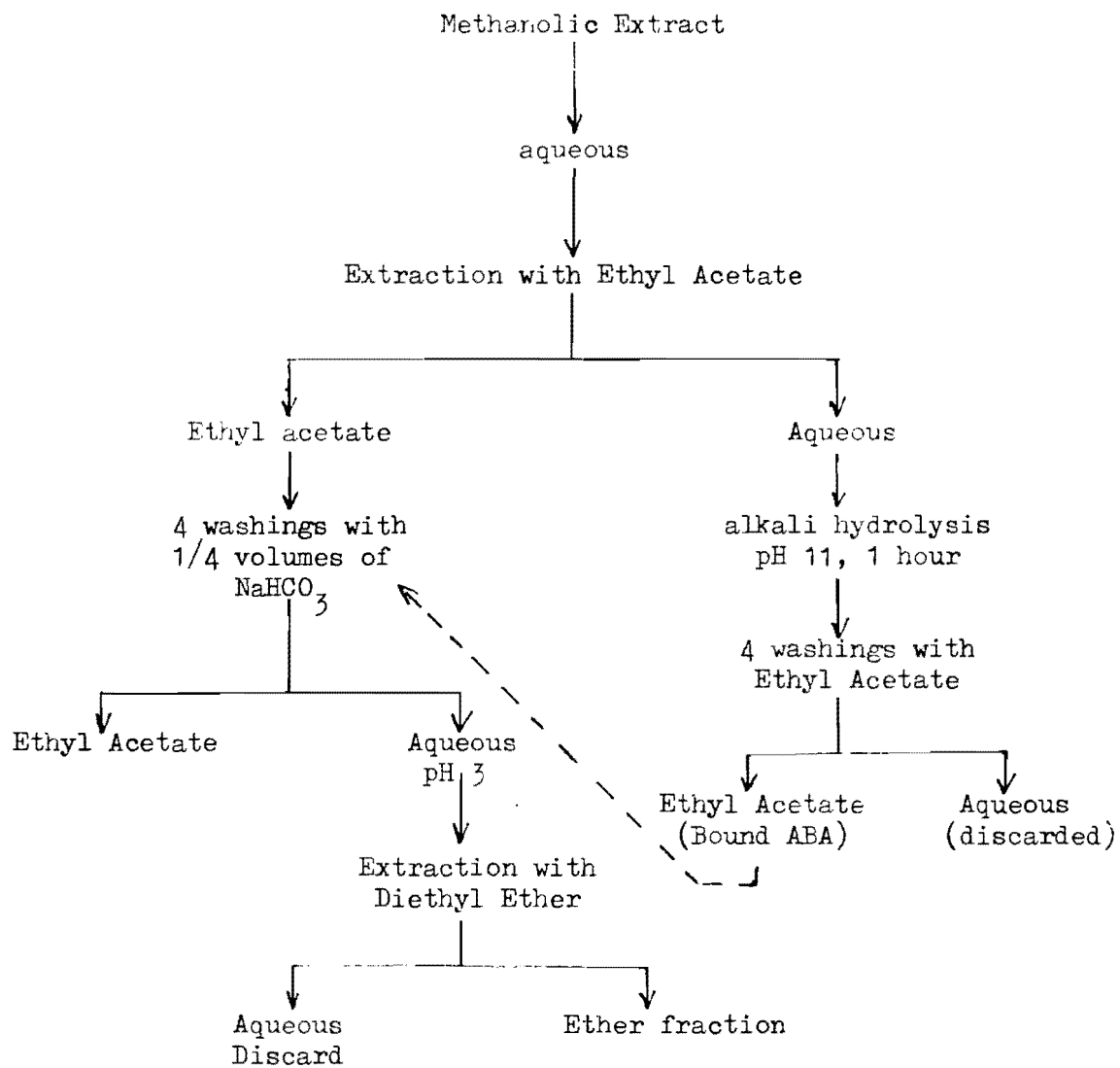


Figure 2. Gas chromatographs of methylated extracts of *Alnus viridis* leaves and buds (A) before and (B) after co-chromatography with the mixed isomers of authentic Me-ABA. Chromatographs were obtained using ^{63}Ni electron capture detector and 3% OV 17 as stationary phase. Column and detector temperatures were 195°C and 295°C , respectively. Samples injected were equivalent to approx. 10^{-3} g DW of tissue and 100 pg of authentic Me-ABA. Arrows indicate peaks corresponding with cis,trans Me-ABA and trans,trans Me-ABA.

- a free ABA fraction of a leaf extract
- b bound ABA fraction of a leaf extract
- c free ABA fraction of an apical extract
- d bound ABA fraction of an apical extract

detector response

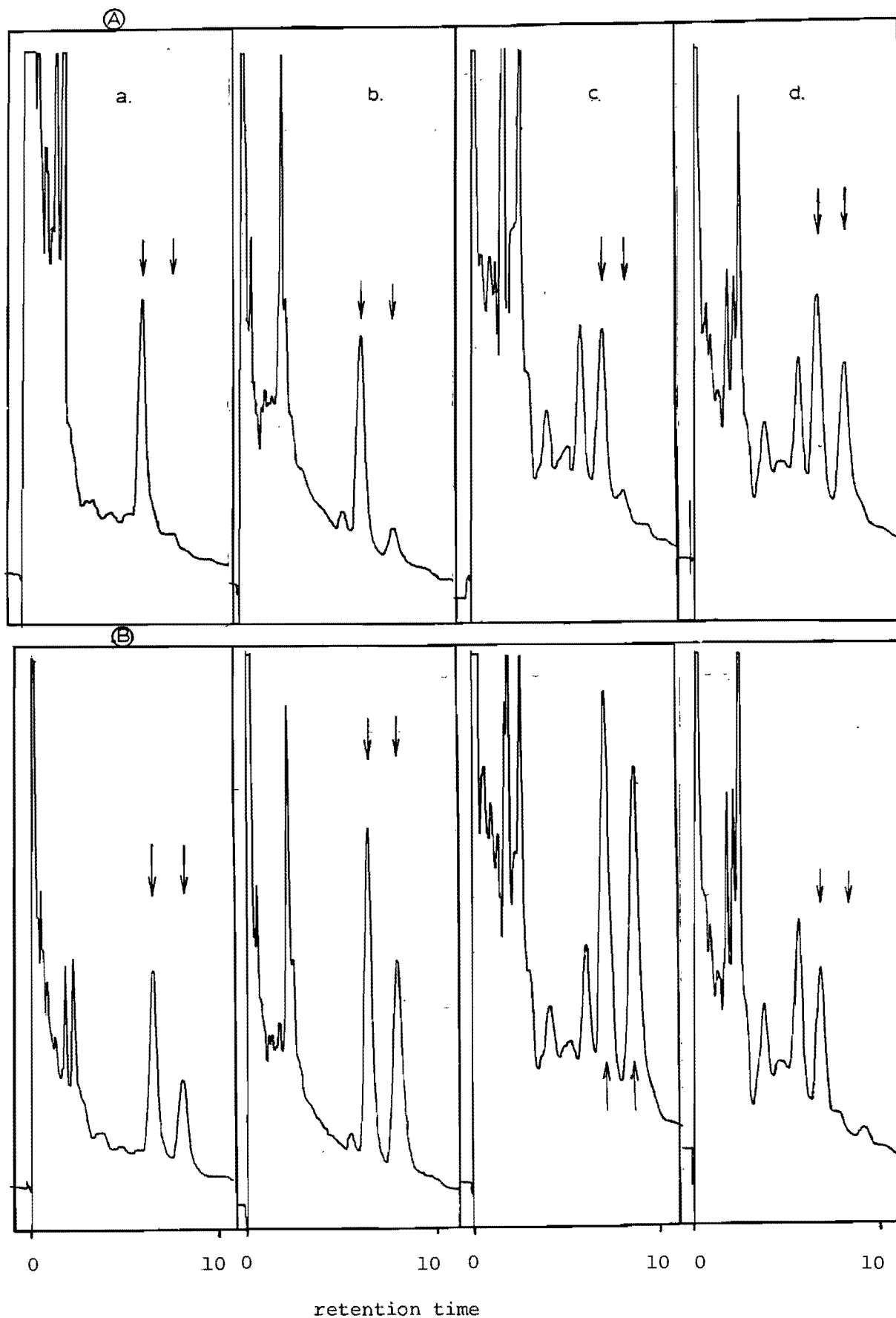
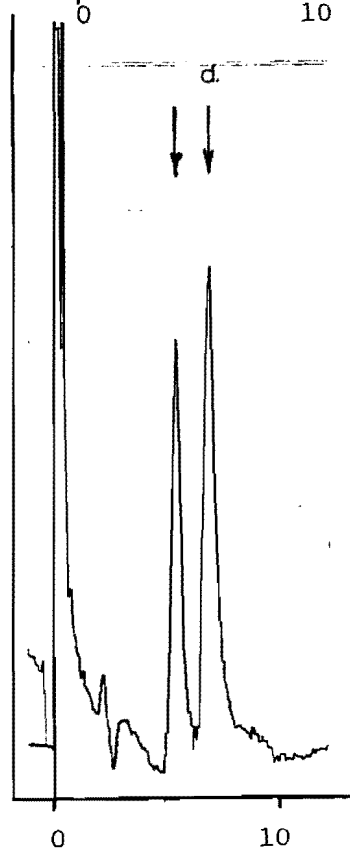
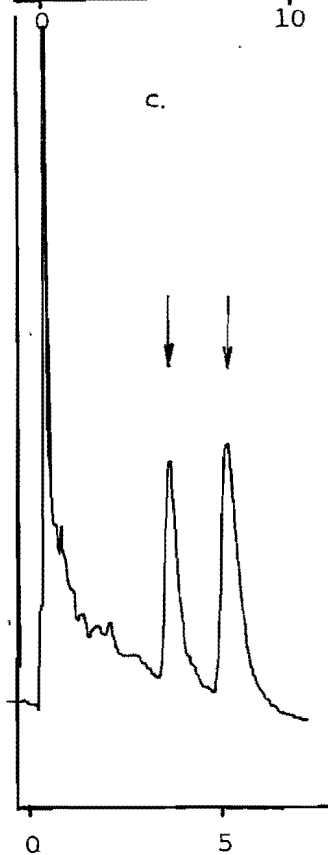
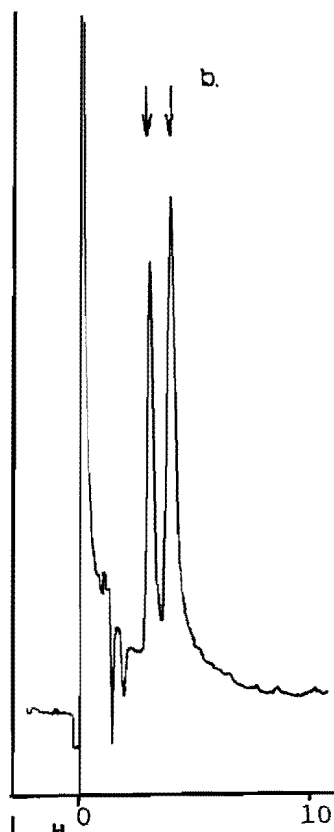
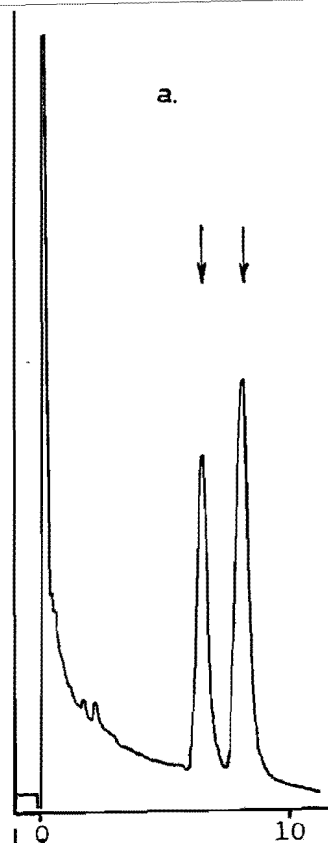


Figure 3. Gas chromatographs of the mixed isomers of authentic ABA on four stationary phases (a) 3% OV 17, (b) 3% OV 1, (c) 3% OV 210, and (d) 10% SE 30 using an electron capture detector. Column temperatures were 195°C, 205°C, 205°C and 210°C, respectively and the detector temperature was 195°C. Samples injected represent 100 pg of authentic Me-ABA. Arrows indicate peaks corresponding with cis,trans Me-ABA and trans,trans Me-ABA.

detector response



retention time (min)

Figure 4. Gas chromatographs of methylated extracts of *Alnus viridis* leaves and buds obtained using an electron capture detector and (A) 3% OV 210, (B) 3% OV1, and (C) 10% SE 30 as stationary phases. Column and detector temperatures were as in figure 3 and other details as in figure 2.

- (a) free ABA fraction of a leaf extract
- (b) bound ABA fraction of a leaf extract
- (c) free ABA fraction of an apical extract
- (d) bound ABA fraction of an apical extract

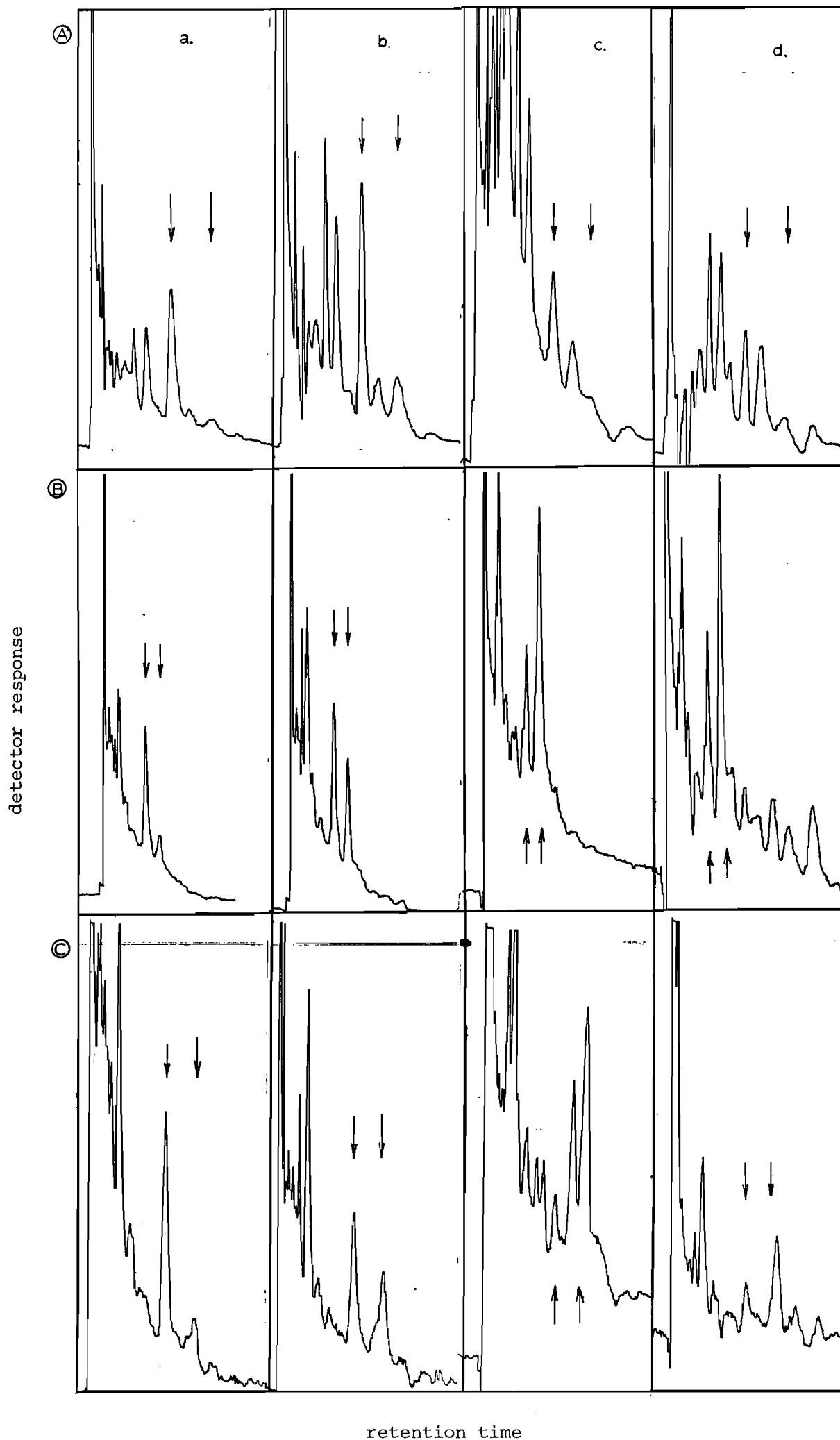


Figure 5. Gas chromatographs of a methylated extract of *Alnus viridis* leaves obtained (A) before and (B) after co-chromatography with the mixed isomers of authentic Me-ABA. Chromatographs were obtained using an electron capture detector and (a) 3% OV 17, (b) 3% OV 1, (c) 3% OV 210 and (d) 10% SE 30 as stationary phases. Samples injected represent approx. 10^{-3} g DW Of tissue. Other details as in figure 3.

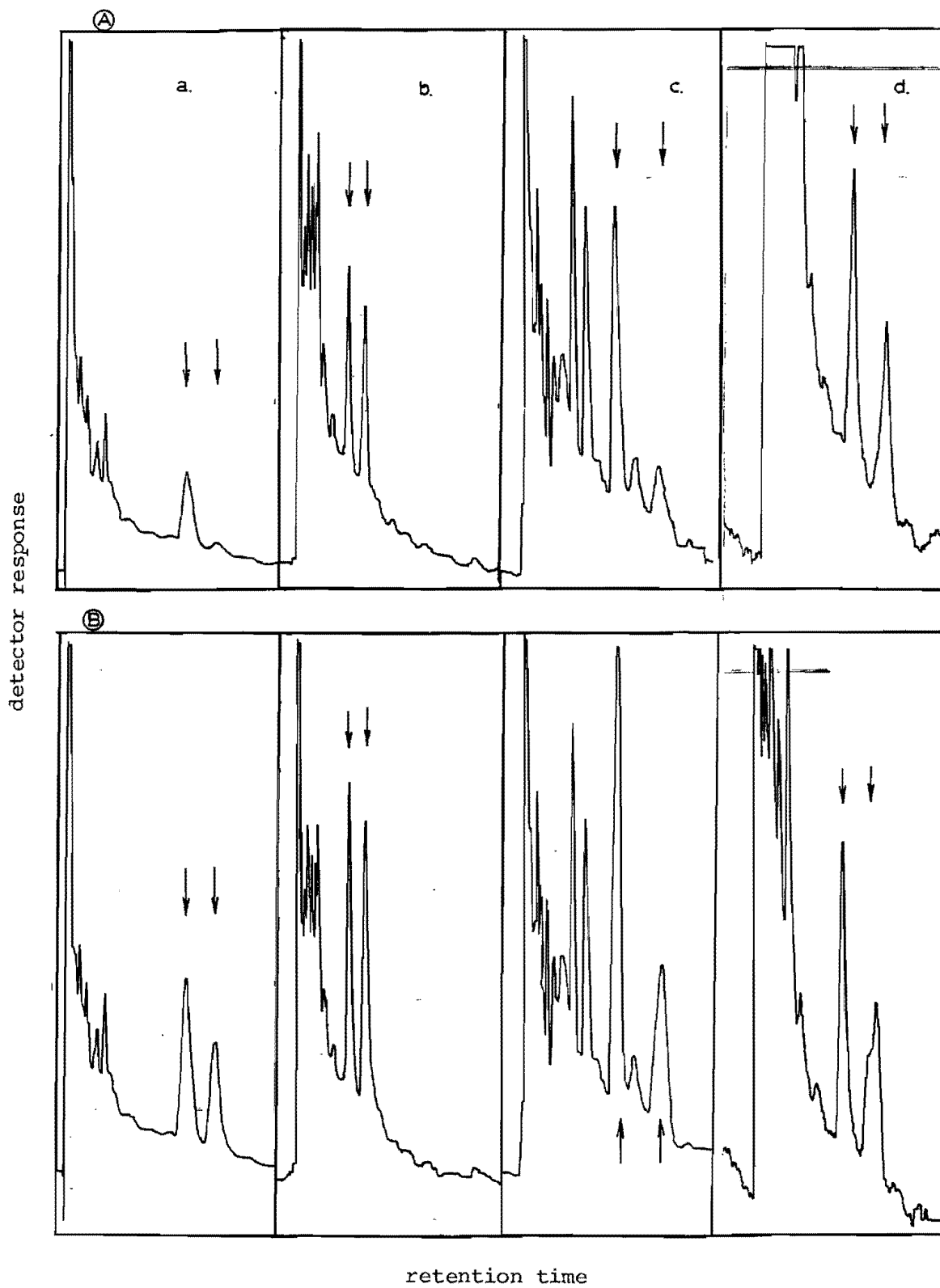
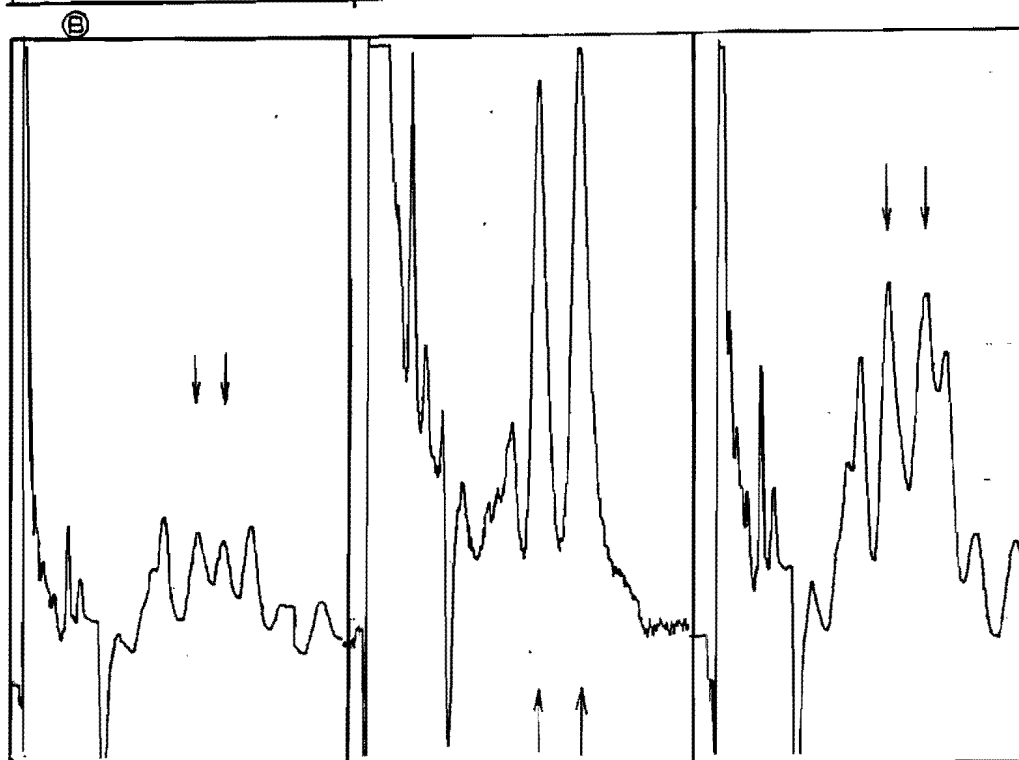
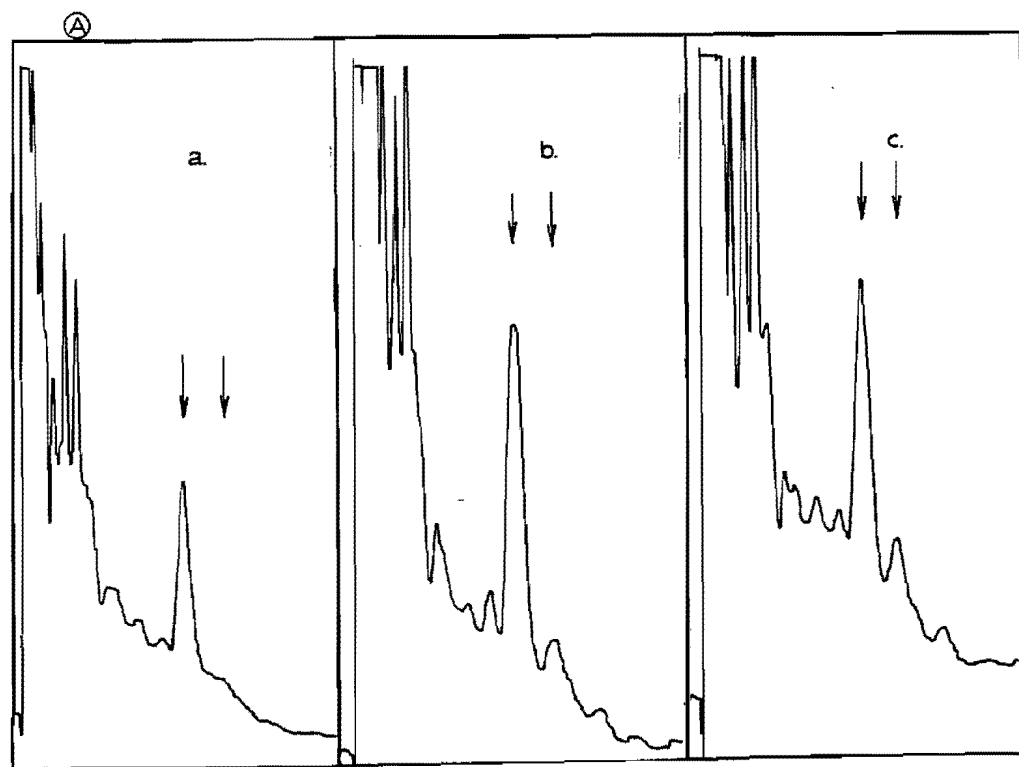


Figure 6. Gas chromatographs of methylated extracts of *Alnus viridis* leaves (A) before and (B) after 1 hour of U.V. light exposure, obtained using an electron capture detector and 3% OV 17 as the stationary phase. Other details as in figure 2.

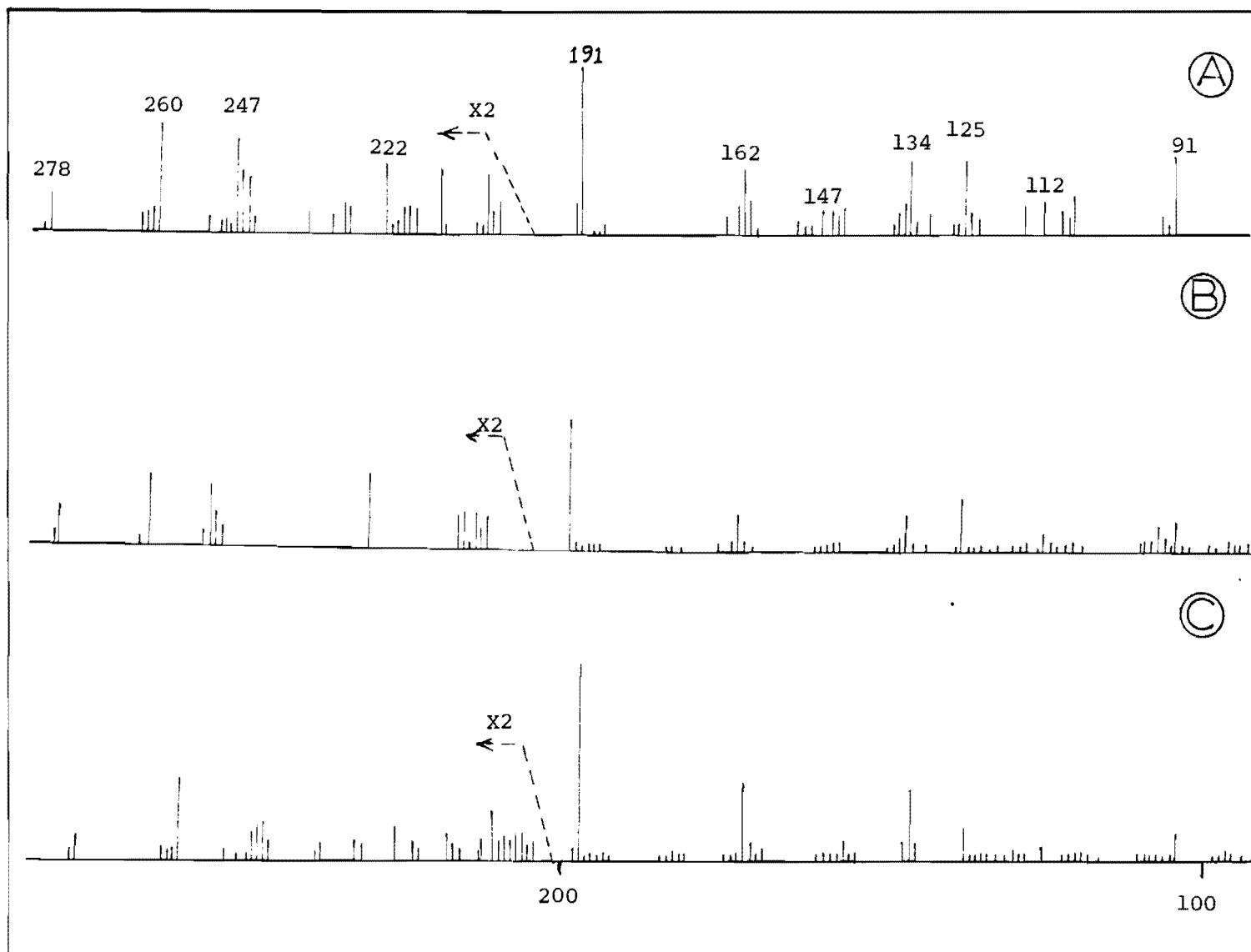
detector response



retention time

Figure 7. Mass spectrum of the mixed isomers of authentic Me-ABA (A) and the methylated extracts of *Alnus viridis* (B) leaves and (C) apices. Ionizing voltage was 70 eV.

relative abundance



m/e

Figure 8. Growth of *Alnus viridis* seedlings under 16 h (LD) and then transfer to 8 h (SD) photoperiods. Plants were maintained at 10°C night and 14°C day (equivalent to the high light intensity period). Arrow indicates the beginning of the SD treatment. Each value represents the percentage increase in height and is the mean of 10 replicates (LD's) or 7 replicates to day 12 and 4 replicates after day 12 of SD treatment. Each replicate consists of two plants and the vertical bars represent twice the S.E.'s of the means.

Leaves and apices were harvested from 3 replicates on day 0 and day 12 of SD treatment.

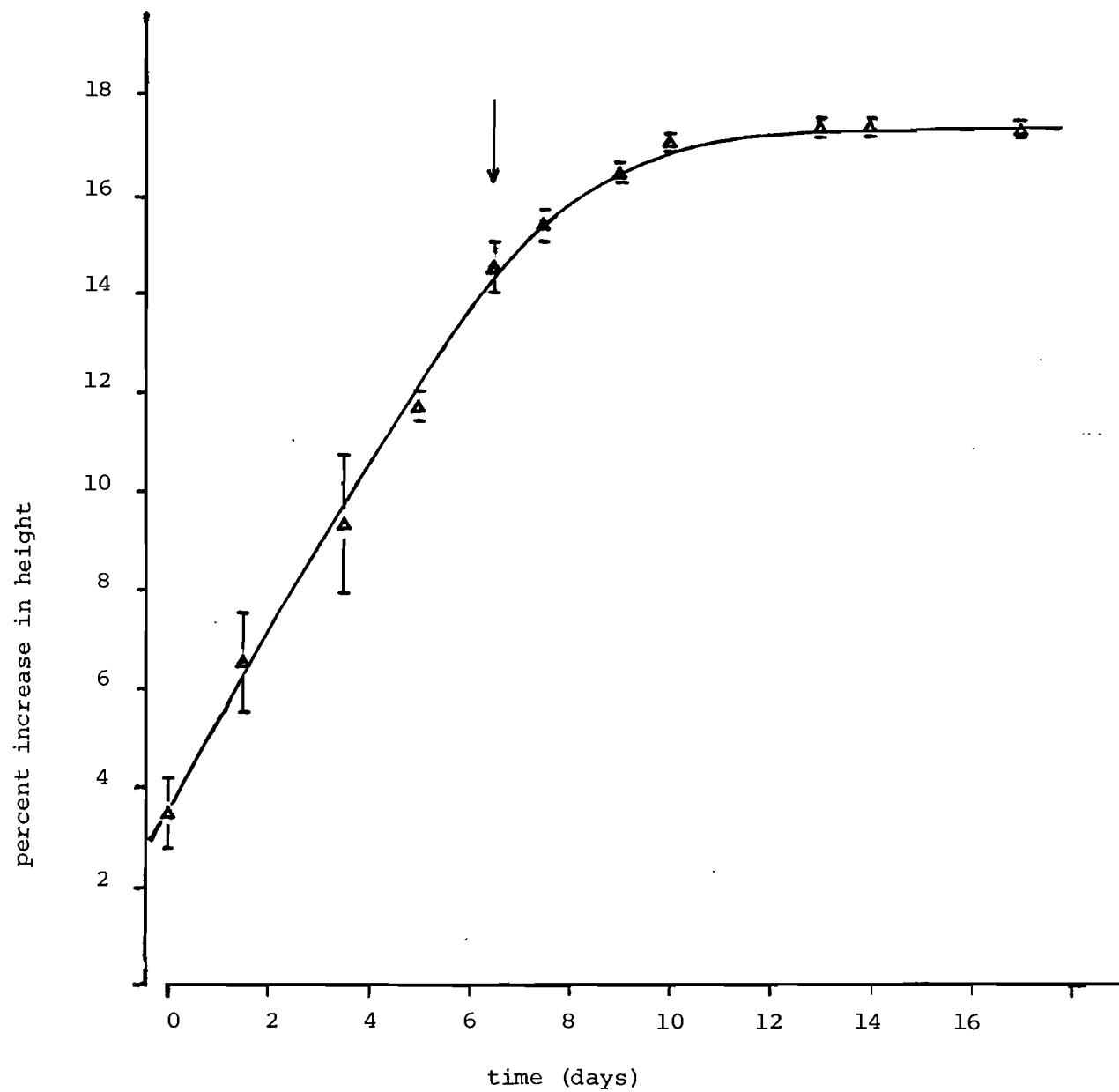


Figure 9. The response of wheat coleoptile sections to ABA.
Each value represents growth as a percentage of the control (growth in water) and is the mean \pm S.E. of three replicates, each consisting of 10 coleoptiles.

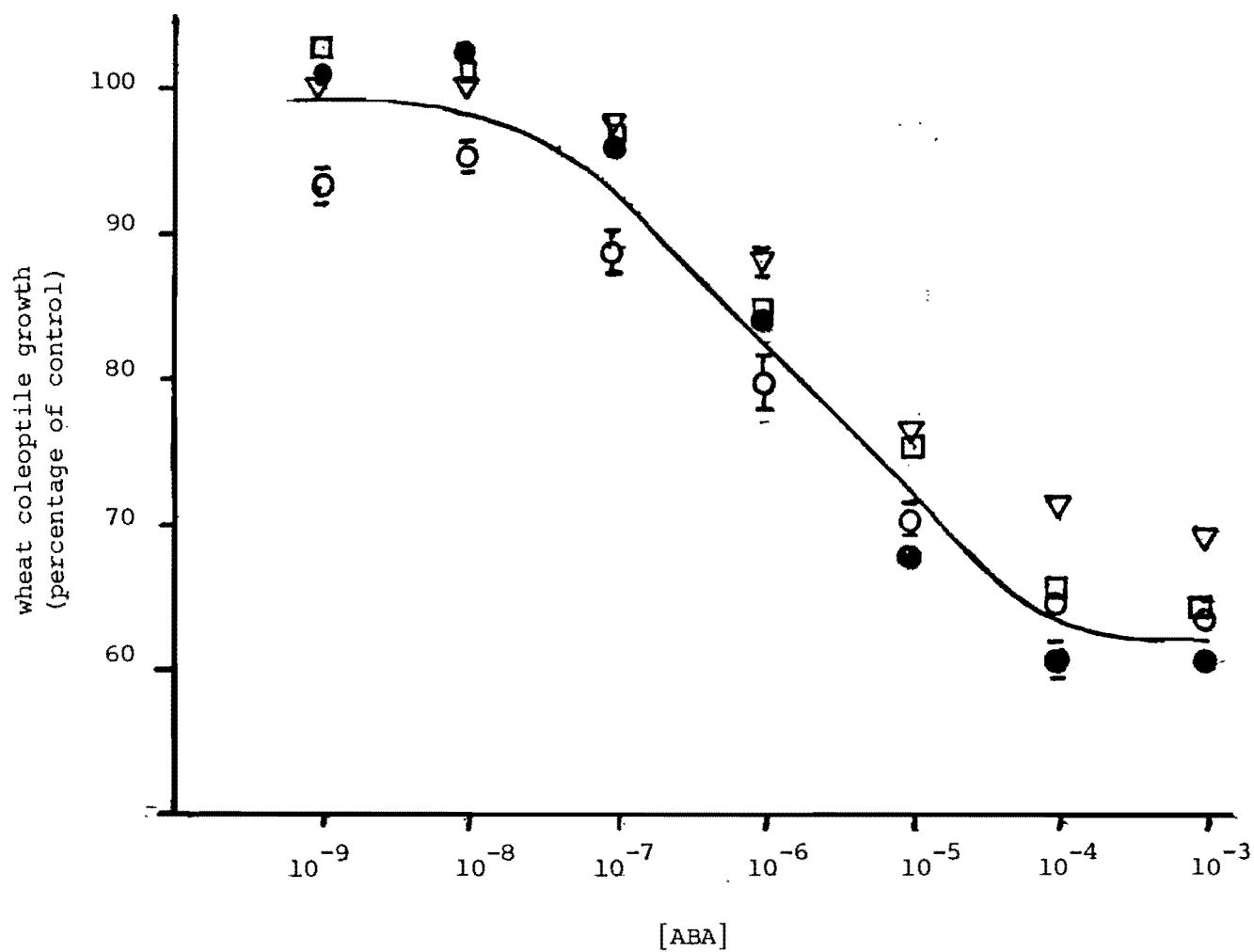


Figure 10. The effect of serial concentrations of ABA and short chain fatty acids (C5 and C10) on the extension growth of wheat coleoptile sections. Each value represents growth as a percentage of the control (growth in water) and is the mean \pm S.E. of 3 replicates, each consisting of 10 coleoptiles.

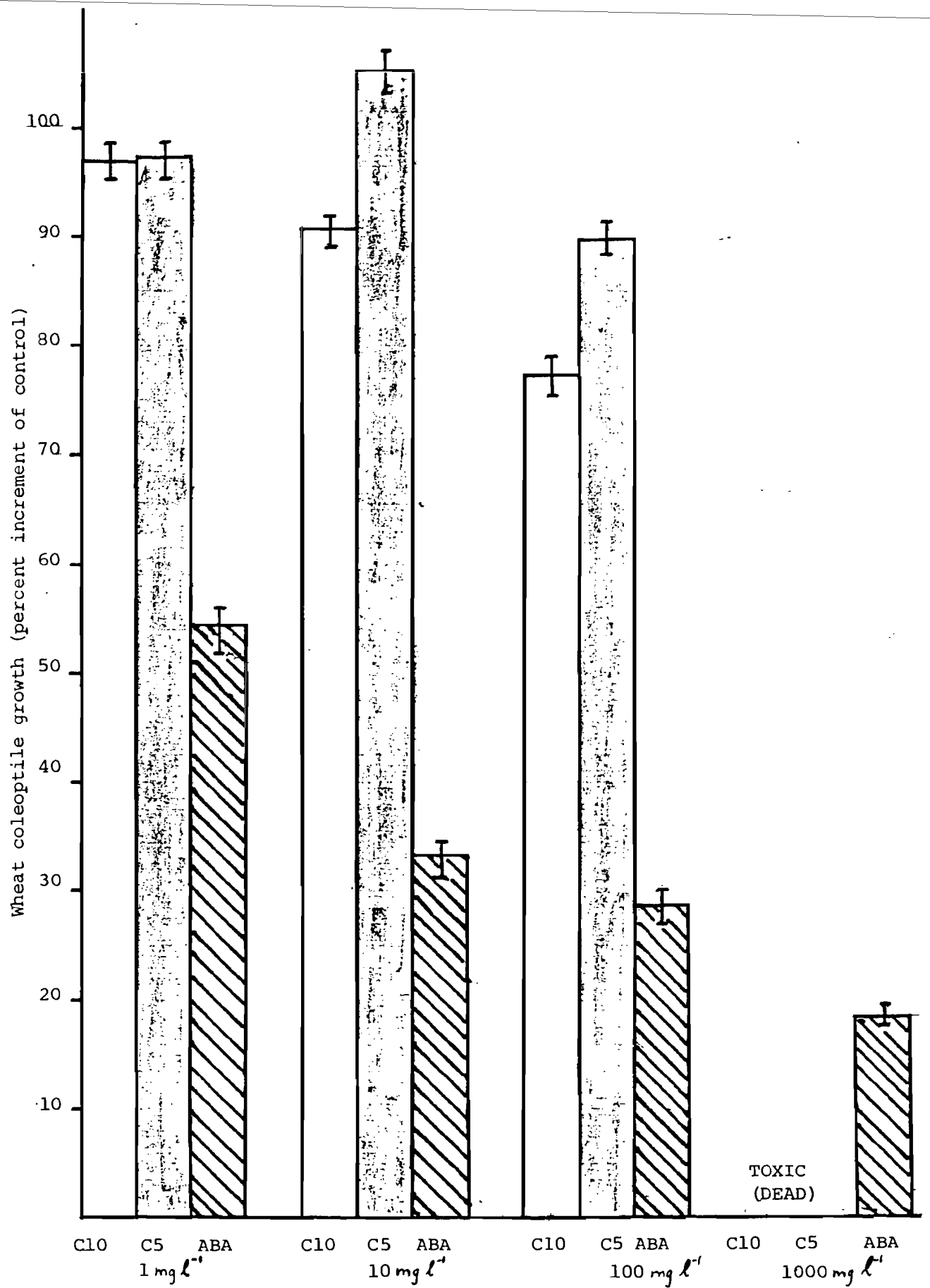


Figure 11. Wheat coleoptile section assay of unwashed chromatographs developed in isopropanol:NH₃:H₂O (10:1:1::v:v:v) solvent. Each R_F section was assayed using 10 coleoptiles. Vertical bars represent twice the standard error.

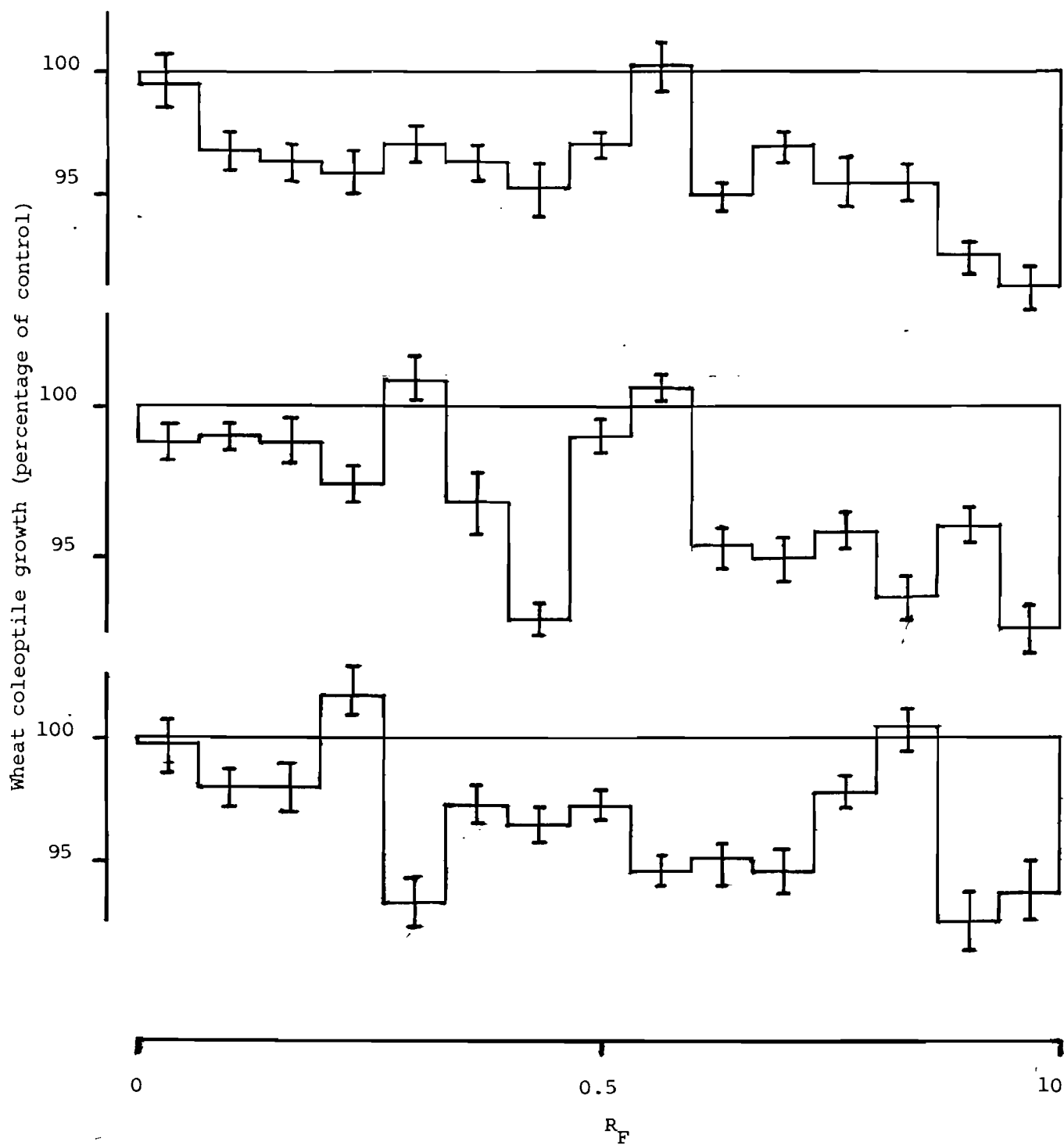


Figure 12. Wheat coleoptile section assay of blank, undeveloped chromatographs. Each R_F section was assayed using 10 coleoptiles. Vertical bars represent twice the standard error.

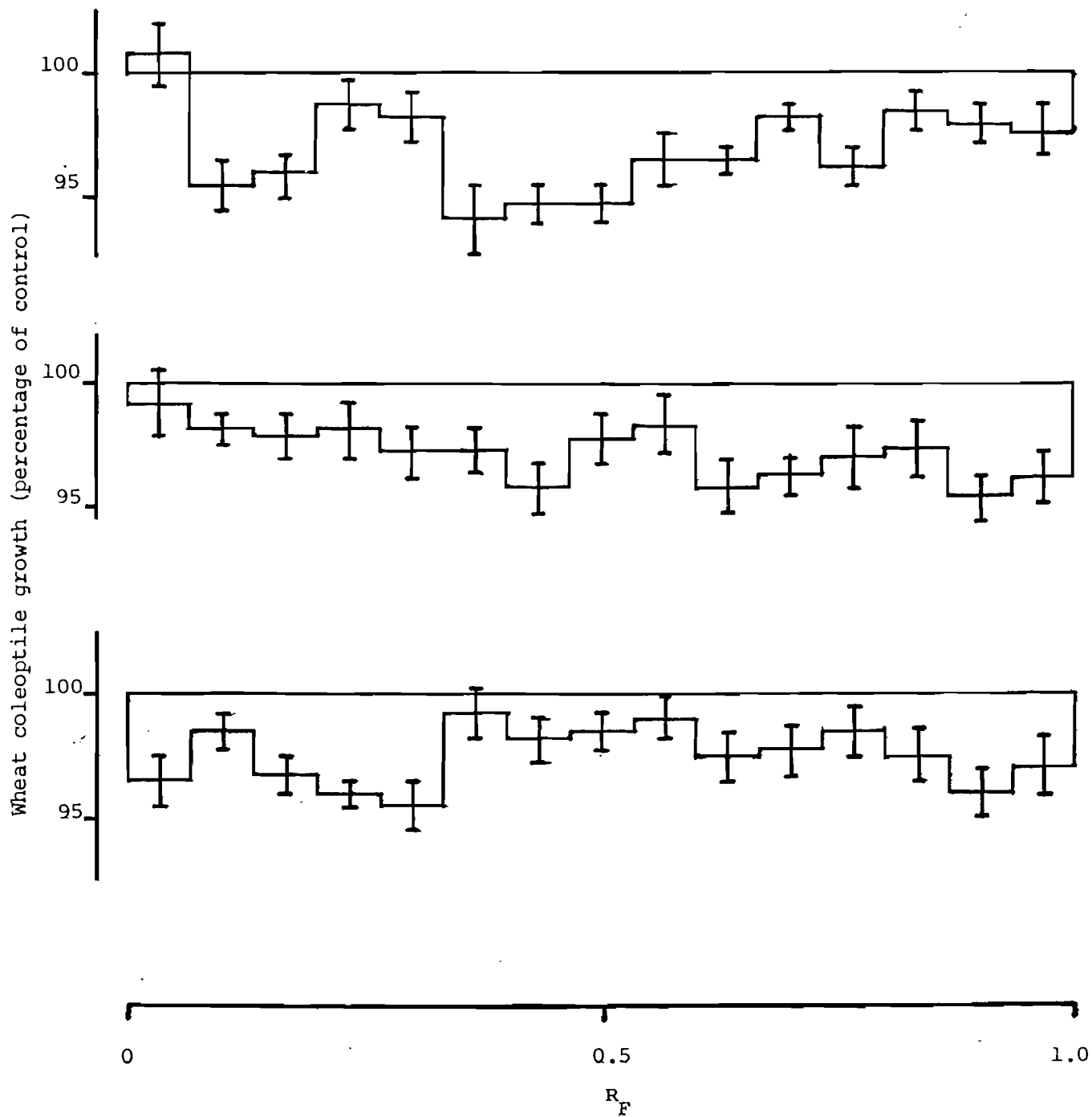


Figure 13. Wheat coleoptile section assay of pre-washed chromatographs developed in isopropanol:NH₃:H₂O (10:1:1::v:v:v) solvent. The chromatographs were pre-washed three times in 80% MeOH. Each R_F section was assayed using 10 coleoptiles. Vertical bars represent twice the standard error.

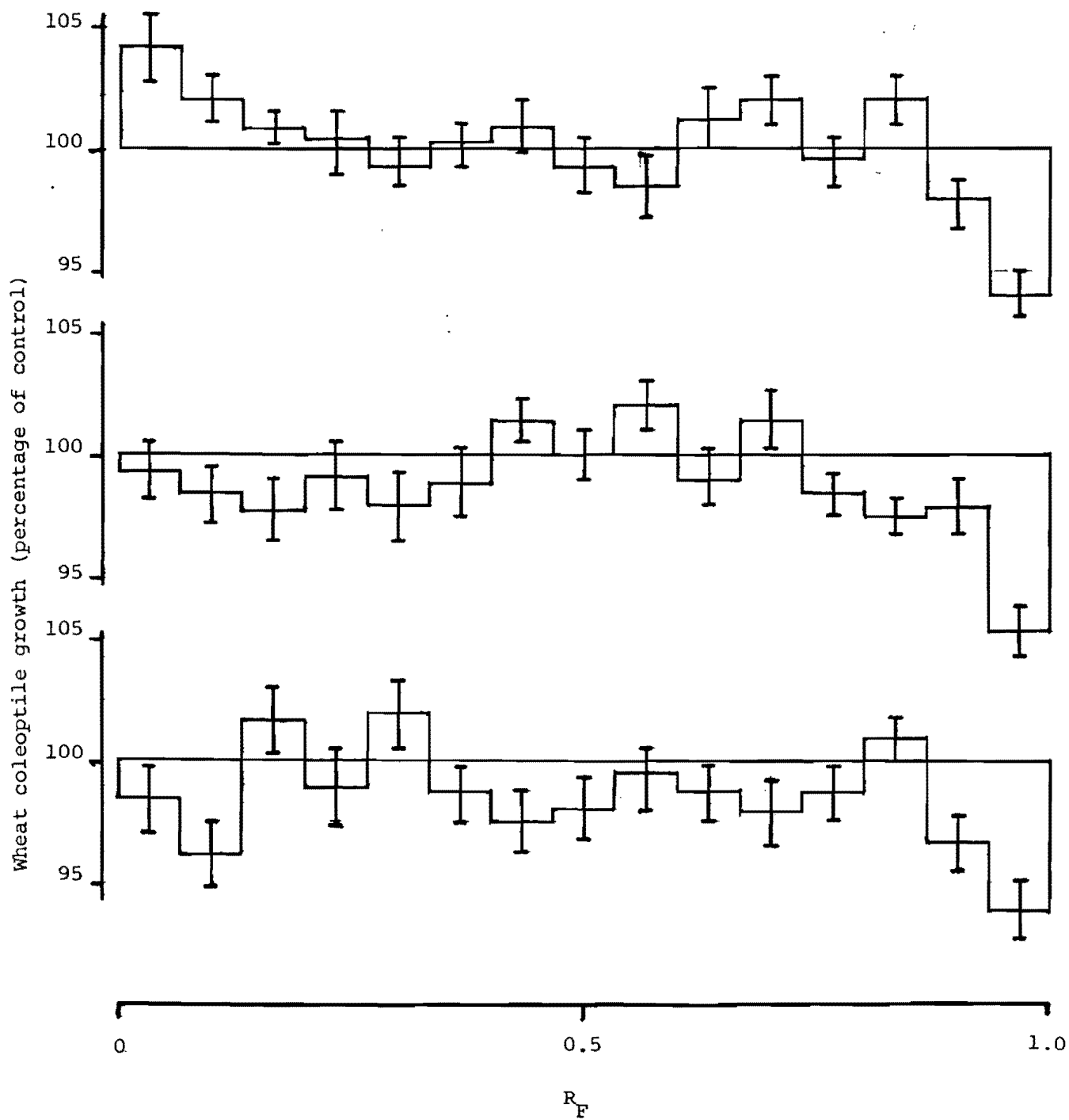


Figure 14. The response of lettuce hypocotyls to ABA. Each value represents growth as a percentage of the control (growth in water) and is the mean \pm S.E. of three replicates, each consisting of 16 hypocotyls.

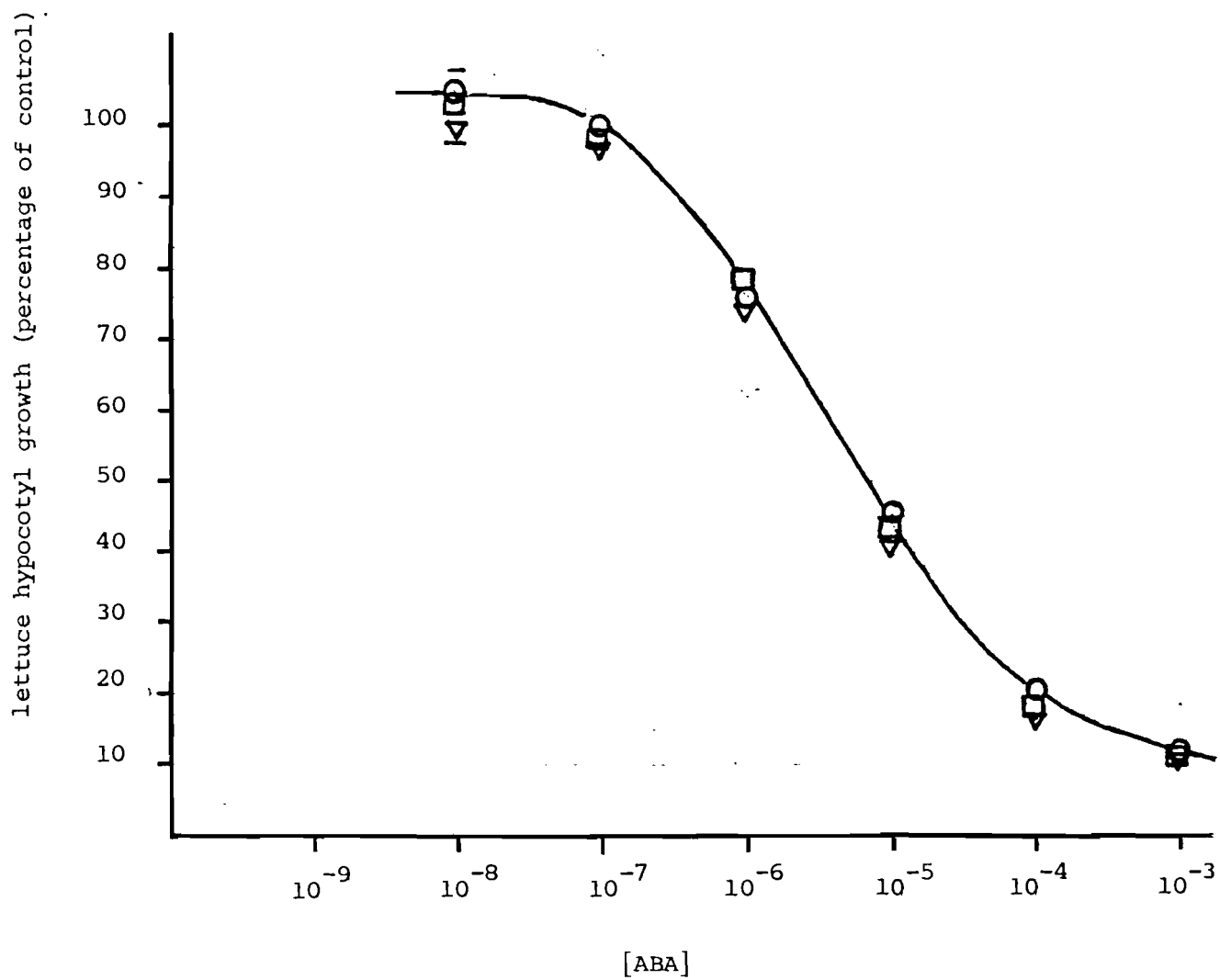


Figure 15. The effect of serial concentrations of ABA, C5, C10 and IAA on the growth of lettuce hypocotyls. Each value represents growth as a percentage of the control (growth in water) and is the mean \pm S.E. of three replicates, each consisting of 16 hypocotyls.

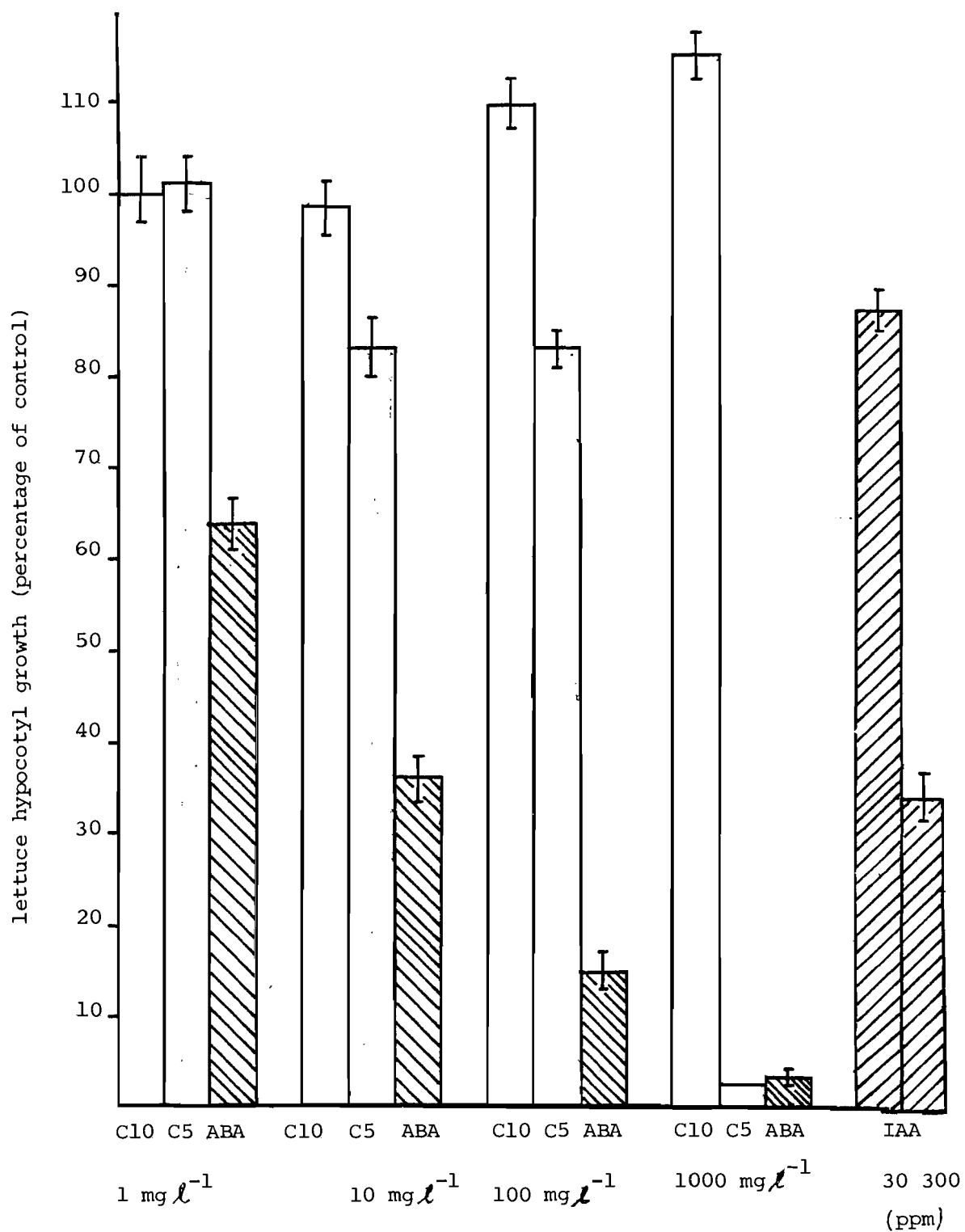


Figure 16. Lettuce hypocotyl assay of pre-washed chromatographs developed in isopropanol:NH₃:H₂O (10:1:1::v:v:v) solvent. The chromatographs were pre-washed three times in 80% MeOH. Each R_F section was assayed using 16 hypocotyls and the vertical bars represent twice the standard error.

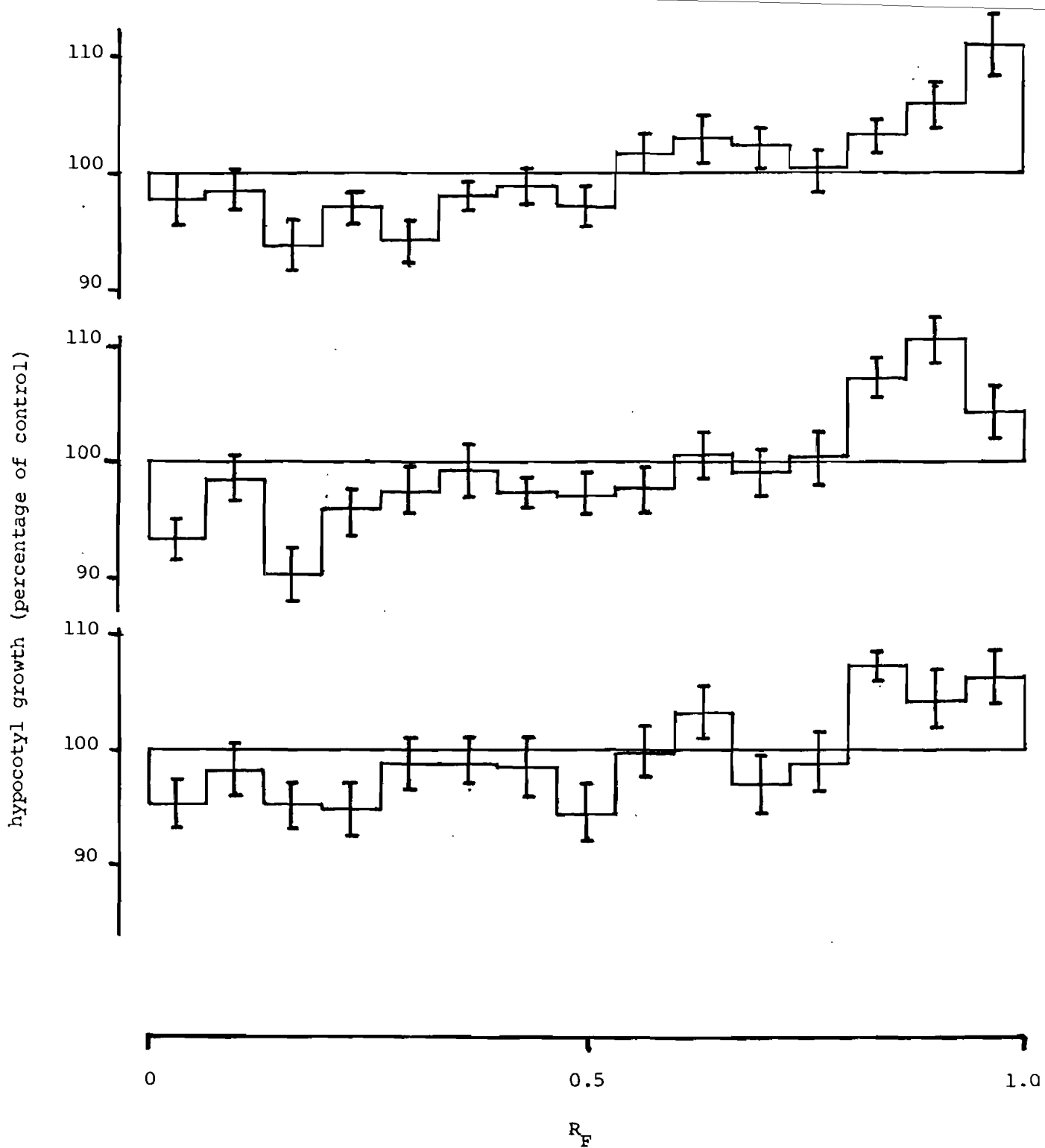


Figure 17. Diagram of a typical chromatograph of an acidic ether-soluble extract of *Alnus* tissue as seen under (A) visible and (B) U.V. light. The chromatographs were loaded with the equivalent of approx. 0.2 g DW of tissue and developed in isopropanol:NH₃:H₂O (10:1:1::v:v:v).

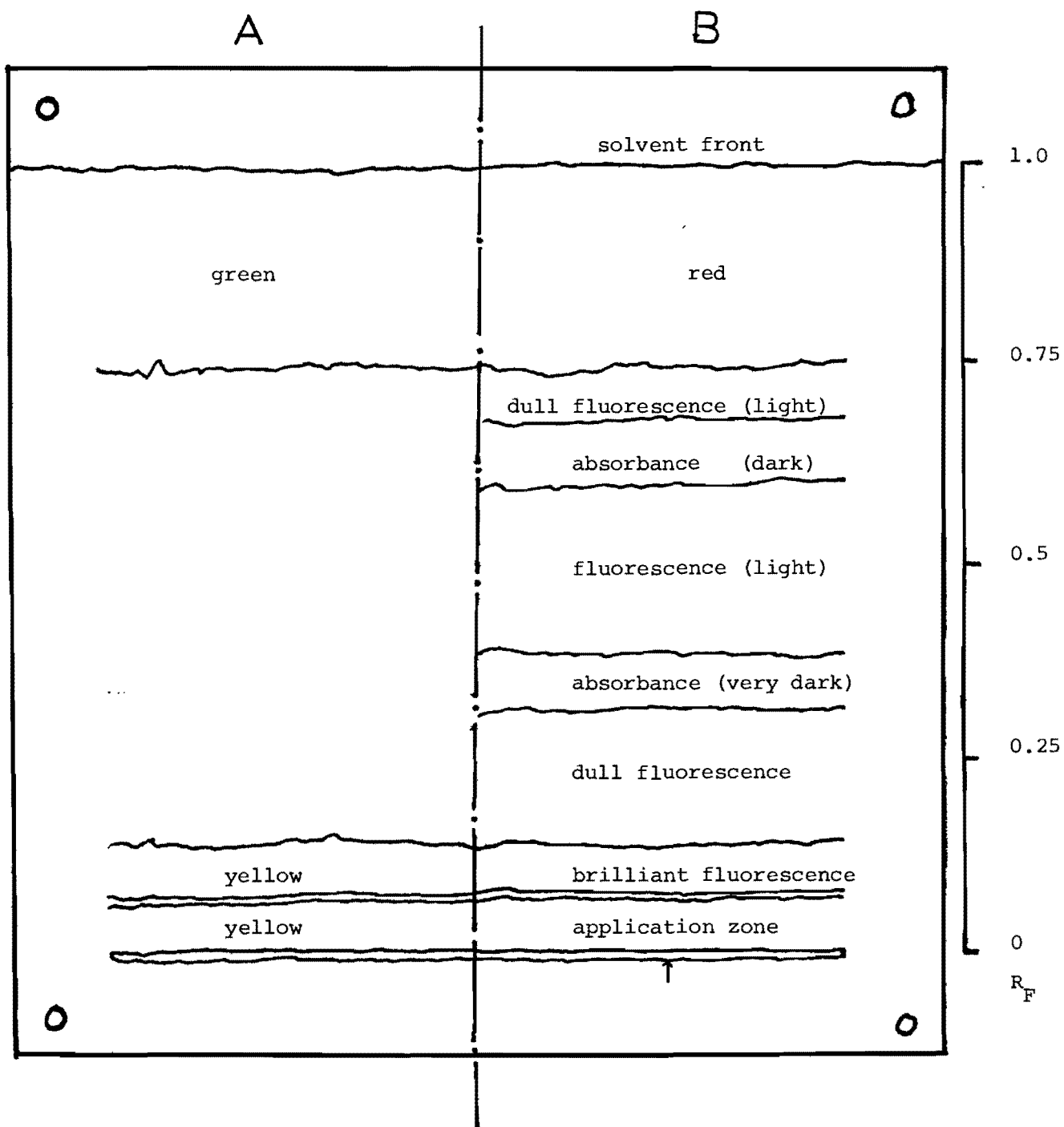


Figure 18. Wheat coleoptile section assay of acidic ether-soluble extracts obtained from leaves of *Alnus glutinosa* trees harvested on 7 March 1977 (H1). The equivalent of 0.25 g DW of leaf material was assayed and the solvent system was isopropanol:NH₃:H₂O (10:1:1::v:v:v). Each R_F section was assayed on 10 coleoptiles and each histogram represents the mean \pm S.E. of three replicate chromatographs. The broken line represents the assay of a control (pre-washed, solvent developed) chromatograph, and the R_F of authentic ABA is indicated by the horizontal bar under some of the histograms.

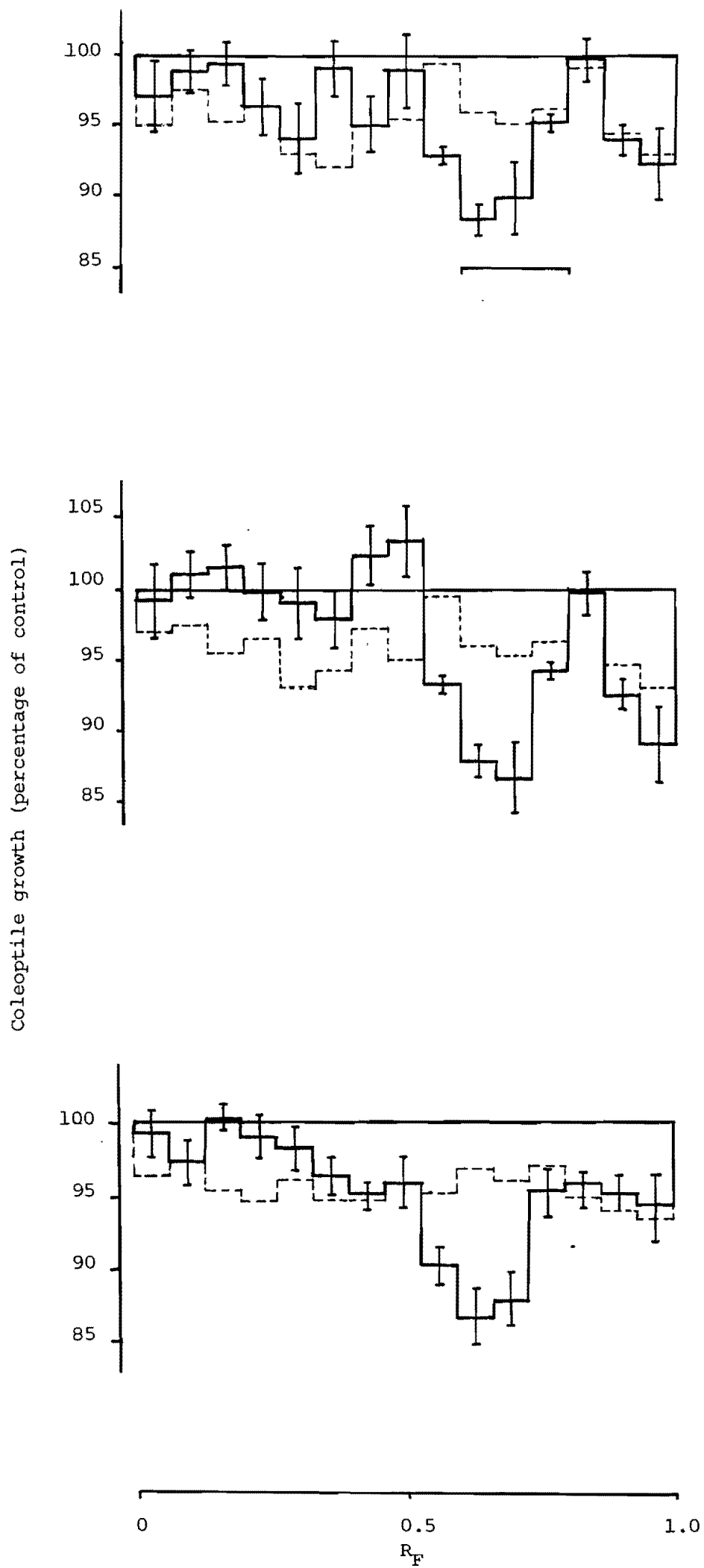


Figure 19. Wheat coleoptile section assay of acidic ether-soluble extracts obtained from leaves of *Alnus glutinosa* trees harvested on 25 March 1977 (H2). The equivalent of 0.25 g DW of leaf material was assayed and the solvent system was isopropanol:NH₃:H₂O (10:1:1::v:v:v). Other details as in figure 18.

coleoptile growth (percentage of control)

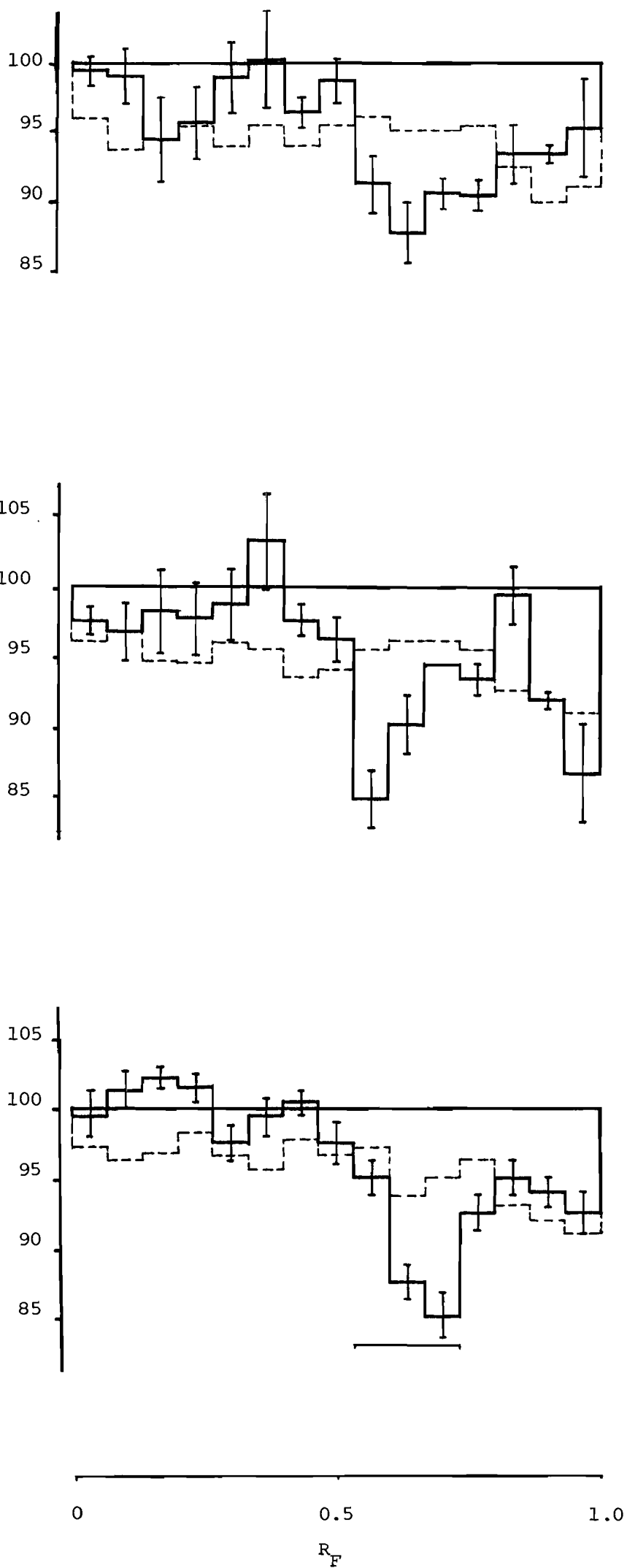
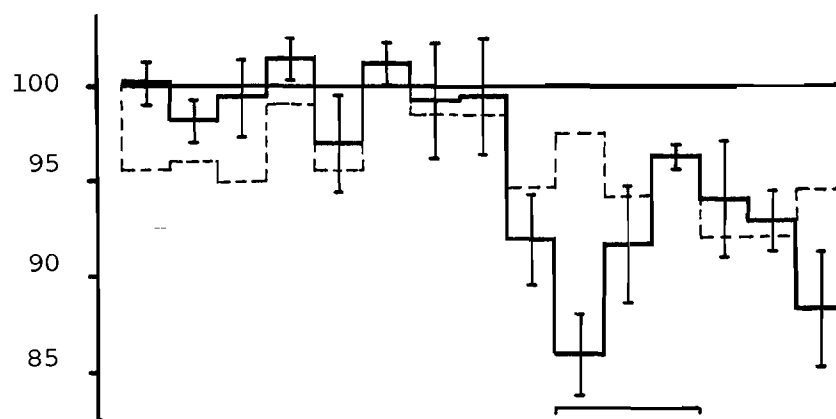


Figure 20. Wheat coleoptile section assay of acidic ether-soluble extracts obtained from leaves of *Alnus glutinosa* trees harvested on 15 April 1977 (H3). The equivalent of 0.25 g DW of leaf material was assayed and the solvent system was isopropanol:NH₄OH:H₂O (10:1:1::v:v:v). Other details as in figure 18.



Coleoptile growth (percentage of control)

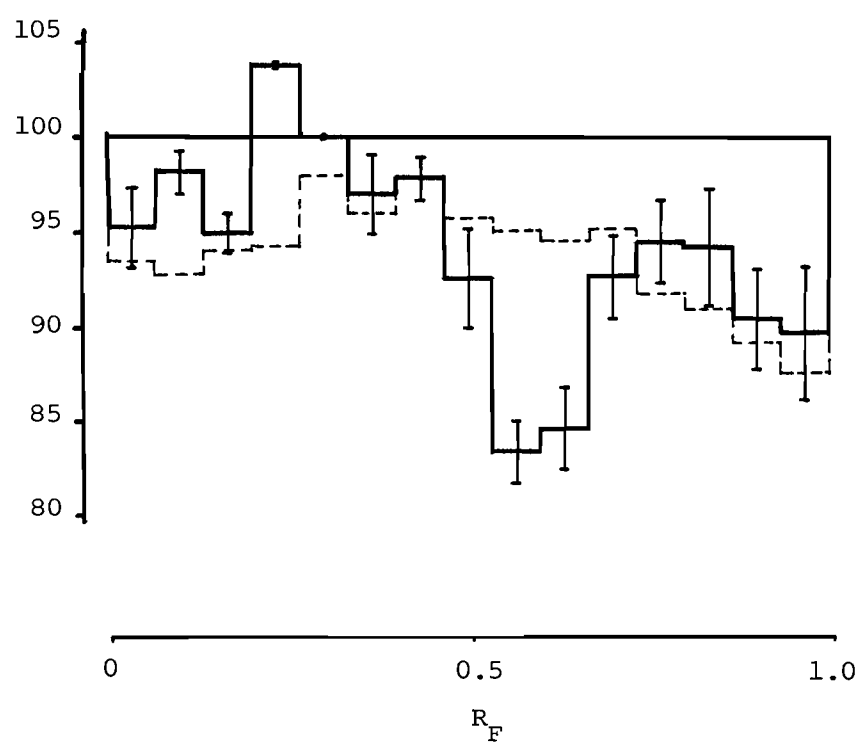
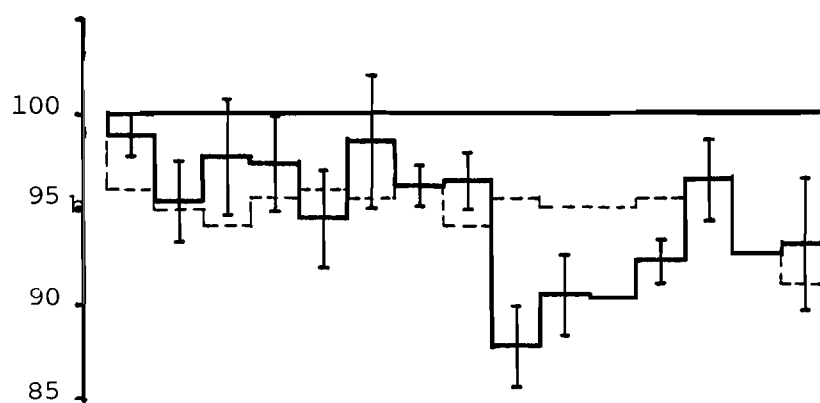


Figure 21. Wheat coleoptile section assay of acidic ether-soluble extracts obtained from leaves of *Alnus viridis* trees harvested on 28 April 1977 (H4). The equivalent of 0.25 g DW of leaf material was assayed and the solvent system was isopropanol:NH₃:H₂O (10:1:1::v:v:v). Other details as in figure 18.

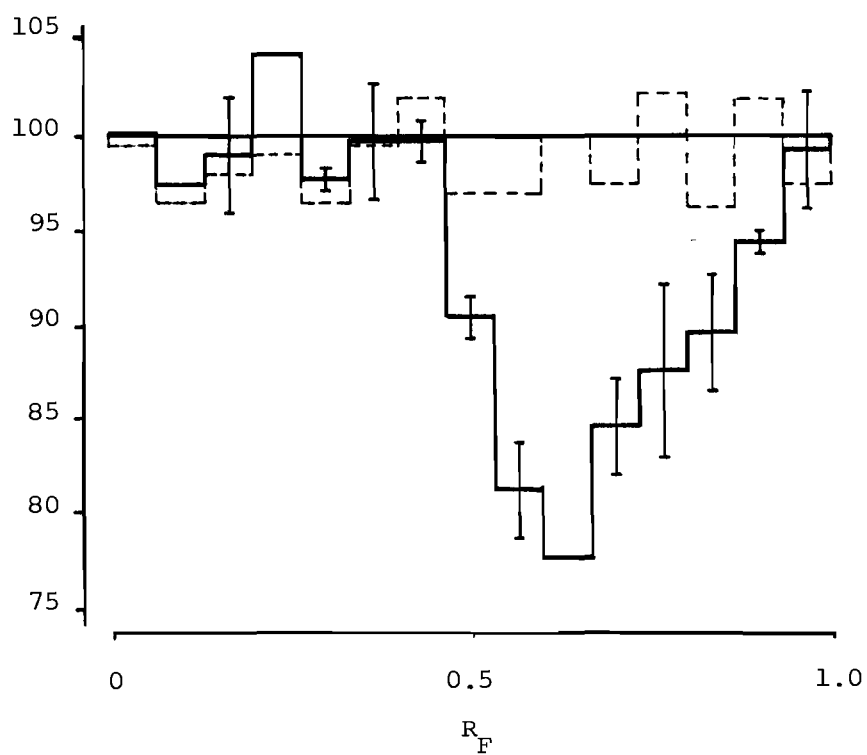
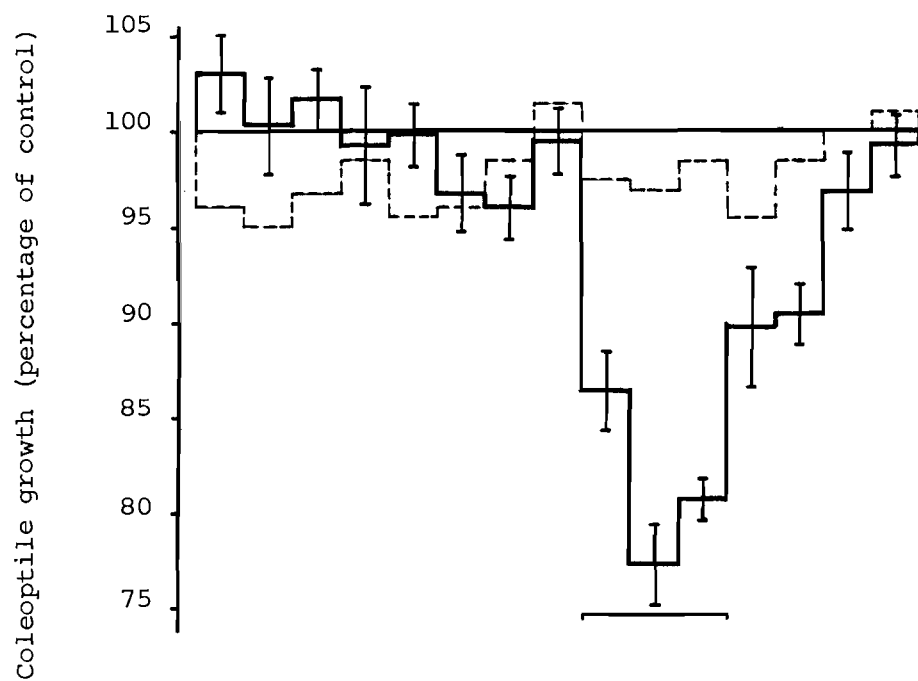
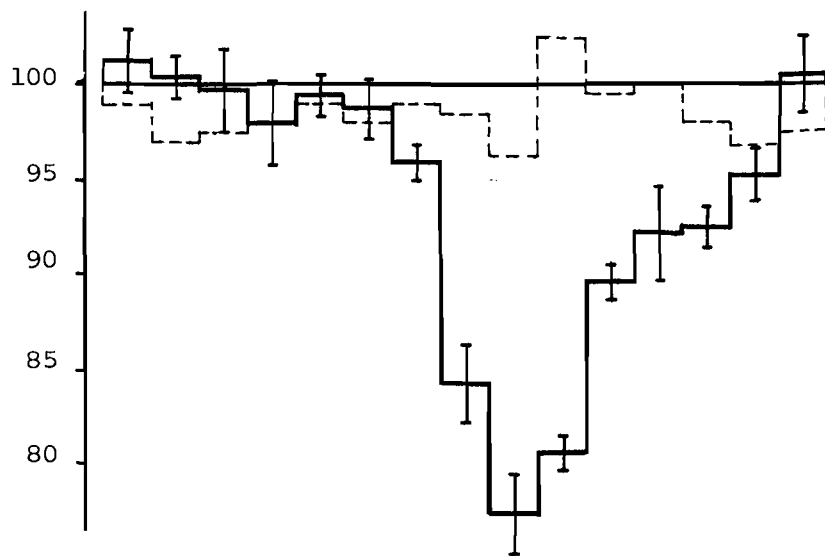


Figure 22. Wheat coleoptile section assay of acidic ether-soluble extracts obtained from leaves of *Alnus glutinosa* trees harvested on 15 May 1977 (H5). The equivalent of 0.25 g DW of leaf material was assayed and the solvent system was isopropanol:NH₃:H₂O (10:1:1::v:v:v). Other details as in figure 18.

Coleoptile growth (percentage of control)

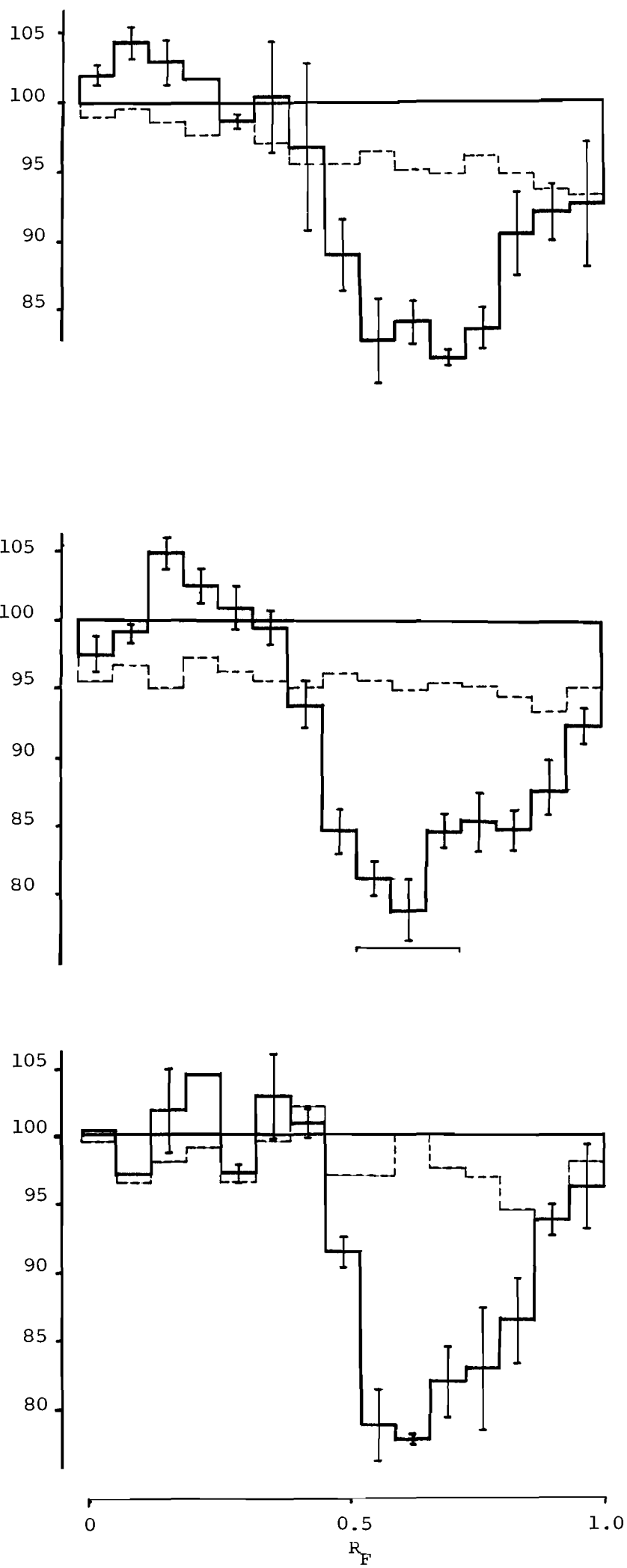


Figure 23. Wheat coleoptile section assay of authentic ABA standards. Aliquots equivalent to (a) 0.0512, (b) 0.512, (c) 5.12 and (d) 51.2 μg of the mixed isomers of authentic ABA were loaded on to the chromatographs. The solvent system was isopropanol: NH_3 : H_2O (10:1:1::v:v:v). Other details as in figure 18.

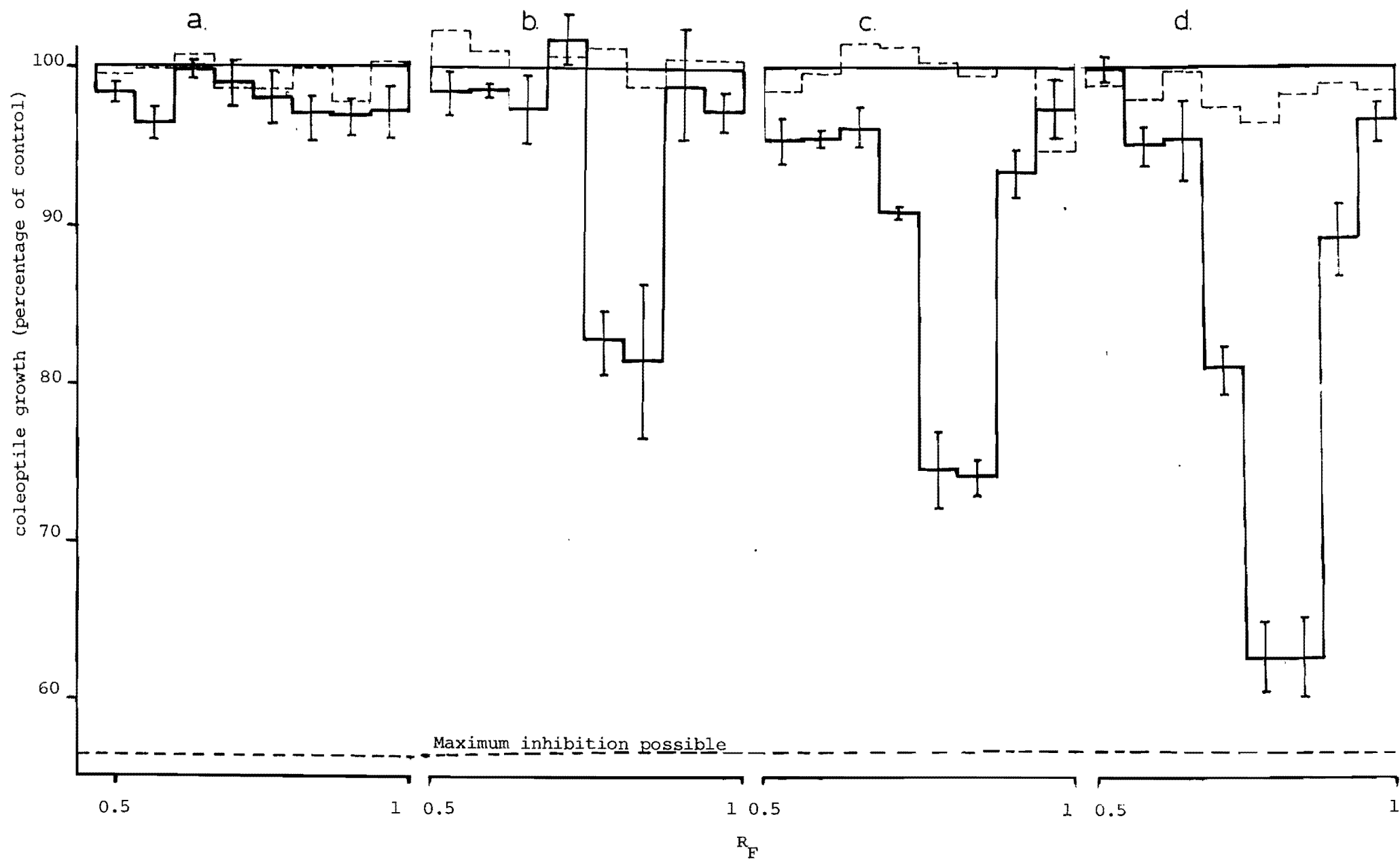


Figure 24. Wheat coleoptile section assay of the serial dilution of an acidic ether-soluble extract obtained from leaves of *Alnus glutinosa* trees harvested on 7 March 1977 (H1). The equivalent of (a) 0.025, (b) 0.125, (c) 0.25, and (d) 0.375 g DW of leaf material was assayed. Other details as in figure 18.

Coleoptile growth (percentage of control)

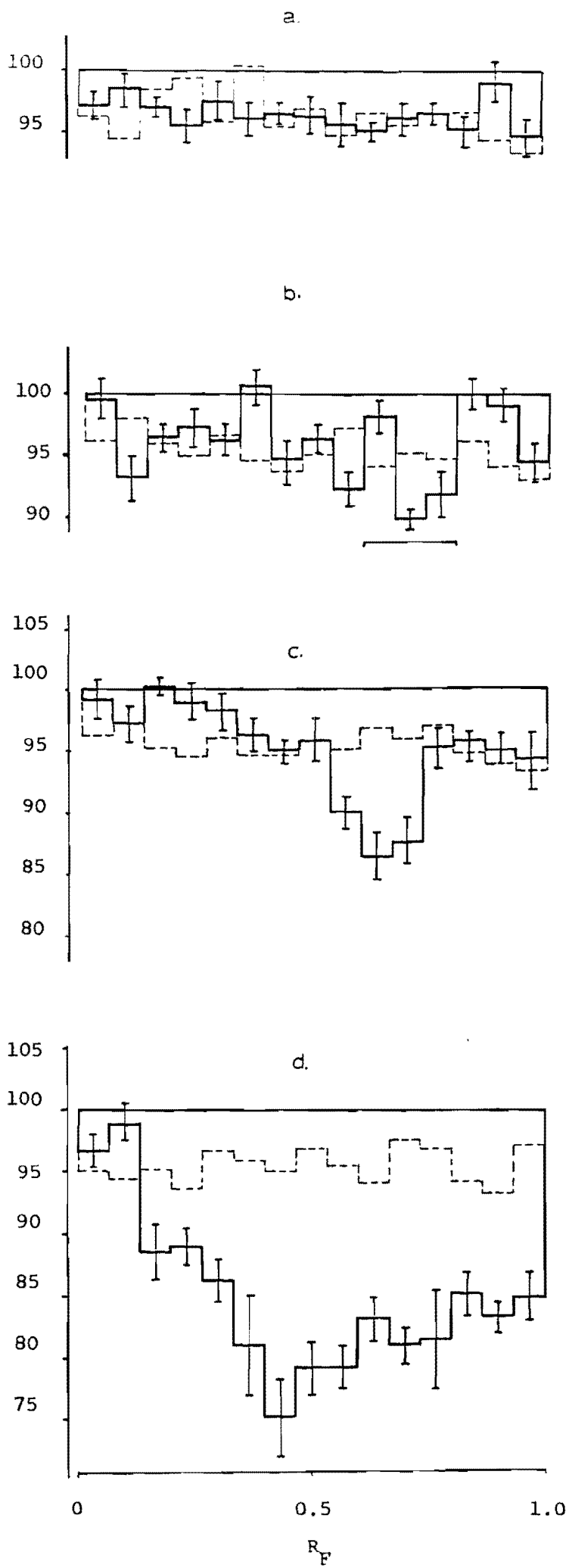


Figure 25. Wheat coleoptile section assay of the serial dilution of an acidic ether-soluble extract obtained from leaves of *Alnus glutinosa* trees harvested on 15 May 1977 (H5). The equivalent of (a) 0.025, (b) 0.125, (c) 0.25 and (d) 0.375 g DW of leaf material was assayed. Other details as in figure 18.

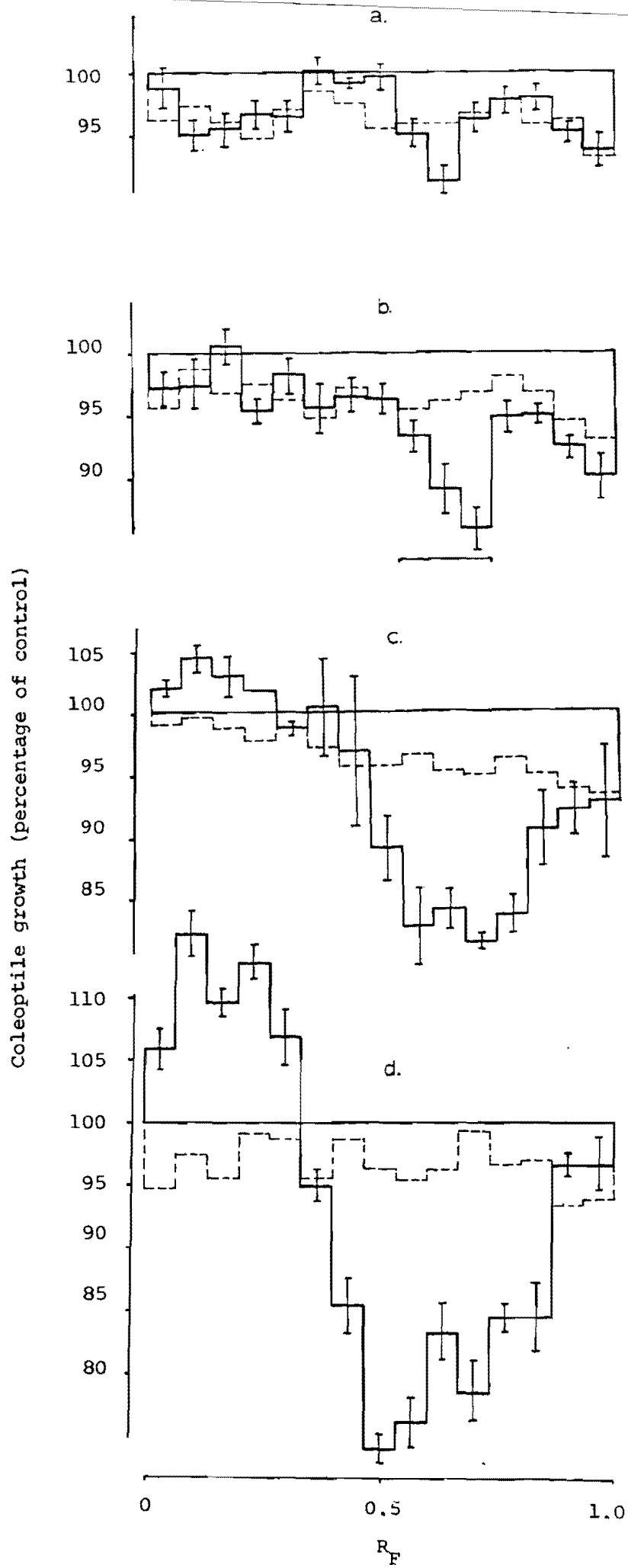


Figure 26. Lettuce hypocotyl assay of acidic ether-soluble extracts obtained from leaves of *Alnus glutinosa* trees harvested on 7 March 1977 (H1). The equivalent of 0.25 g DW of leaf material was assayed and the solvent system was isopropanol: NH_3 : H_2O (10:1:1::v:v:v). Each R_F section was assayed on 10 coleoptiles and each histogram represents the mean \pm S.E. of three replicate chromatographs. The broken line represents the assay of a control (pre-washed solvent developed) chromatograph and the R_F authentic ABA is indicated by the horizontal bar under some of the histograms.

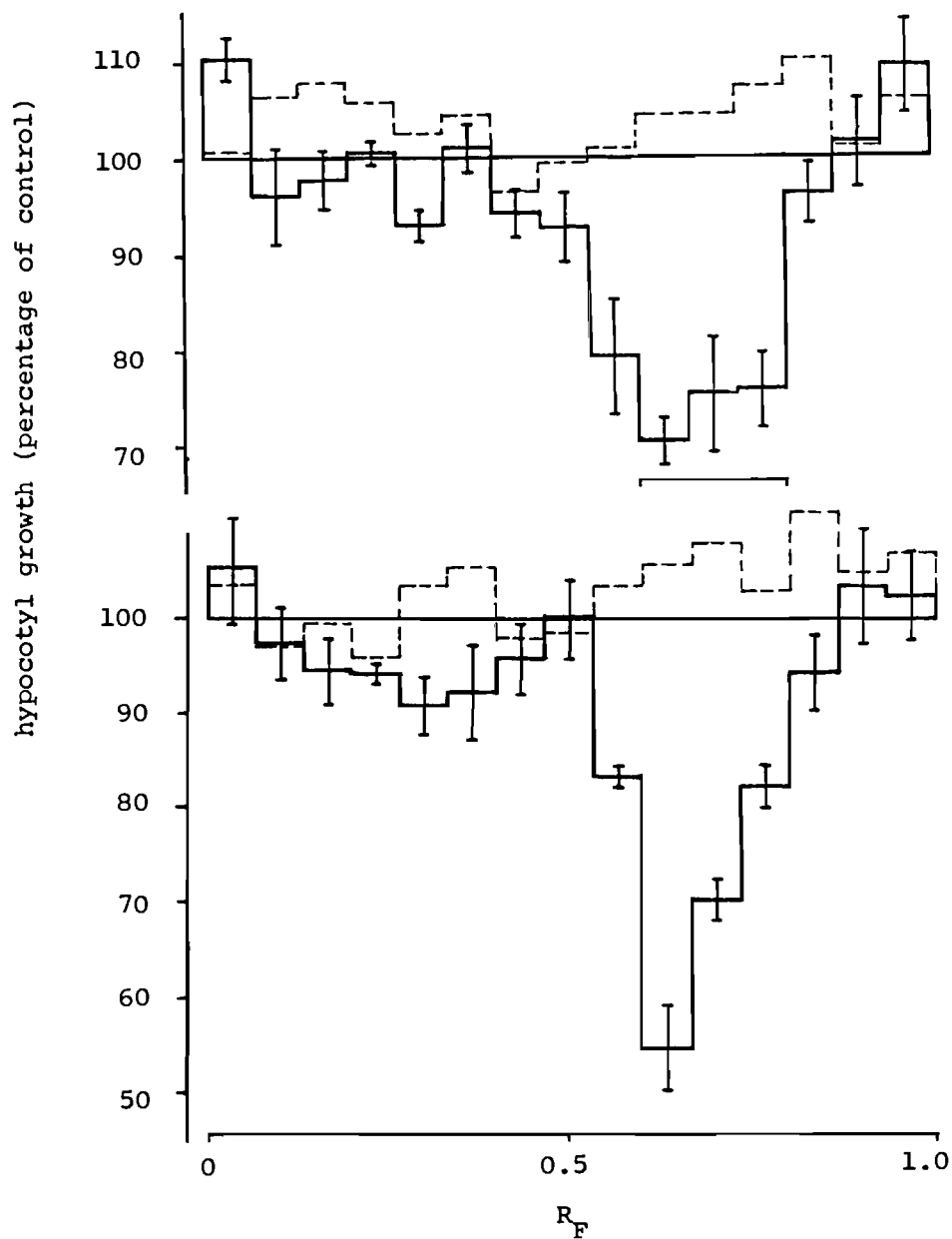
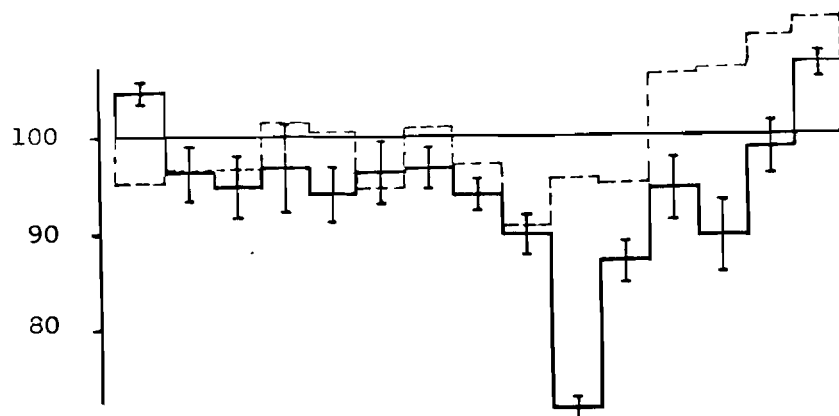


Figure 27. Lettuce hypocotyl assay of acidic ether-soluble extracts obtained from leaves of *Alnus glutinosa* trees harvested on 25 March 1977 (H2). The equivalent of 0.25 g DW of leaf material was assayed and the solvent system was isopropanol:NH₃:H₂O (10:1:1::v:v:v). Other details as in figure 26.

hypocotyl growth (percentage of control)

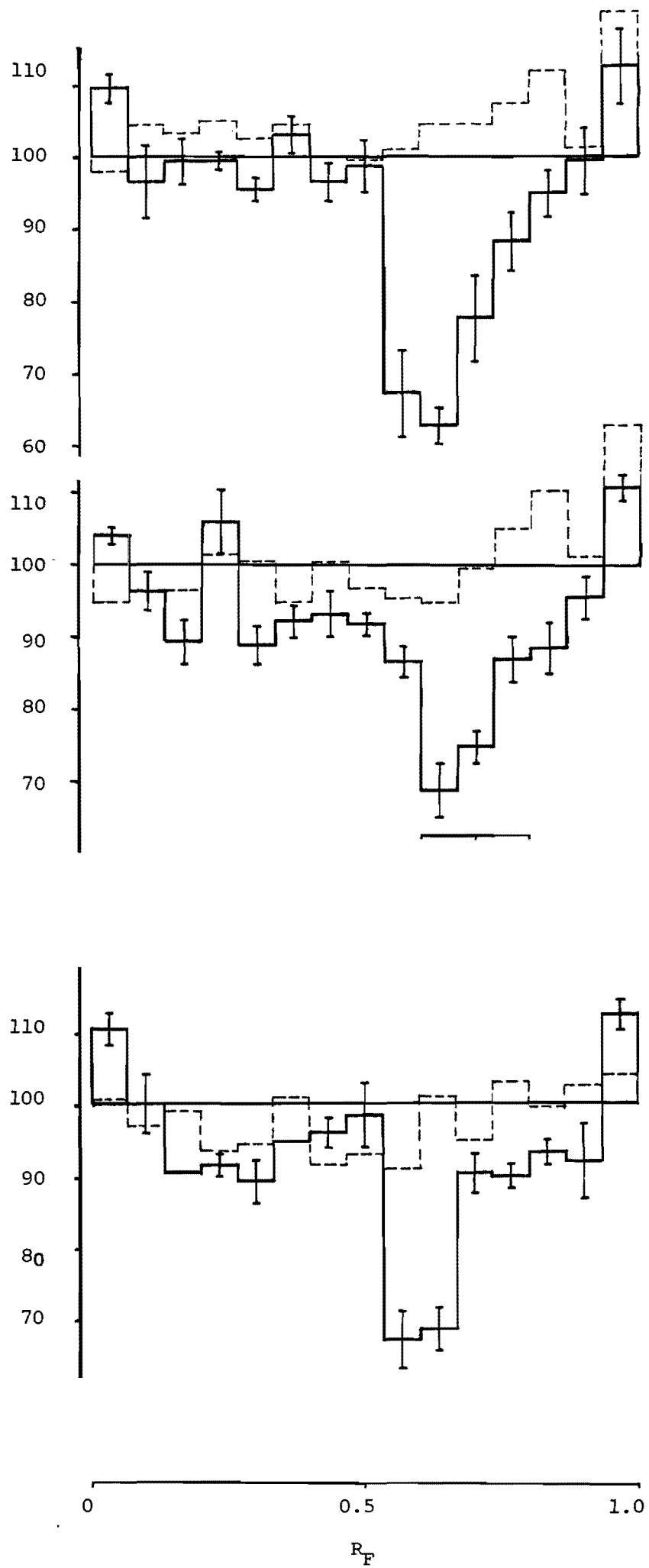


Figure 28. Lettuce hypocotyl assay of acidic ether-soluble extracts obtained from leaves of *Alnus glutinosa* trees harvested on 15 April 1977 (H3). The equivalent of 0.25 g DW of leaf material was assayed and the solvent system was isopropanol:NH₃:H₂O (10:1:1::v:v:v). Other details as in figure 26.

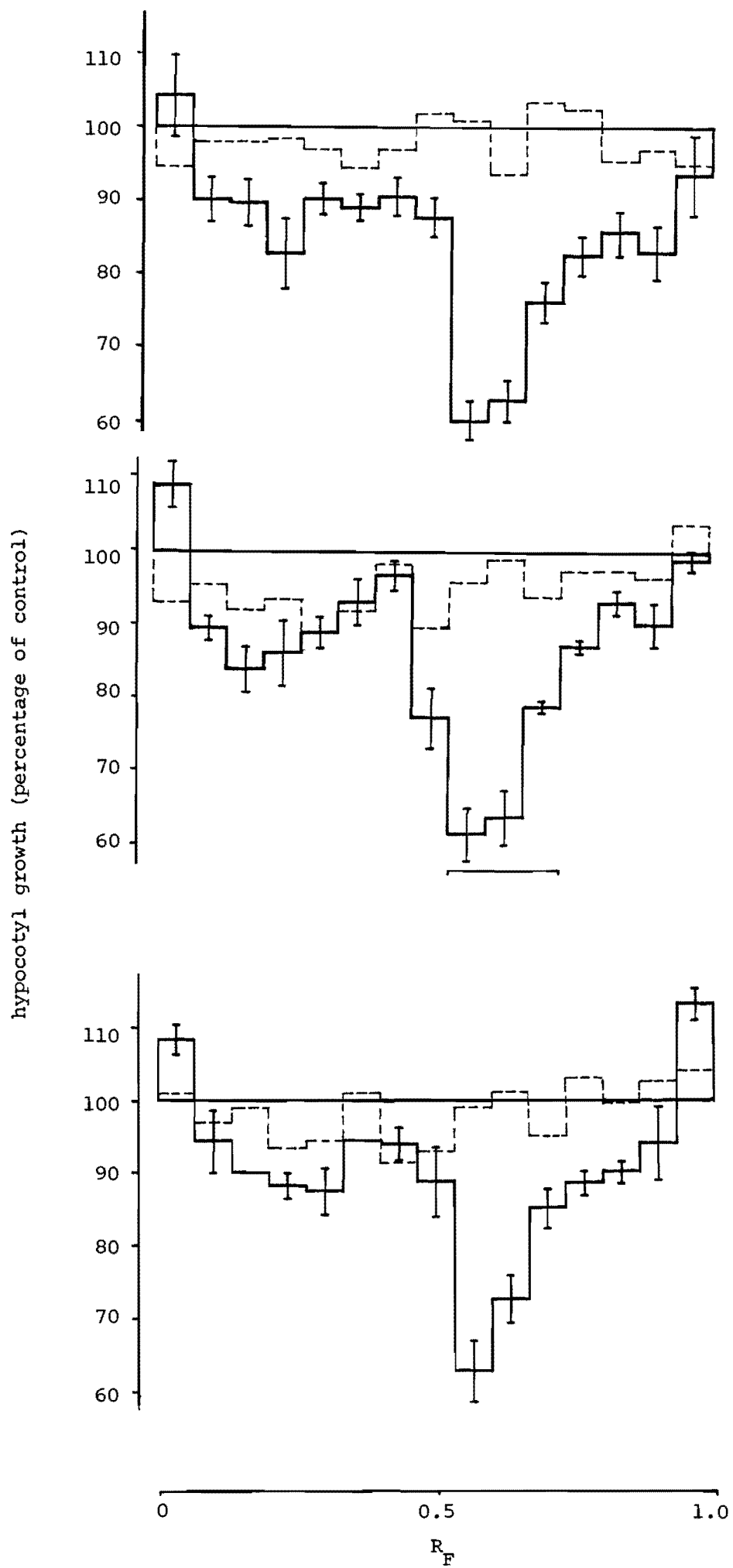


Figure 29. Lettuce hypocotyl assay of acidic ether-soluble extracts obtained from leaves of *Alnus glutinosa* trees harvested on 28 April 1977 (H4). The equivalent of 0.25 g DW of leaf material was assayed and the solvent system was isopropanol:NH₃:H₂O (10:1:1::v:v:v). Other details as in figure 26.

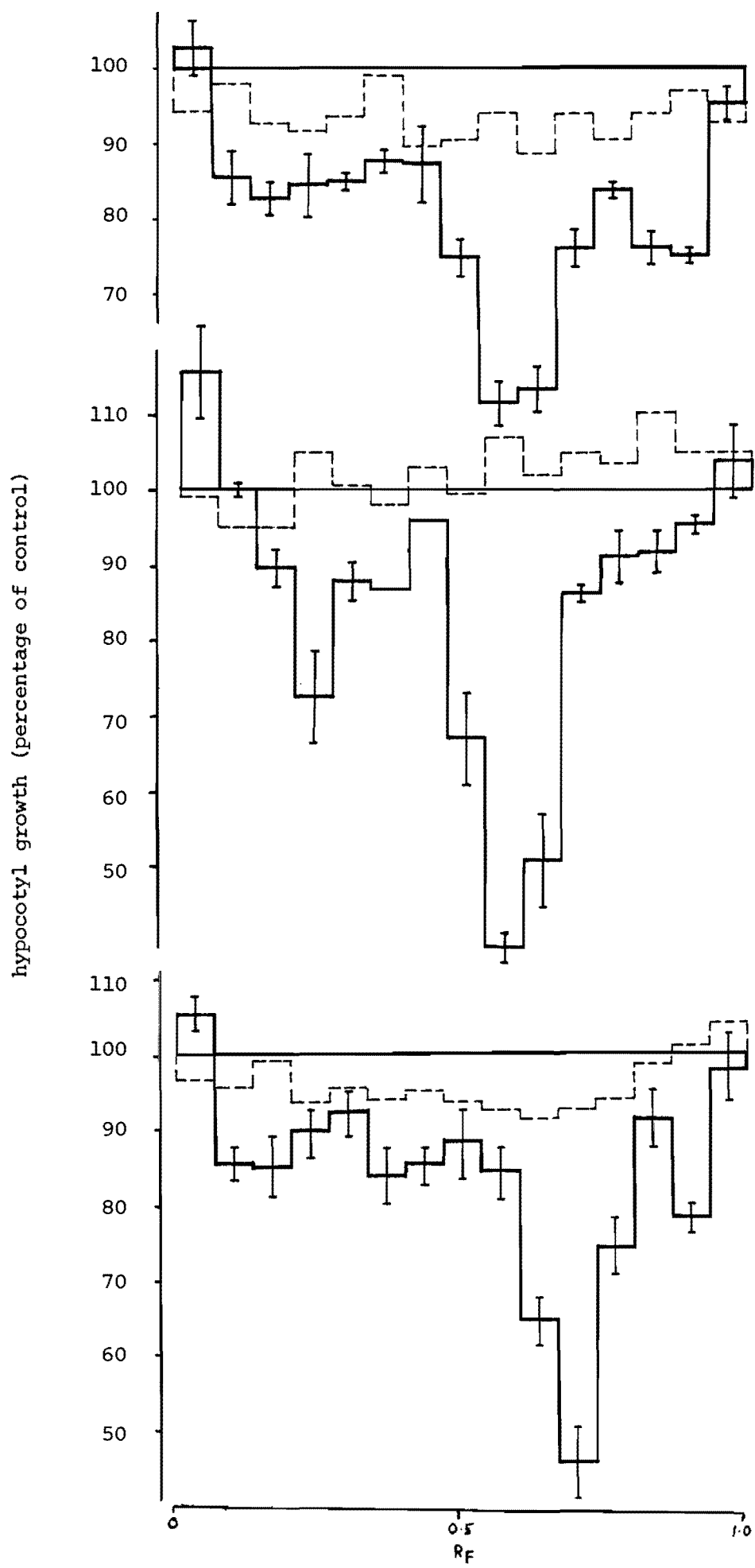


Figure 30. Lettuce hypocotyl assay of acidic ether-soluble extracts obtained from leaves of *Alnus glutinosa* trees harvested on 15 May 1977 (H5). The equivalent of 0.25 g DW of leaf material was assayed and the solvent system was isopropanol:NH₃:H₂O (10:1:1::v:v:v). Other details as in figure 26.

hypocotyl growth (percentage of control)

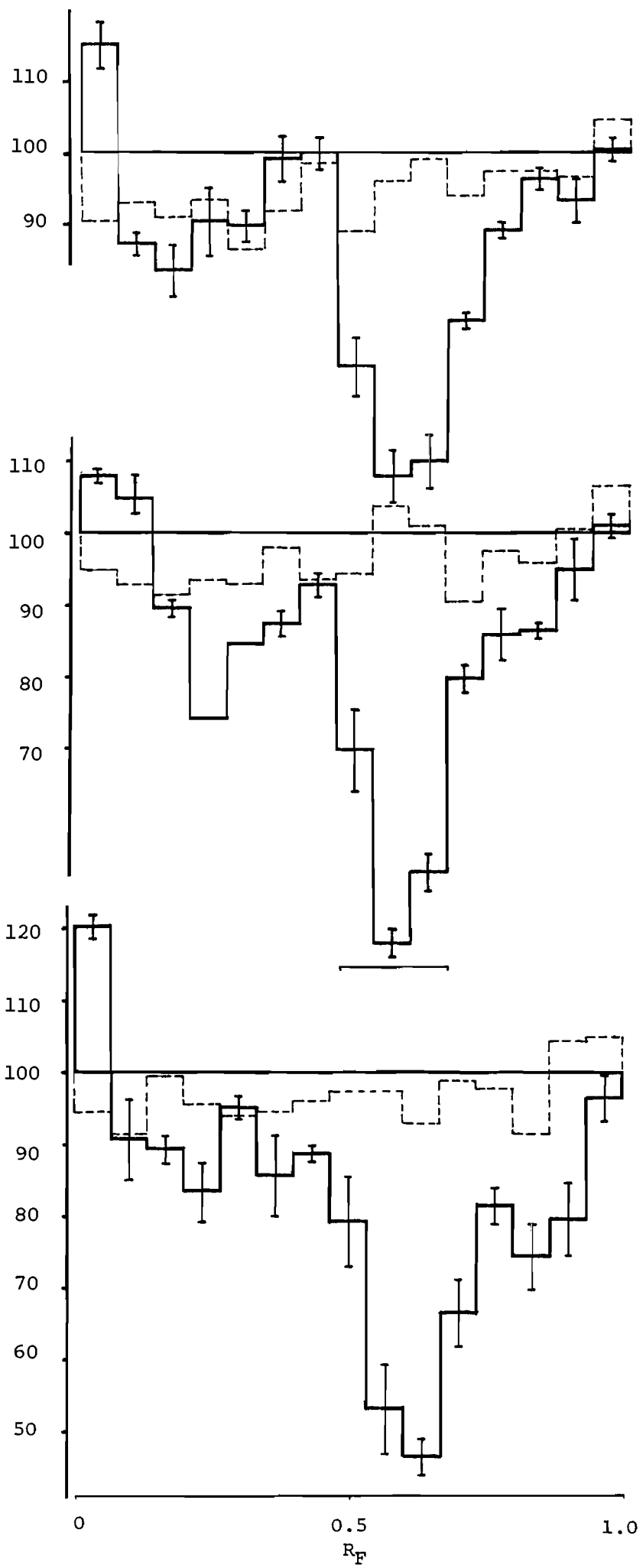


Figure 31. Lettuce hypocotyl assay of authentic ABA standards. Aliquots equivalent to (a) 0.0512, (b) 0.512, (c) 5.12) and (d) 51.2 μg of the mixed isomers of authentic ABA were loaded on to the chromatographs. The solvent system was isopropanol: NH_3 : H_2O (10:1:1::v:v:v). Other details as in figure 26.

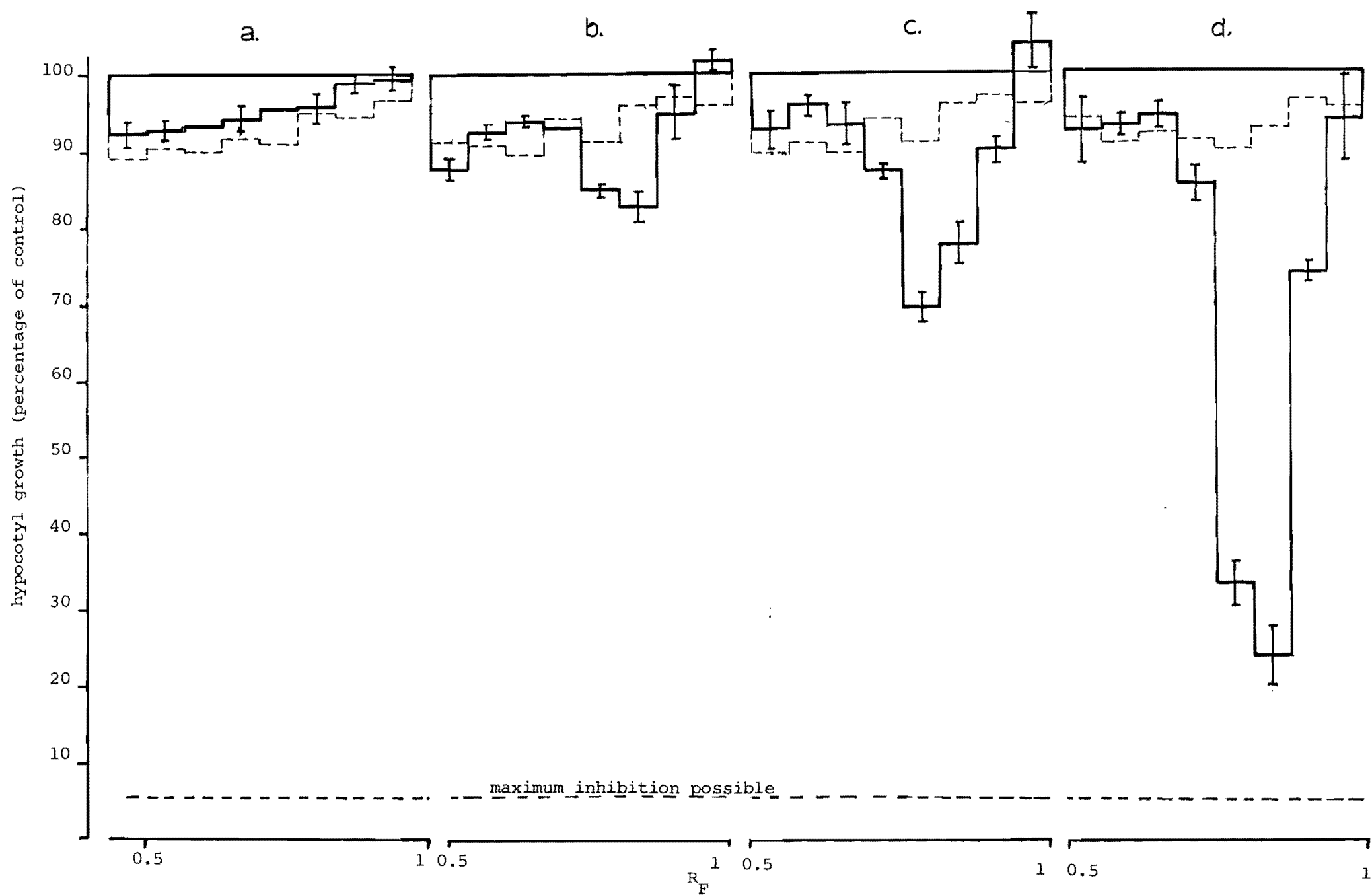


Figure 32. Lettuce hypocotyl assay of the serial dilution of an acid ether-soluble extract obtained from leaves of *Alnus glutinosa* trees harvested on 7 March 1977 (H1). The equivalent of (a) 0.025, (b) 0.125, (c) 0.25 and (d) 0.375 g DW of leaf material was assayed. Other details as in figure 26.

hypocotyl growth (percentage of control)

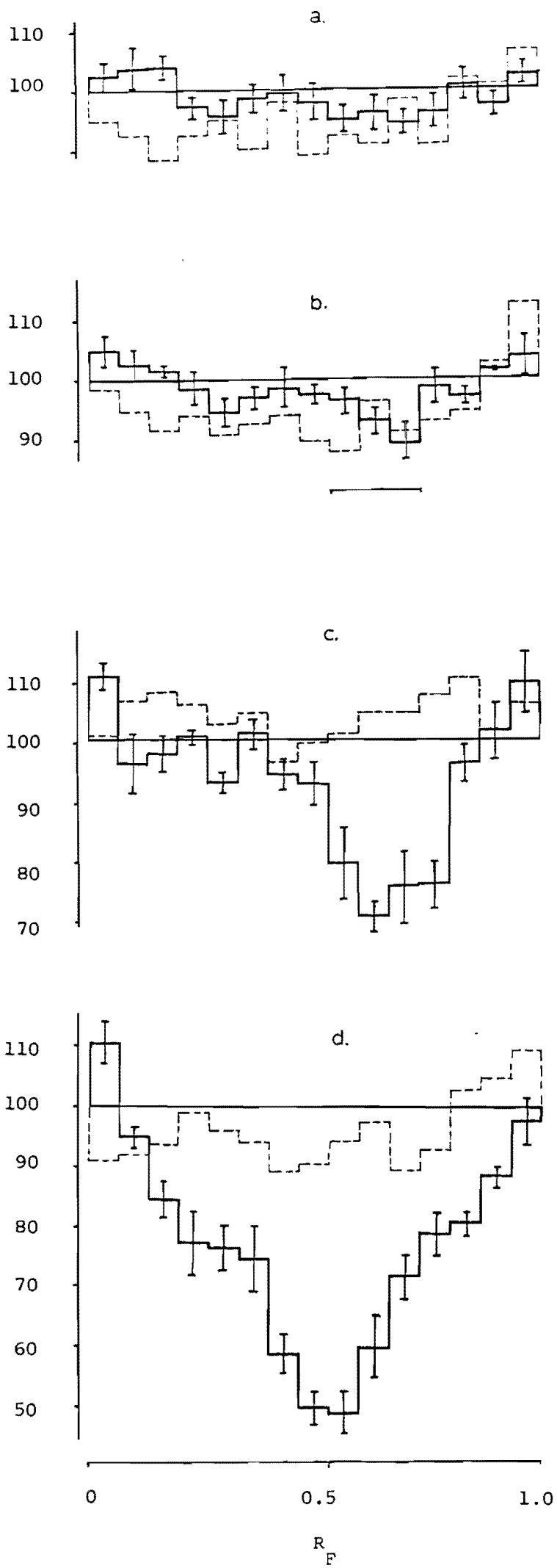


Figure 33. Lettuce hypocotyl assay of the serial dilution of an acidic ether-soluble extract obtained from leaves of *Alnus glutinosa* trees harvested on 15 May 1977 (H5). The equivalent of (a) 0.025, (b) 0.125, (c) 0.25 and (d) 0.375 g DW of leaf material was assayed. Other details as in figure 26.

hypocotyl growth (percentage of control)

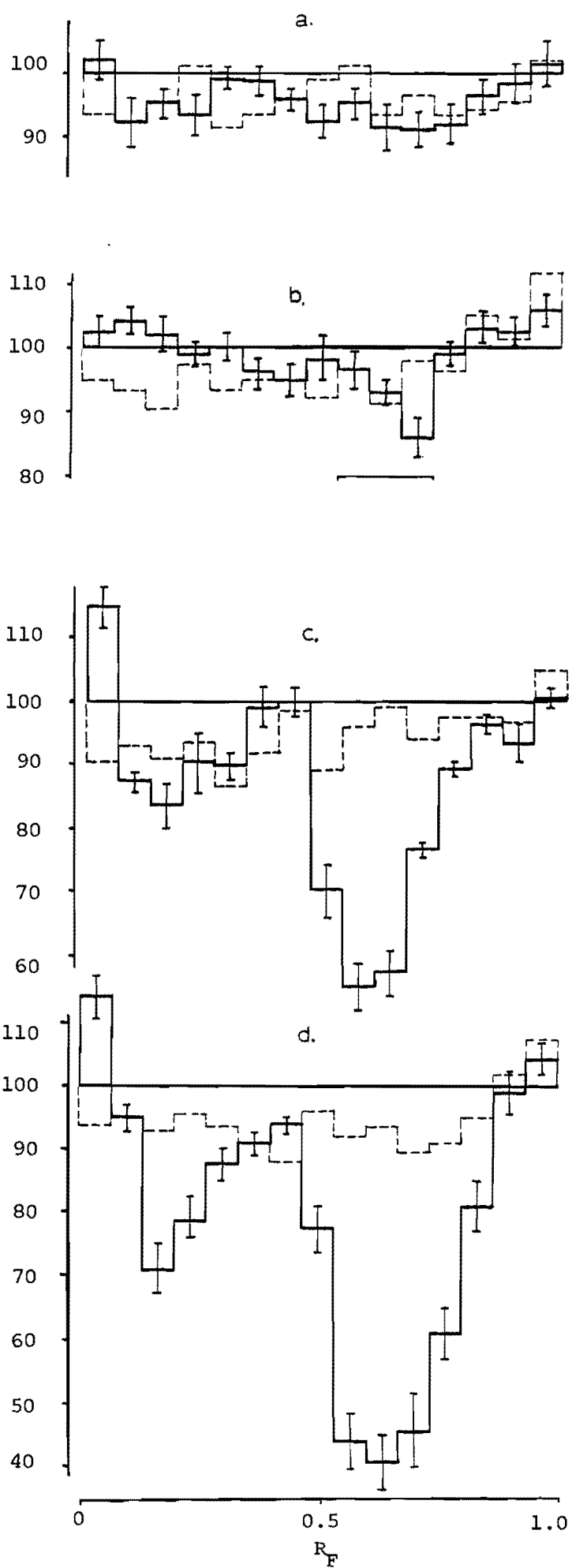


Figure 34. Wheat coleoptile section assay of acid ether-soluble extracts obtained from apical of *Alnus glutinosa* trees harvested on 7 March 1977 (H1). The equivalent of 0.20 g DW of apical material was assayed and the solvent system was isopropanol:NH₃:H₂O (10:1:1::v:v:v). Other details as in figure 18.

Coleoptile growth (percentage of control)

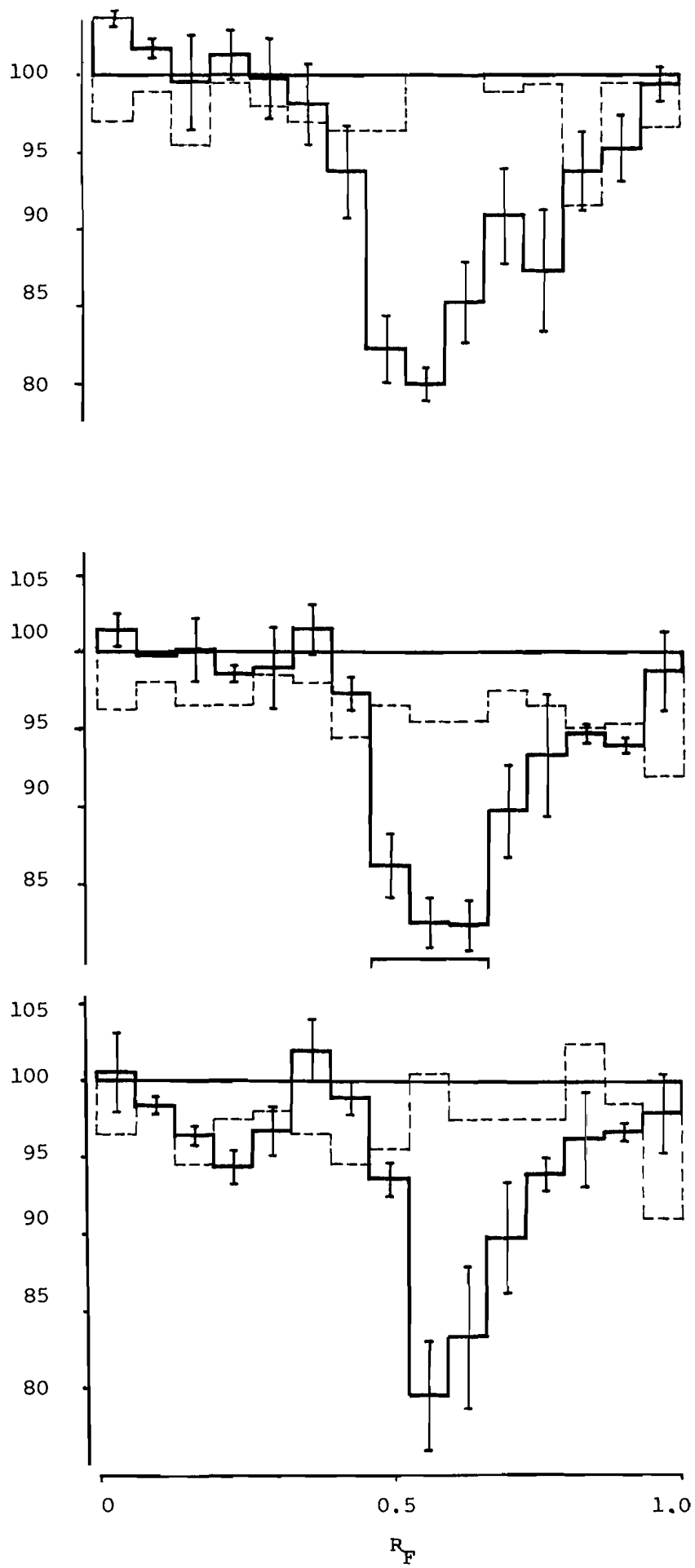


Figure 35. Wheat coleoptile section assay of acidic ether-soluble extracts obtained from apices of *Alnus glutinosa* trees harvested on 25 March 1977 (H2). The equivalent of 0.20 g DW of apical material was assayed and the solvent system was isopropanol:NH₃:H₂O (10:1:1::v:v:v). Other details as in figure 18.

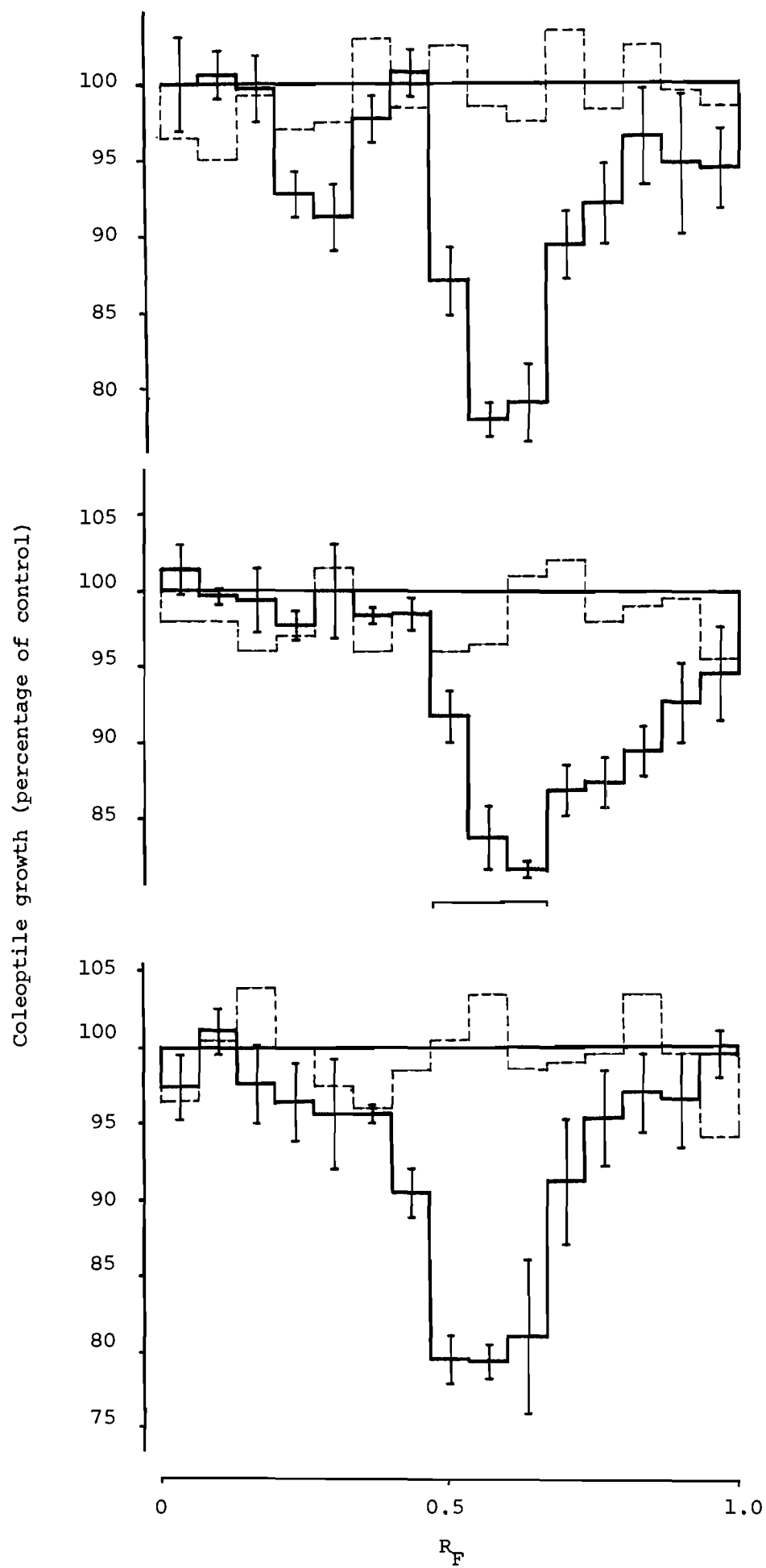


Figure 36. Wheat coleoptile section assay of acidic ether-soluble extracts obtained from apices of *Alnus glutinosa* trees harvested on 15 April 1977 (H3). The equivalent of 0.20 g DW of apical material was assayed and the solvent system was isopropanol:NH₃:H₂O (10:1:1::v:v:v). Other details as in figure 18.

Coleoptile growth (percentage of control)

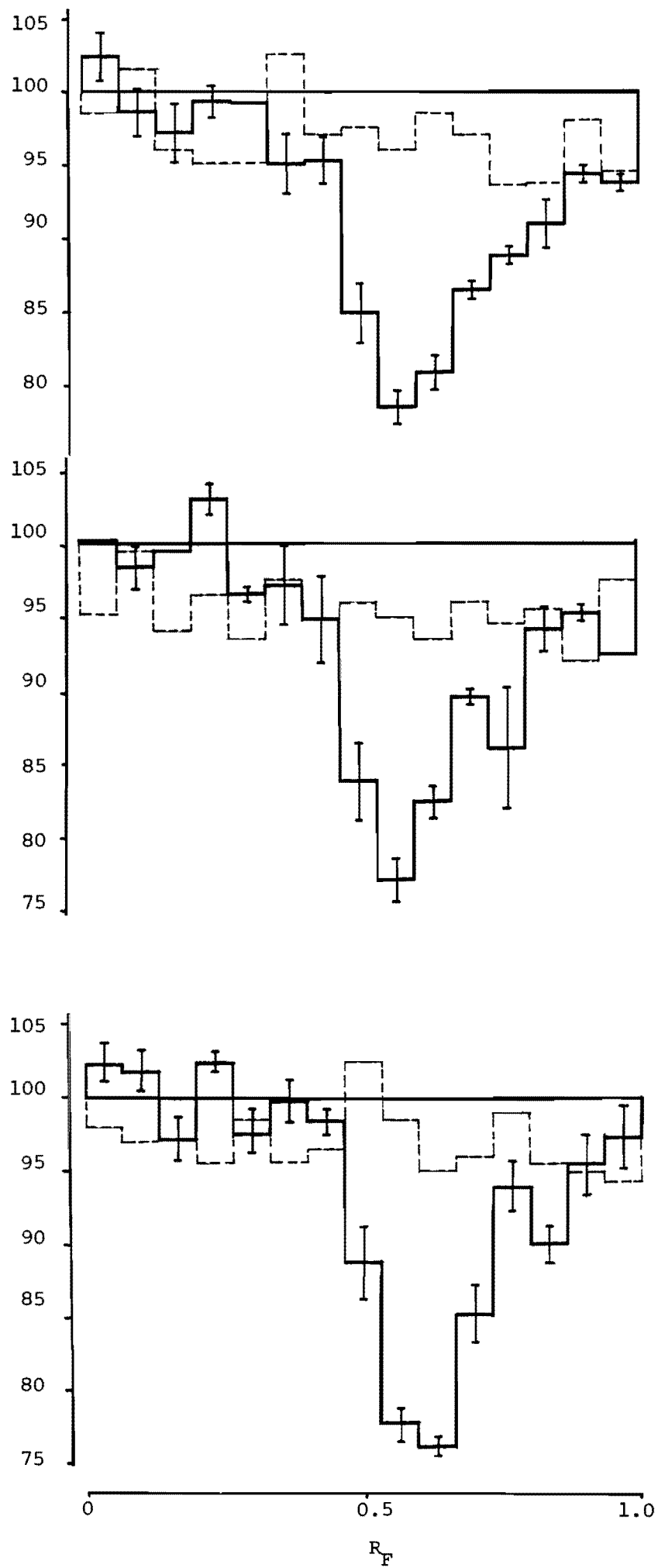


Figure 37. Wheat coleoptile assay of acidic ether-soluble extracts obtained from apices of *Alnus glutinosa* trees harvested on 25 April 1977 (H4). The equivalent of 0.20 g DW of apical material was assayed and the solvent system was isopropanol:NH₃:H₂O (10:1:1::v:v:v). Other details as in figure 18.

Coleoptile growth (percentage of control)

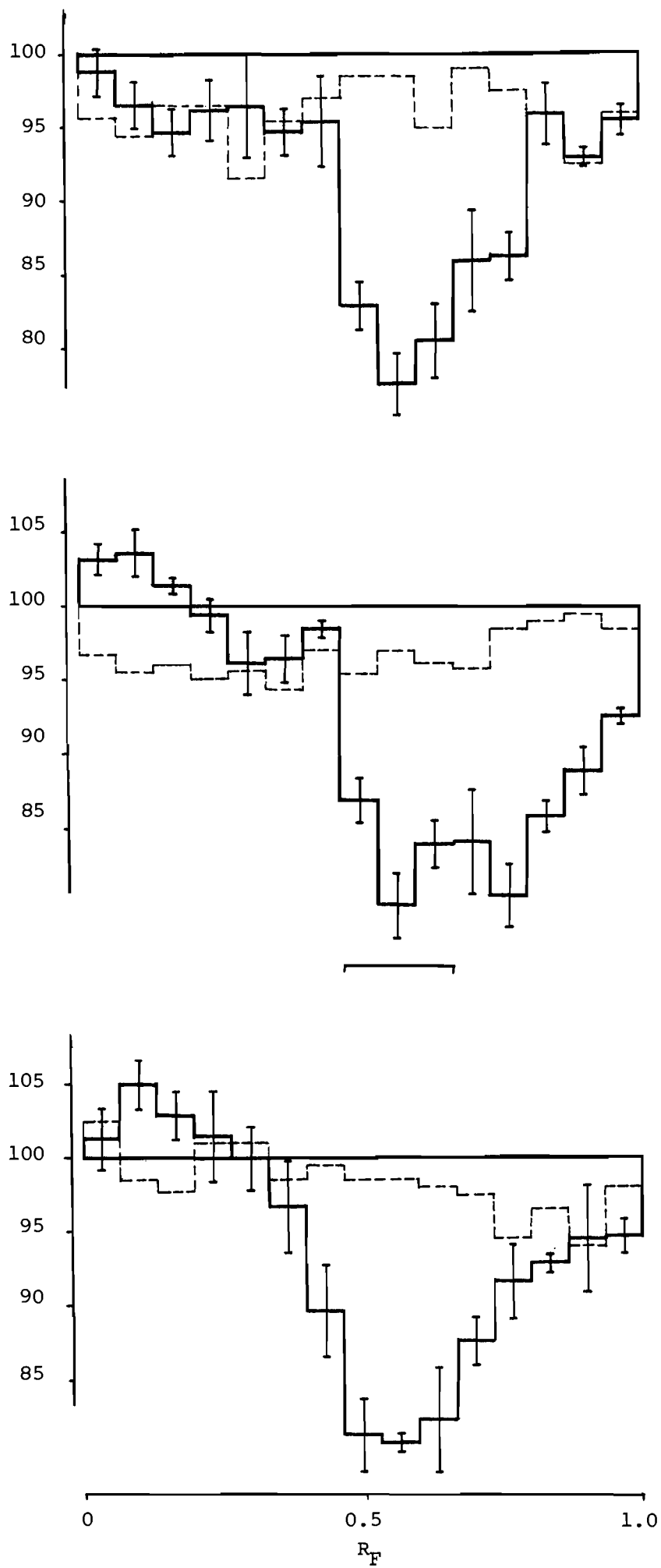


Figure 38. Wheat coleoptile section assay of acidic ether-soluble extracts obtained from apices of *Alnus glutinosa* trees harvested on 15 May 1977 (H5). The equivalent of 0.20 g DW of apical material was assayed and the solvent system was isopropanol:NH₃:H₂O (10:1:1::v:v:v). Other details as in figure 18.

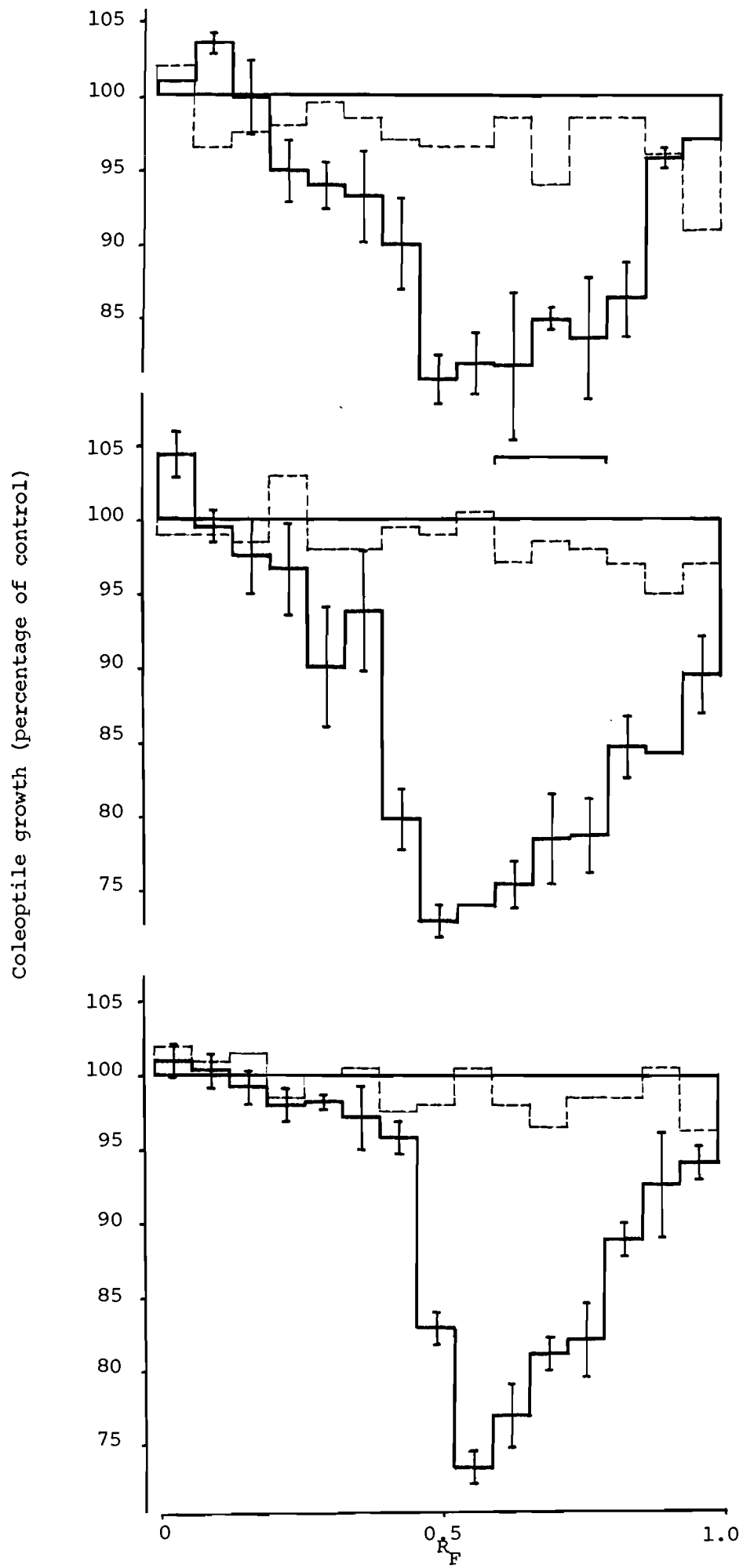
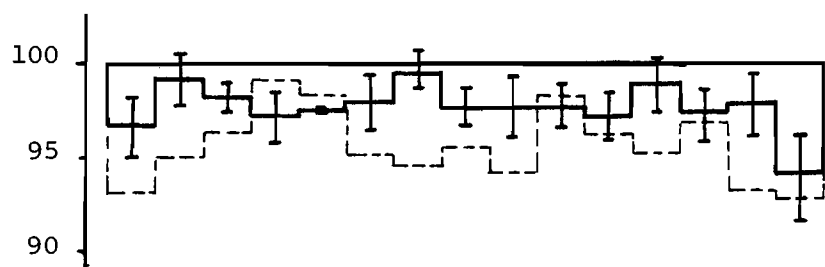


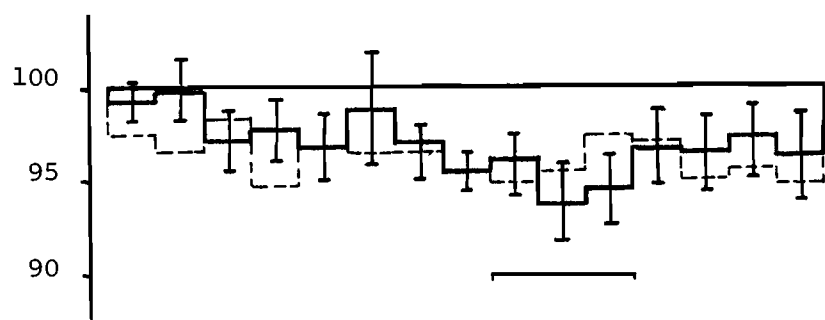
Figure 39. Wheat coleoptile section assay of the serial dilution of an acidic ether-soluble extract obtained from apices of *Alnus glutinosa* trees harvested on 7 March 1977 (H1). The equivalent of (a) 0.02, (b) 0.10 and (c) 0.20 g DW of apical material was assayed. Other details as in figure 18.

Coleoptile growth (percentage of control)

a.



b.



c.

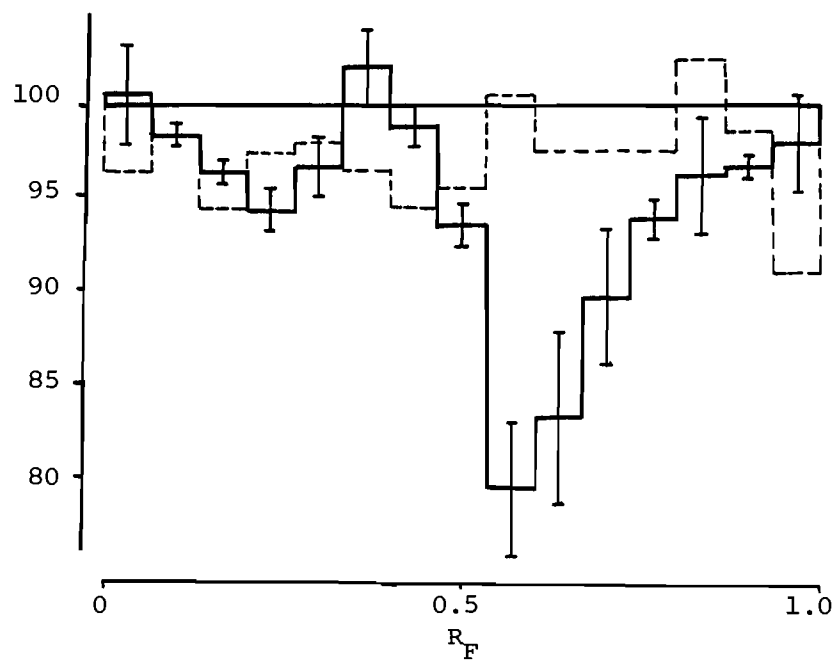
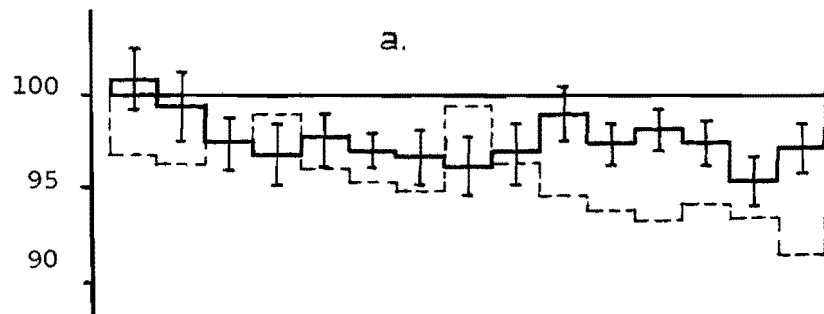


Figure 40. Wheat coleoptile section assay of the serial dilution of an acidic ether-soluble extract obtained from apices of *Alnus glutinosa* trees harvested on 15 May 1977 (H5). The equivalent of (a) 0.02, (b) 0.10 and (c) 0.20 g of apical material was assayed. Other details as in figure 18.



b.



c.

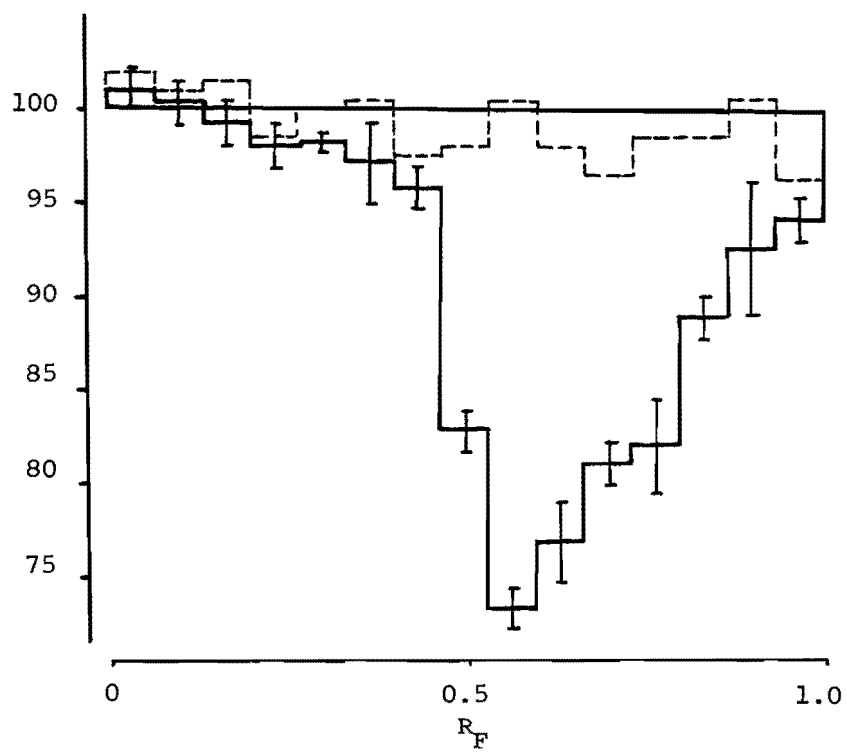


Figure 41. Lettuce hypocotyl assay of acid ether-soluble extracts obtained from apices of *Alnus glutinosa* trees harvested on 7 March 1977 (H1). The equivalent of 0.20 g DW of apical material was assayed and the solvent system was isopropanol:NH₃:H₂O (10:1:1::v:v:v). Other details as in figure 26.

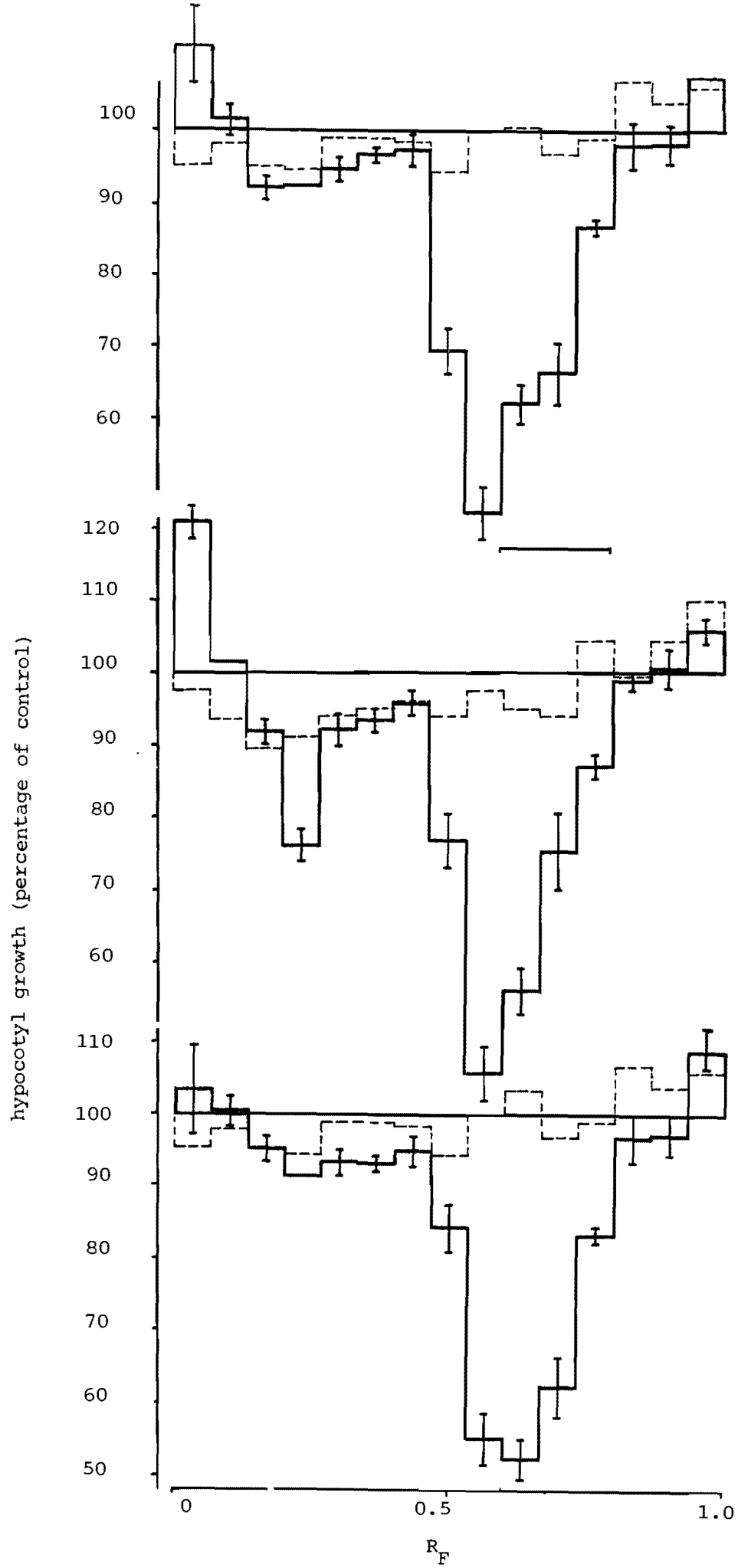


Figure 42. Lettuce hypocotyl assay of acidic ether-soluble extracts obtained from apices of *Alnus glutinosa* trees harvested on 25 March 1977 (H2). The equivalent of 0.20 g DW of apical material was assayed and the solvent system was isopropanol:NH₃:H₂O (10:1:1::v:v:v). Other details as in figure 26.

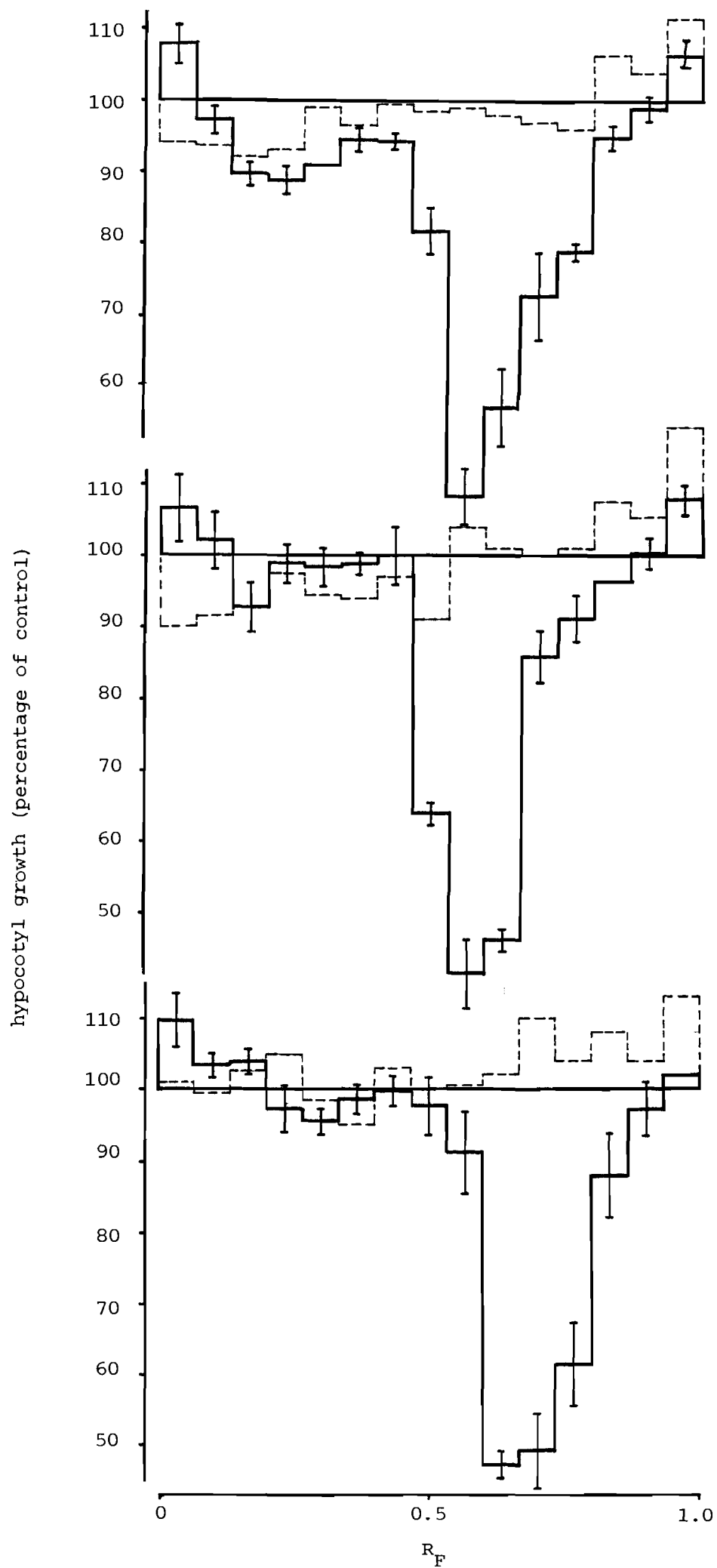


Figure 43. Lettuce hypocotyl assay of acidic ether-soluble extracts obtained from apices of *Alnus glutinosa* trees harvested on 15 April 1977 (H3). The equivalent of 0.20 g DW of apical material was assayed and the solvent system was isopropanol:NH₃:H₂O (10:1:1::v:v:v). Other details as in figure 26.

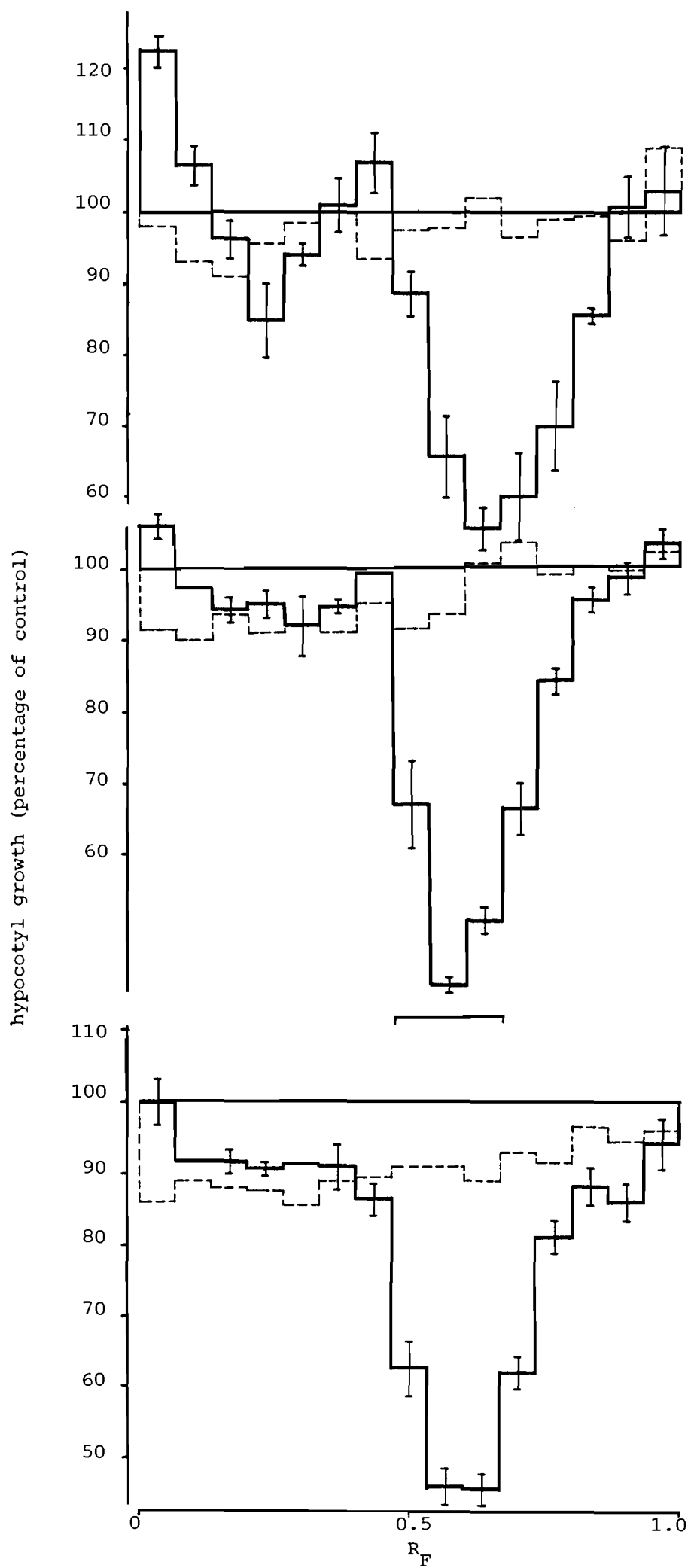


Figure 44. Lettuce hypocotyl assay of acidic ether-soluble extracts obtained from apices of *Alnus glutinosa* trees harvested on 28 April 1977 (H4). The equivalent of 0.20 g DW of apical material was assayed and the solvent system was isopropanol:NH₃:H₂O (10:1:1::v:v:v). Other details as in figure 26.

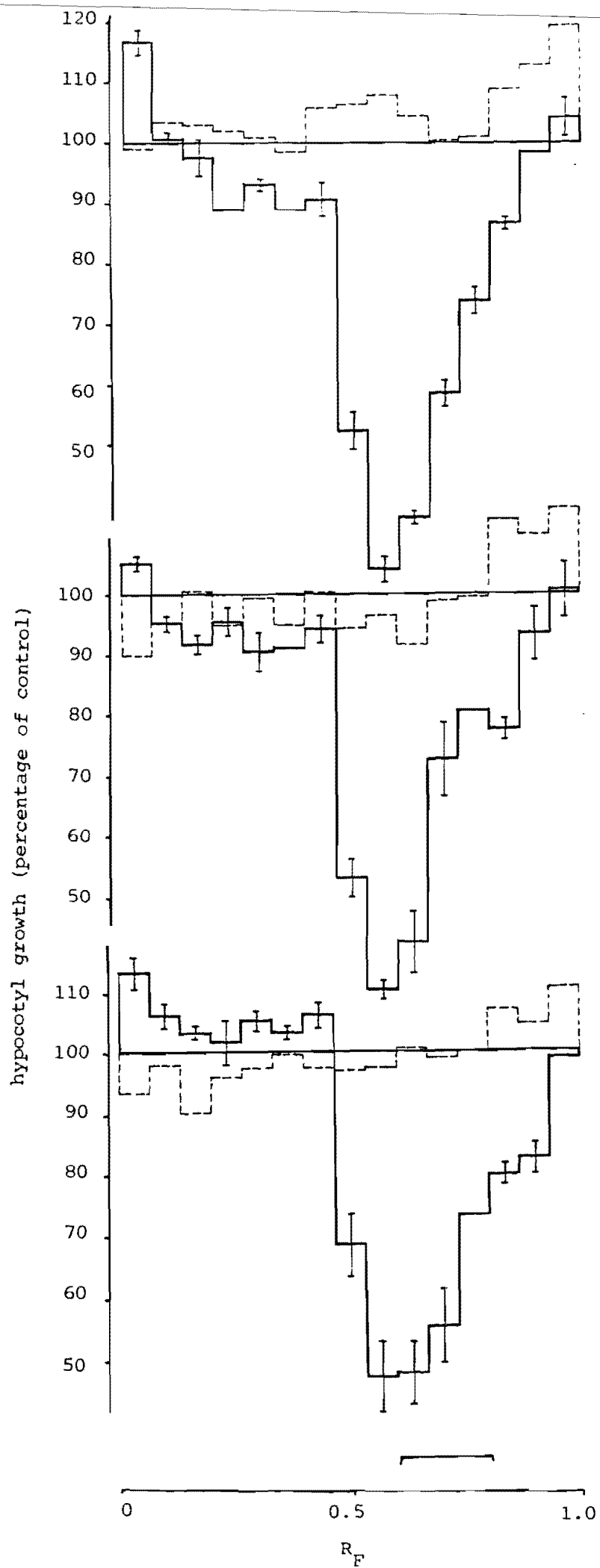


Figure 45. Lettuce hypocotyl assay of acidic ether-soluble extracts obtained from apices of *Alnus glutinosa* trees harvested on 25 May 1977 (H5). The equivalent of 0.20 g DW of dry material was assayed and the solvent system was isopropanol:NH₃:H₂O (10:1:1::v:v:v). Other details as in figure 26.

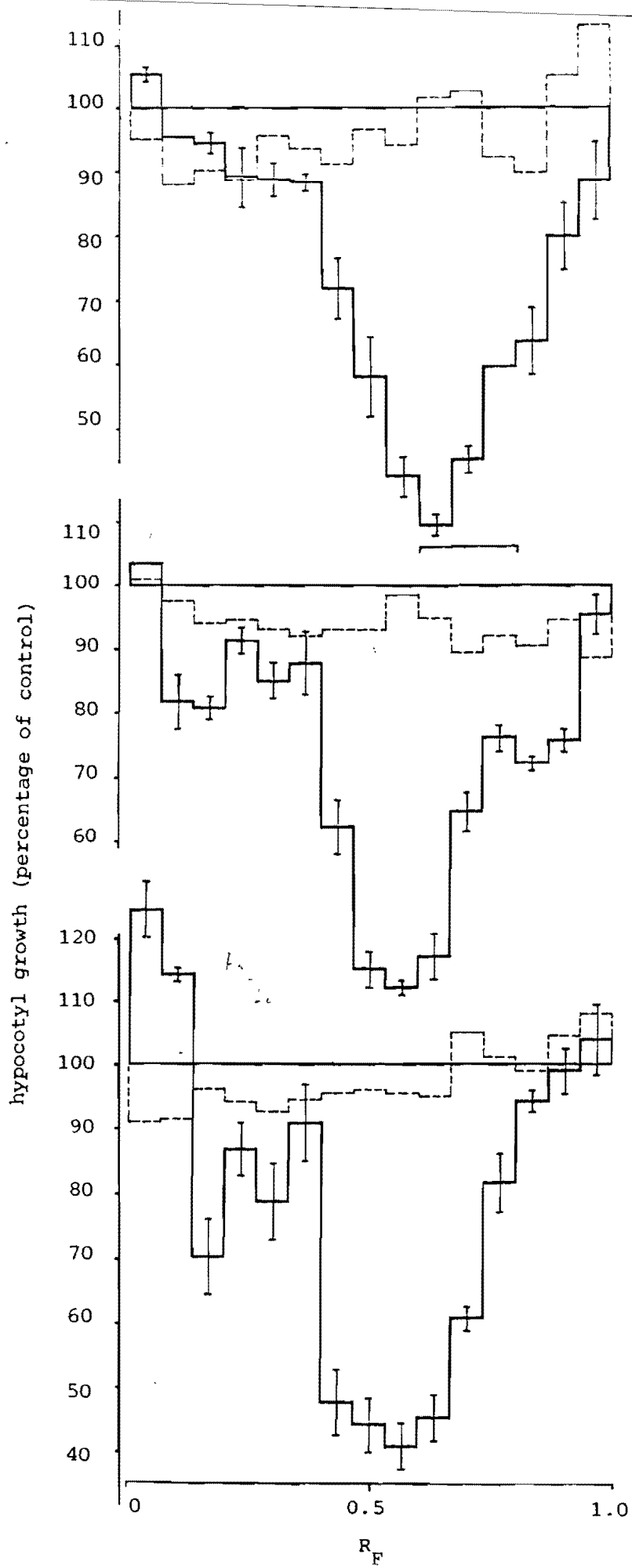


Figure 46. Lettuce hypocotyl assay of the serial dilution of an acidic ether-soluble extract obtained from apices of *Alnus glutinosa* trees harvested on 7 March 1977 (H1). The equivalent of (a) 0.02, (b) 0.10 and (c) 0.20 g DW of apical material was assayed. Other details as in figure 26.

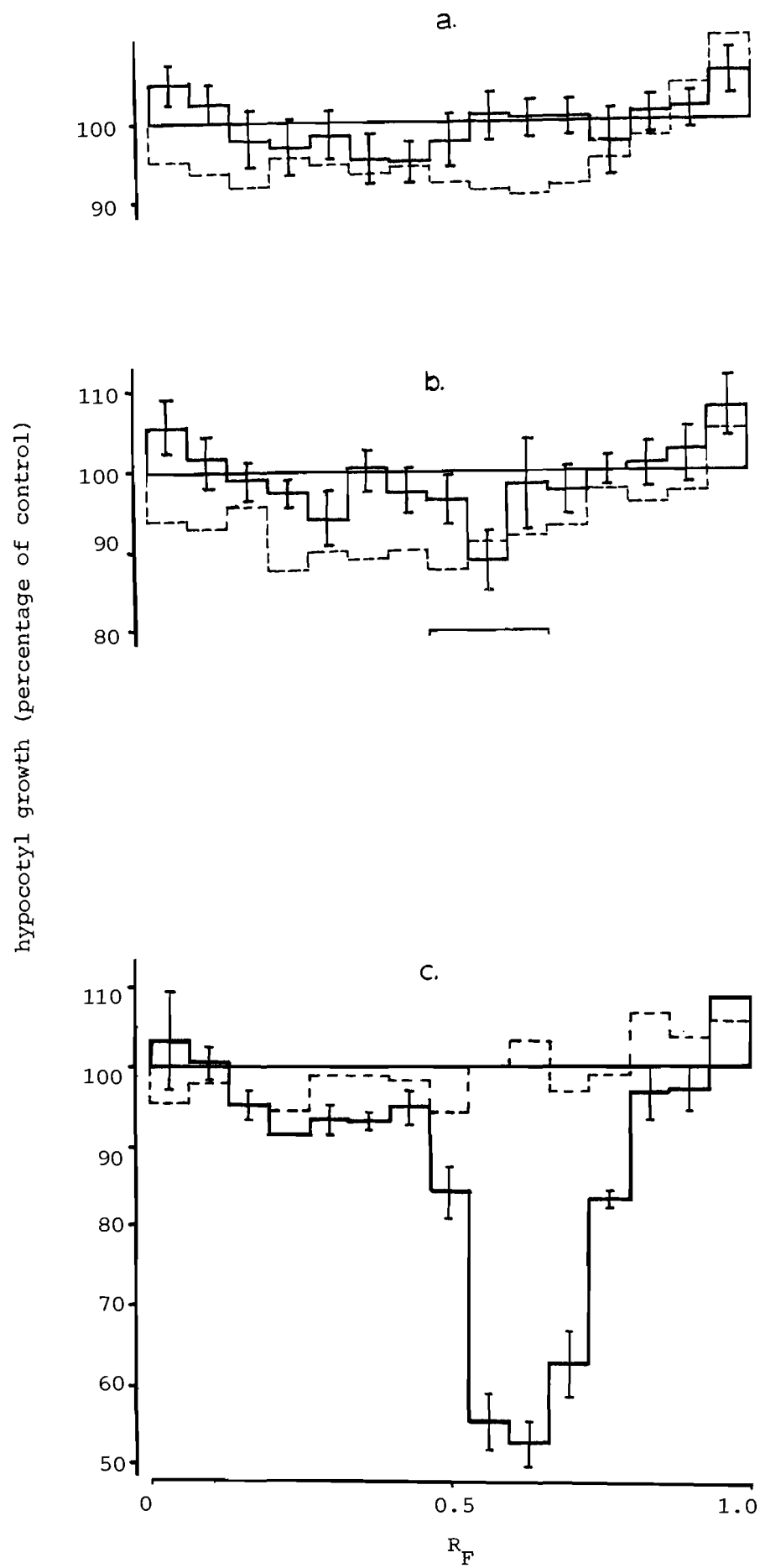


Figure 47. Lettuce hypocotyl assay of the serial dilution of an acid ether-soluble extract obtained from apices of *Alnus glutinosa* trees harvested on 15 May 1977 (H5). The equivalent of (a) 0.02, (b) 0.10 and (c) 0.20 g DW of apical material was assayed. Other details as in figure 26.

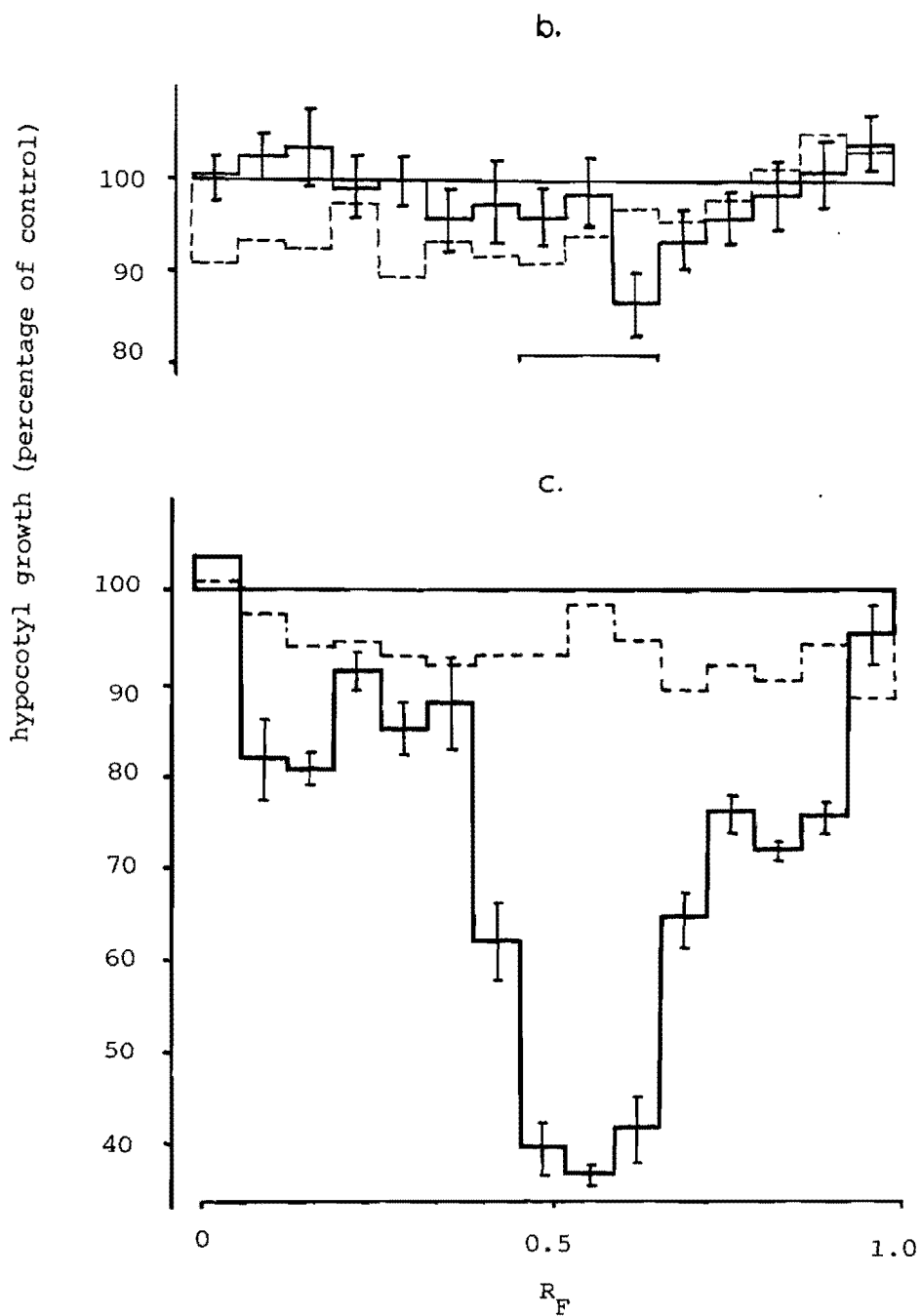
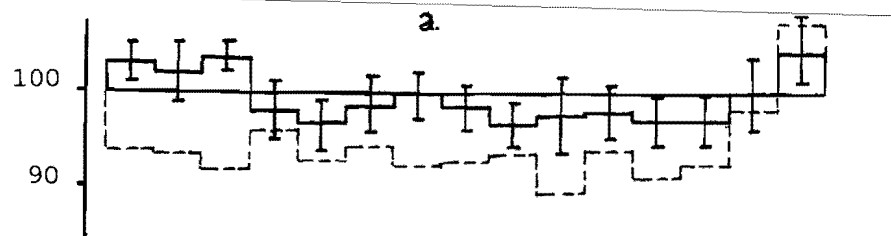
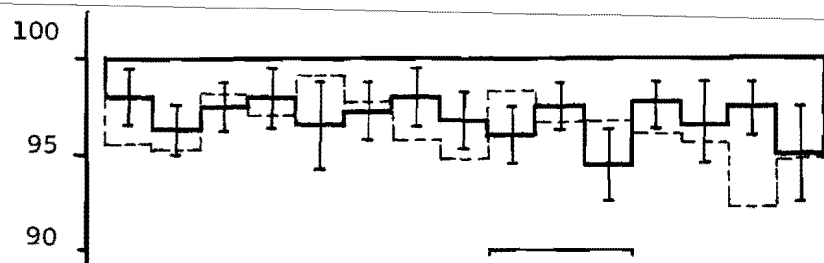
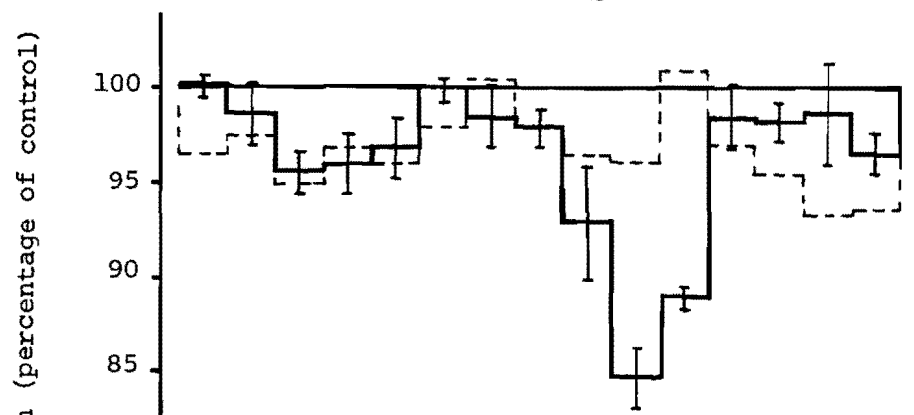


Figure 48. Wheat coleoptile section assay of the serial dilution of an acidic ether-soluble extract obtained from leaves of *Alnus glutinosa* trees harvested on 13 March 1978 (H1). The equivalent of (a) 0.02, (b) 0.125 and (c) 0.25 g DW of leaf material was assayed. Other details as in figure 18.



b.



c.

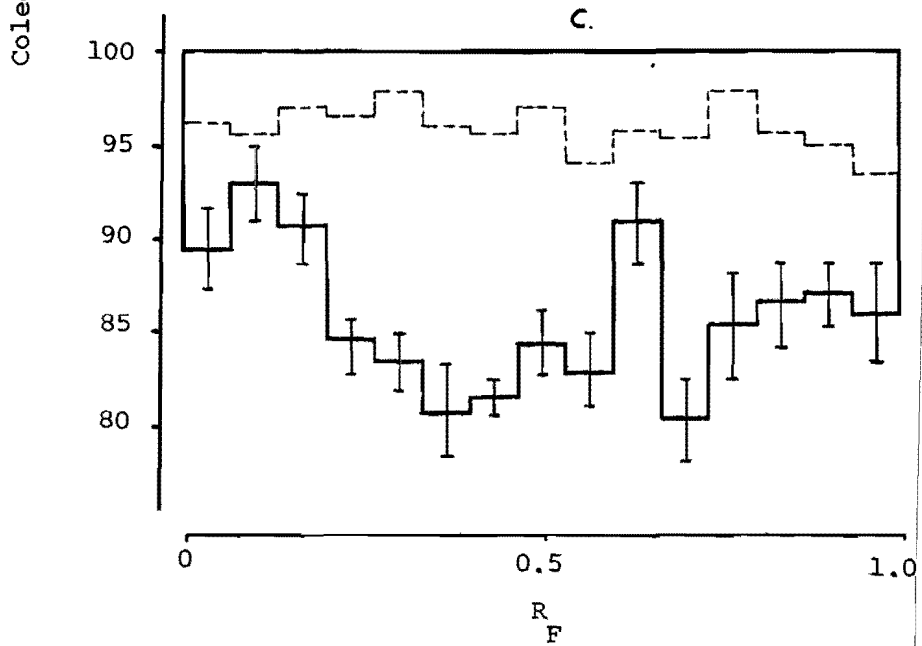


Figure 49. Wheat coleoptile section assay of the serial dilution of an acidic ether-soluble extract obtained from leaves of *Alnus glutinosa* trees harvested on 2 May 1978 (H3). The equivalent of (a) 0.02, (b) 0.125 and (c) 0.25 g DW of leaf material was assayed. Other details as in figure 18.

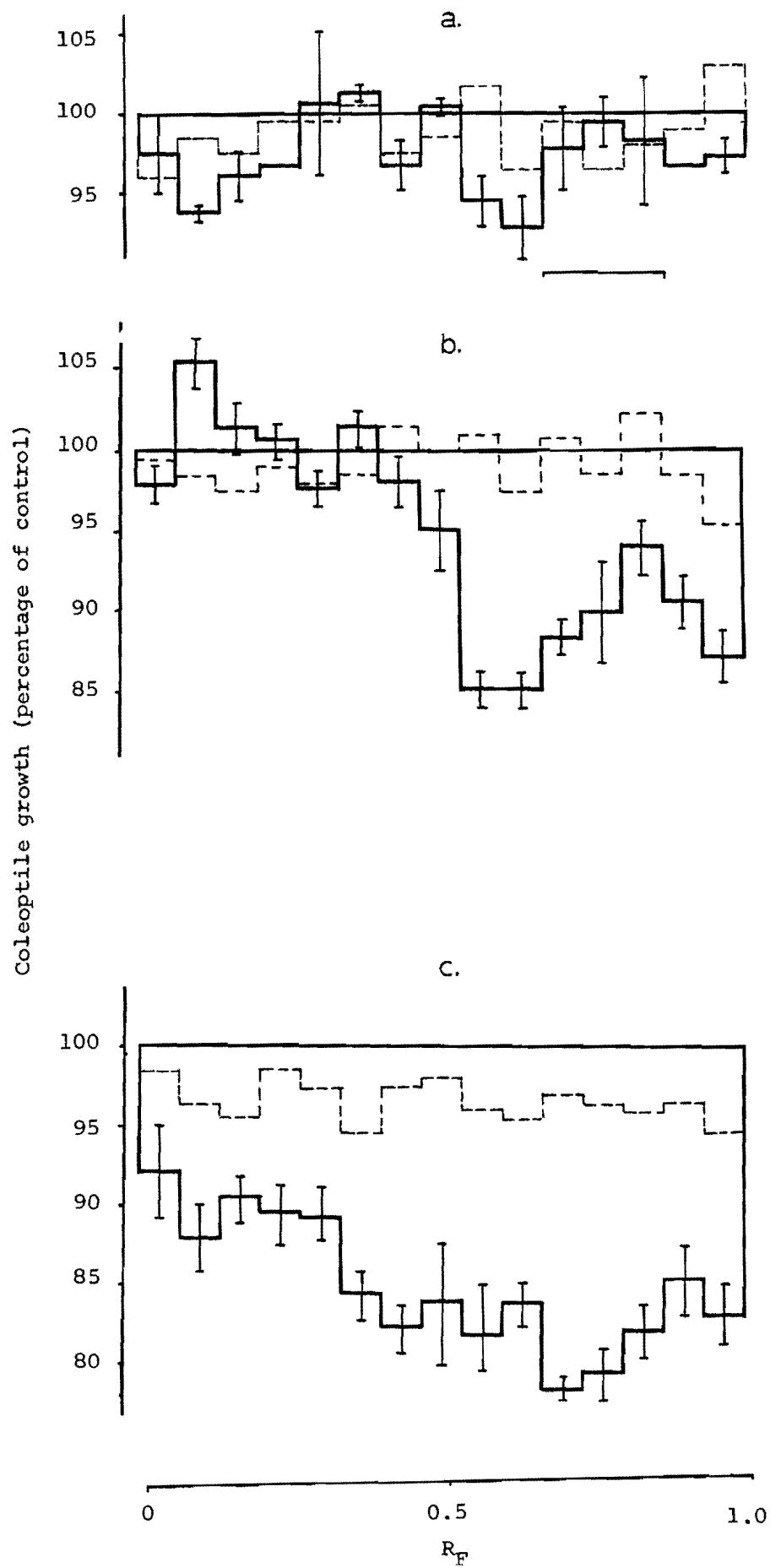


Figure 50. Lettuce hypocotyl assay of the serial dilution of an acid ether-soluble extract obtained from leaves of *Alnus glutinosa* trees harvested on 13 March 1978 (H1). The equivalent of (a) 0.02, (b) 0.125 and (c) 0.25 g DW of leaf material was assayed. Other details as in figure 26.

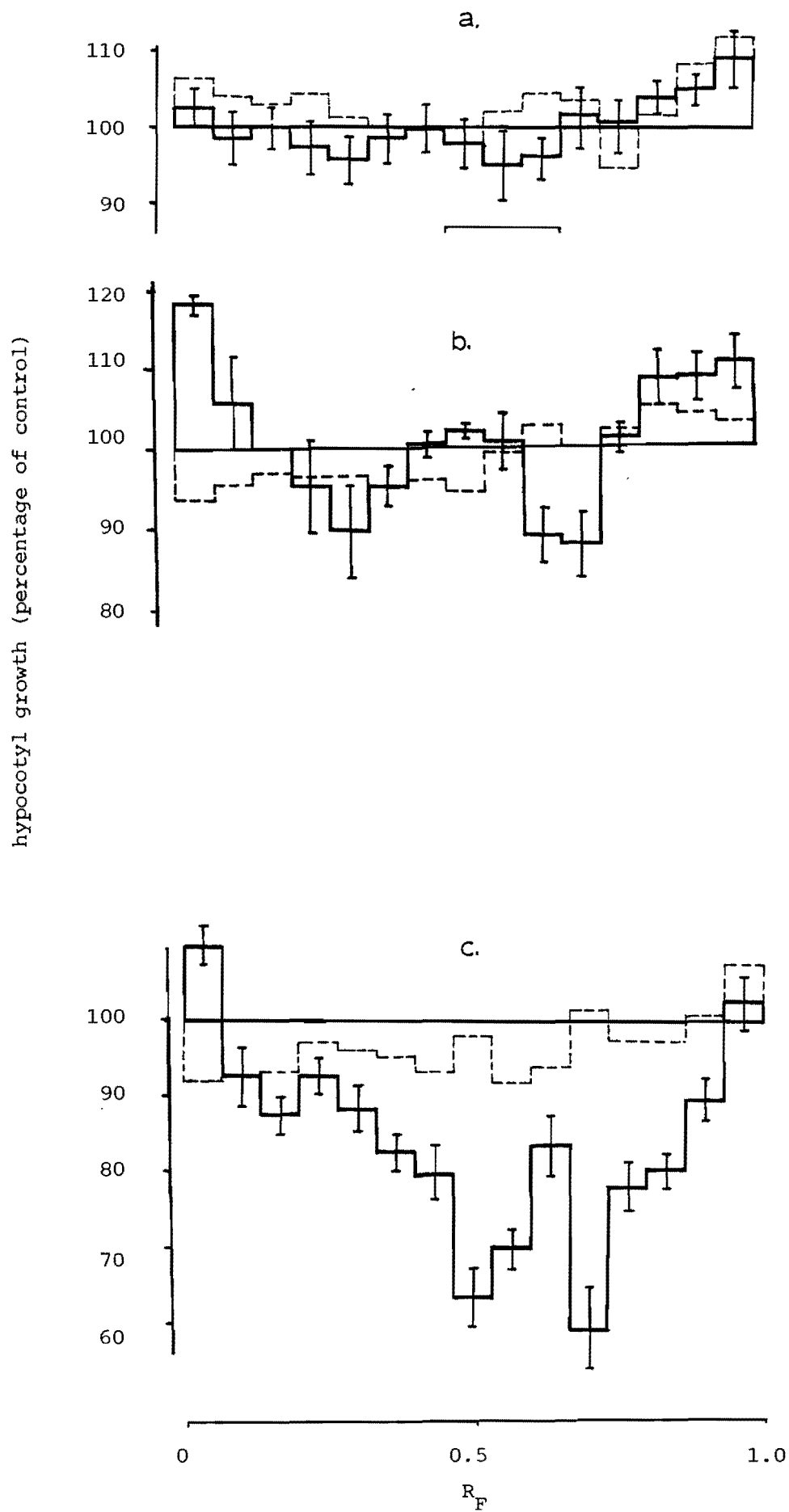


Figure 51. Lettuce hypocotyl assay of the serial dilution of an acid ether-soluble extract obtained from leaves of *Alnus glutinosa* trees harvested on 2 May 1978 (H3). The equivalent of (a) 0.02, (b) 0.125 and (c) 0.25 g DW of apical material was assayed. Other details as in figure 26.

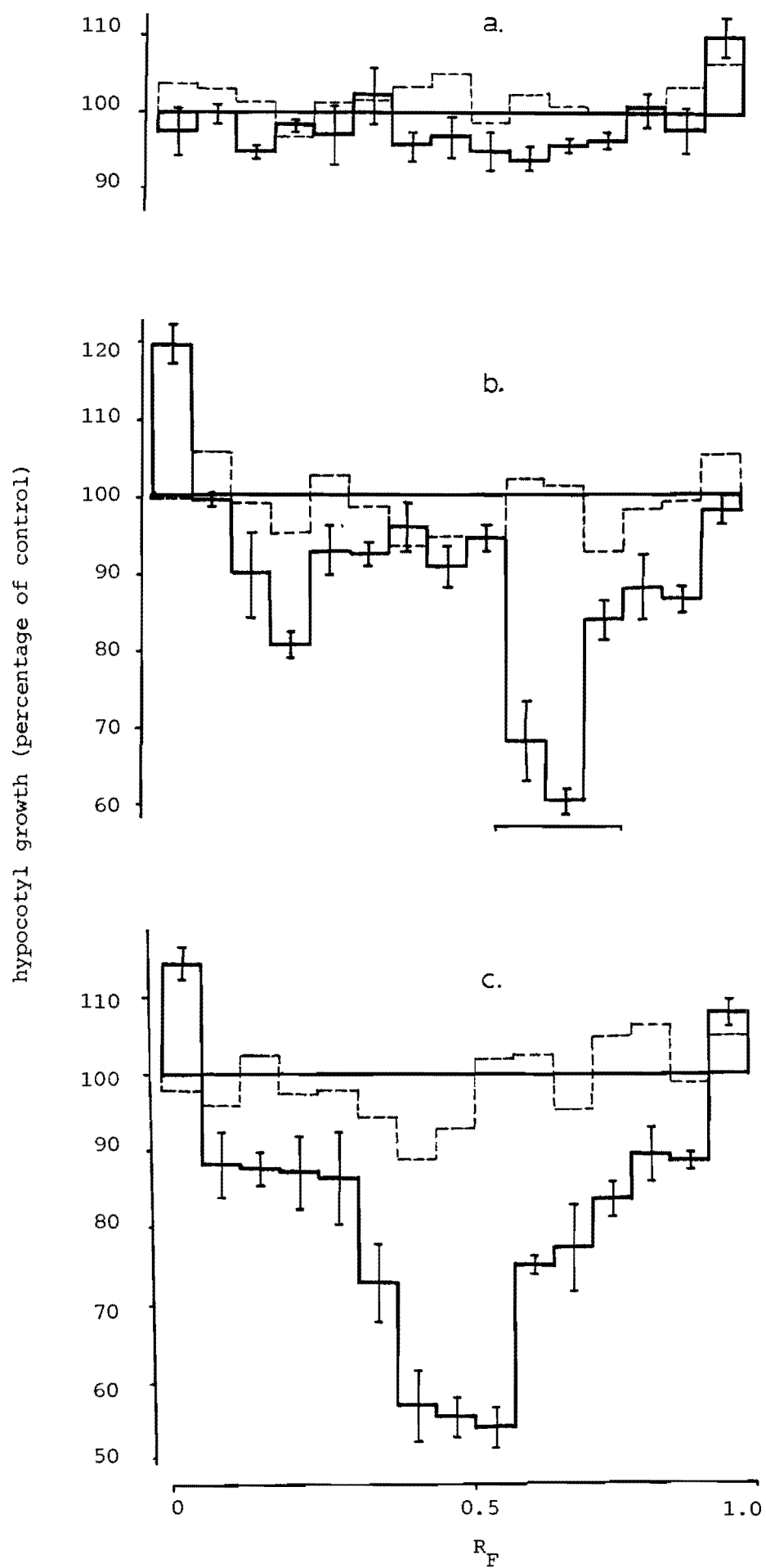


Figure 52. Wheat coleoptile section assay of acidic ether-soluble extracts obtained from leaves of *Alnus glutinosa* trees harvested on 13 March 1978 (H1). The equivalent of 0.125 g DW of leaf material was assayed and the solvent system was isopropanol:NH₃:H₂O (10:1:1::v:v:v). Other details as in figure 18.

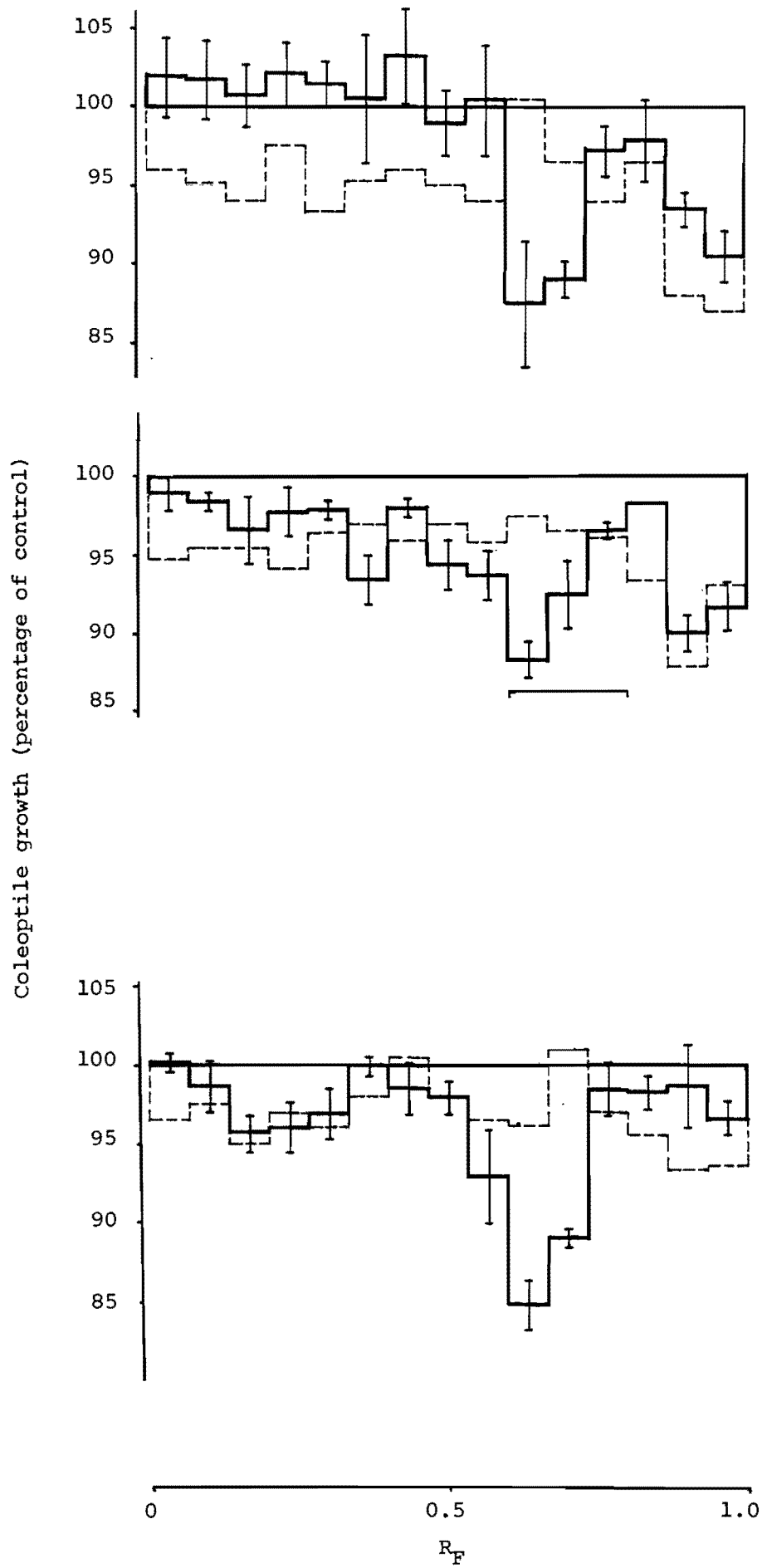


Figure 53. Wheat coleoptile section assay of acidic ether-soluble extracts obtained from leaves of *Alnus glutinosa* trees harvested on 14 April 1978 (H2). The equivalent of 0.125 g DW of leaf material was assayed and the solvent system was isopropanol:NH₃:H₂O (10:1:1::v:v:v). Other details as in figure 18.

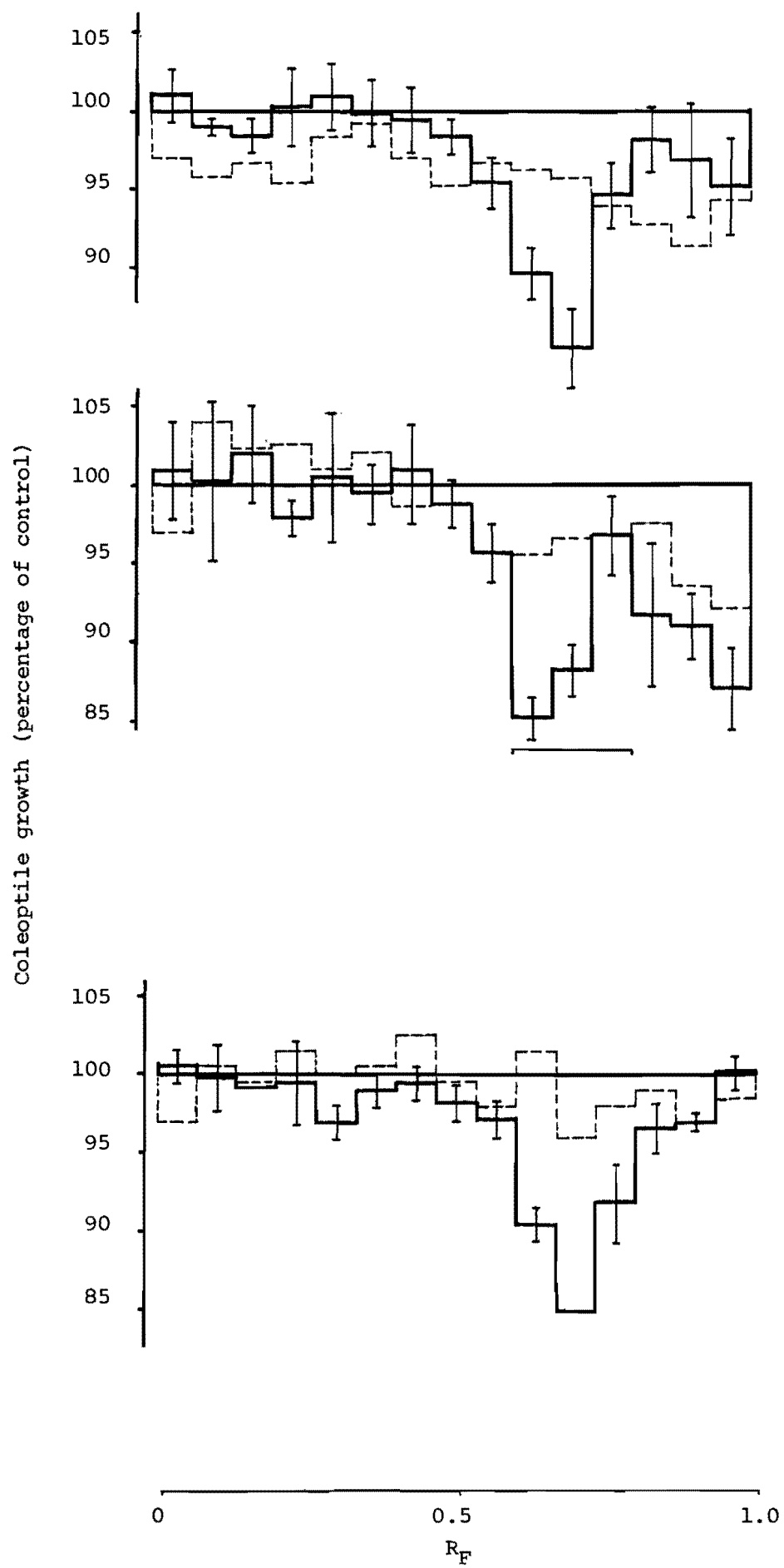


Figure 54. Wheat coleoptile section assay of acidic ether-soluble extracts obtained from leaves of *Alnus glutinosa* trees harvested on 2 May 1978 (H3). The equivalent of 0.125 g DW of leaf material was assayed and the solvent system was isopropanol:NH₃:H₂O (10:1:1::v:v:v). Other details as in figure 18.

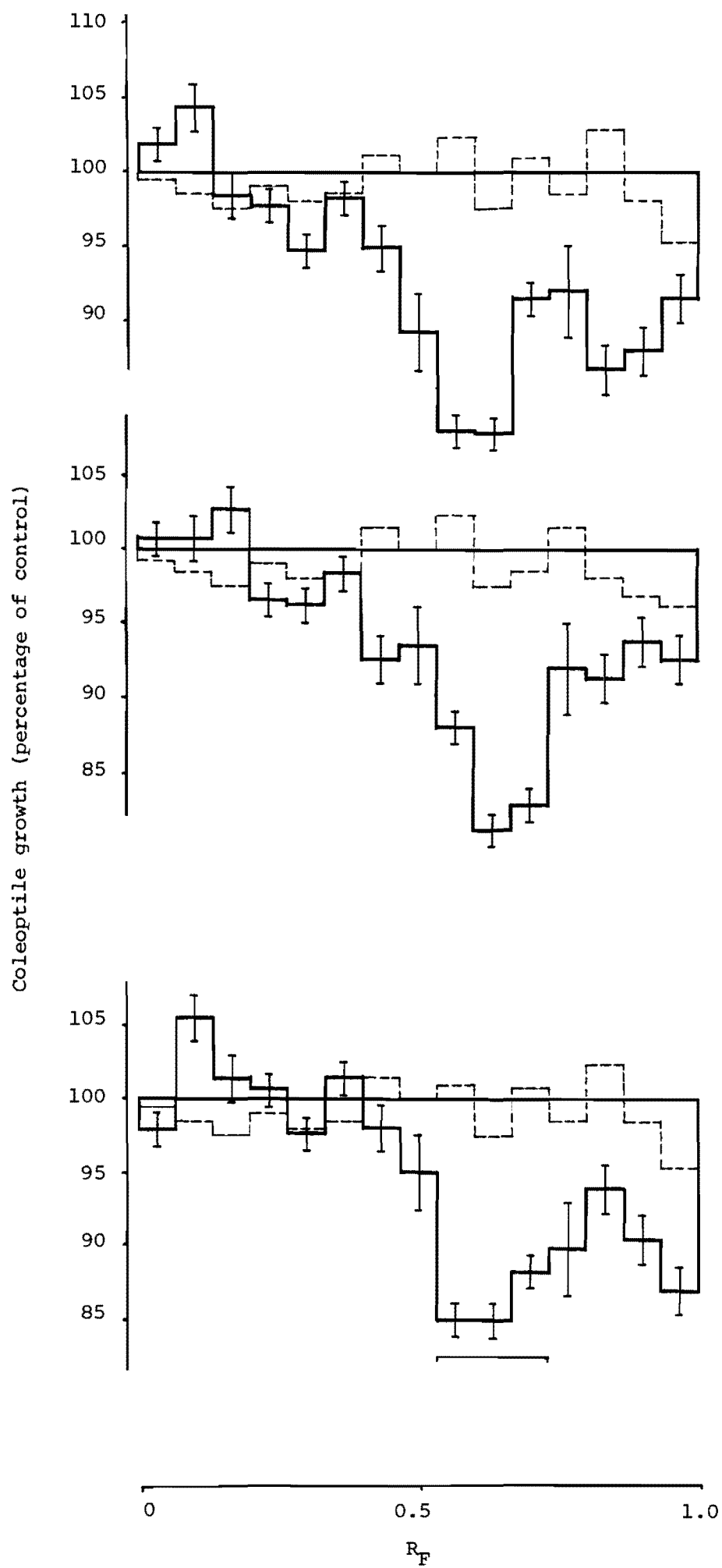


Figure 55. Lettuce hypocotyl assay of acidic ether-soluble extracts obtained from leaves of *Alnus glutinosa* trees harvested on 13 March 1978 (H1). The equivalent of 0.125 g DW of leaf material was assayed and the solvent system was isopropanol:NH₃:H₂O (10:1:1::v:v:v). Other details as in figure 26.

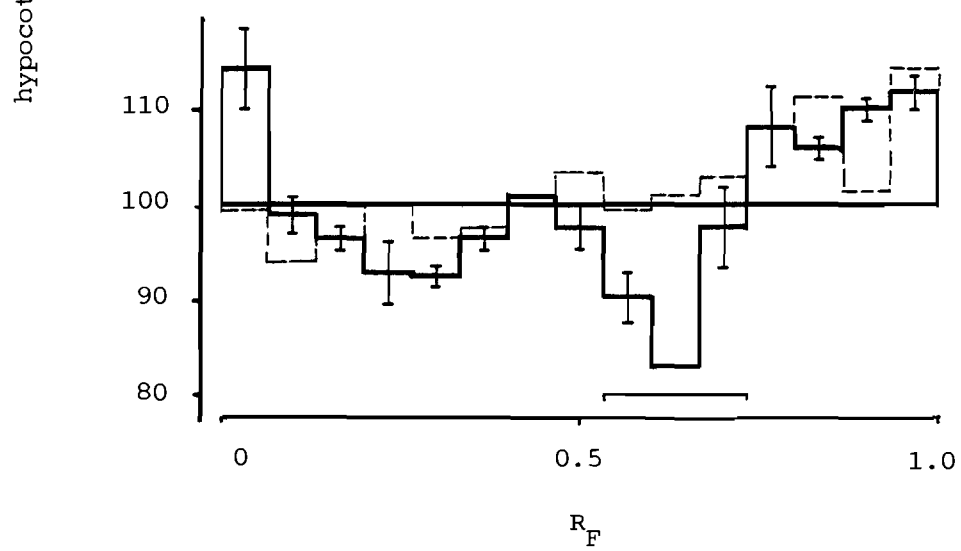
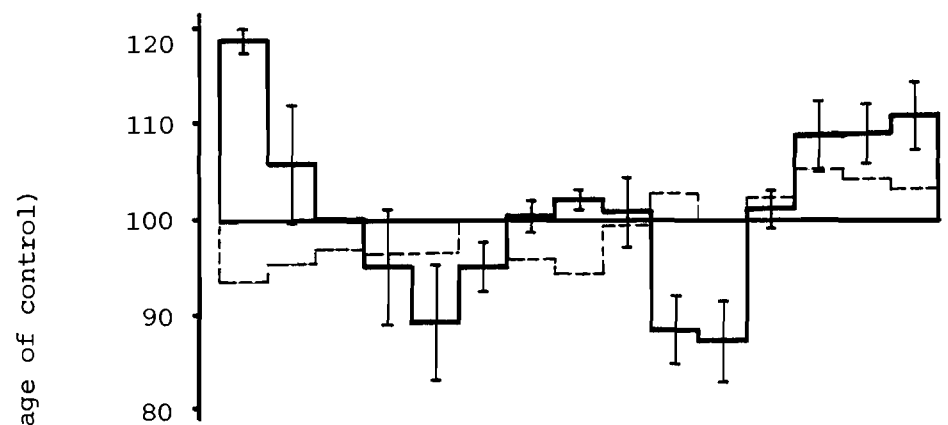
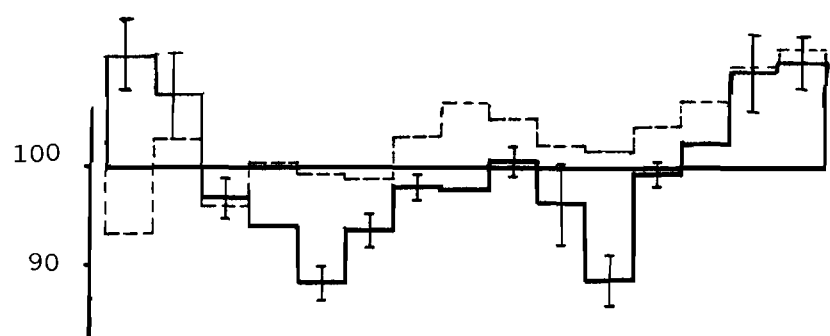


Figure 56. Lettuce hypocotyl assay of acidic ether-soluble extracts obtained from leaves of *Alnus glutinosa* trees harvested on 14 April 1978 (H2). The equivalent of 0.125 g DW of leaf material was assayed and the solvent system was isopropanol:NH₃:H₂O (10:1:1::v:v:v). Other details as in figure 26.

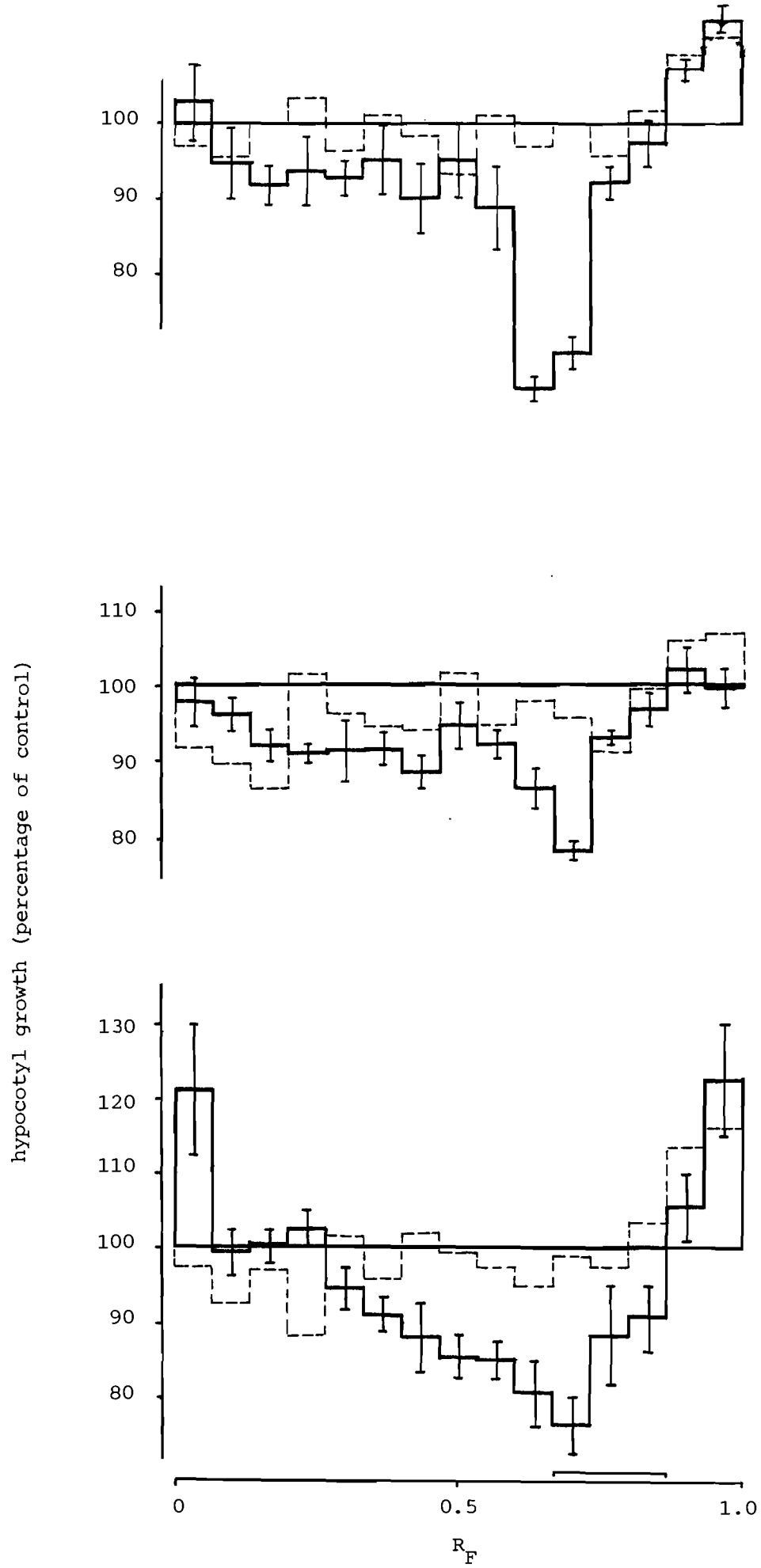


Figure 57. Lettuce hypocotyl assay of acidic ether-soluble extracts obtained from leaves of *Alnus glutinosa* trees harvested on 2 May 1978 (H3). The equivalent of 0.125 g DW of leaf material was assayed and the solvent system was isopropanol:NH₃:H₂O (10:1:1::v:v:v). Other details as in figure 26.

hypocotyl growth (percentage of control)

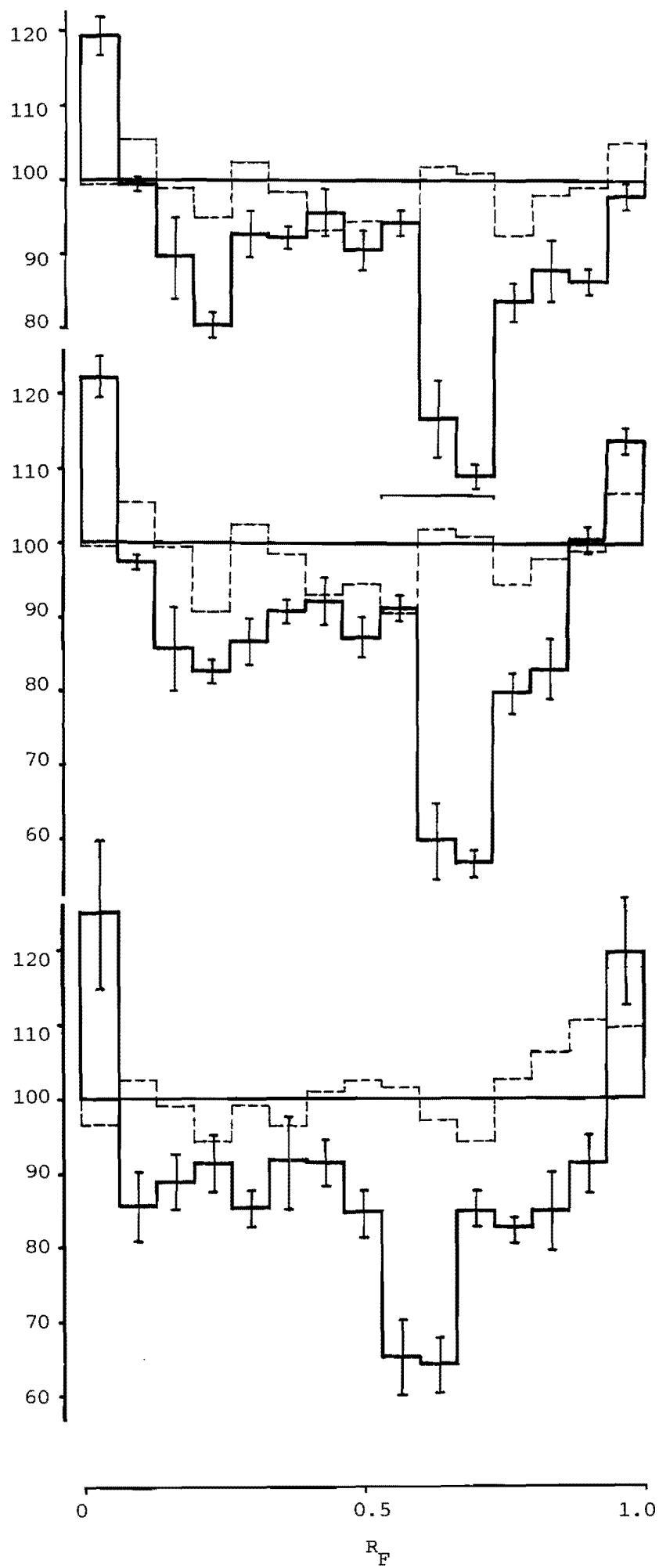


Figure 58. Wheat coleoptile section assay of acidic ether-soluble extracts obtained from apices of *Alnus glutinosa* trees harvested on 13 March 1978 (H1). The equivalent of 0.20 g DW of apical material was assayed and the solvent system was isopropanol:NH₃:H₂O (10:1:1::v:v:v). Other details as in figure 18.

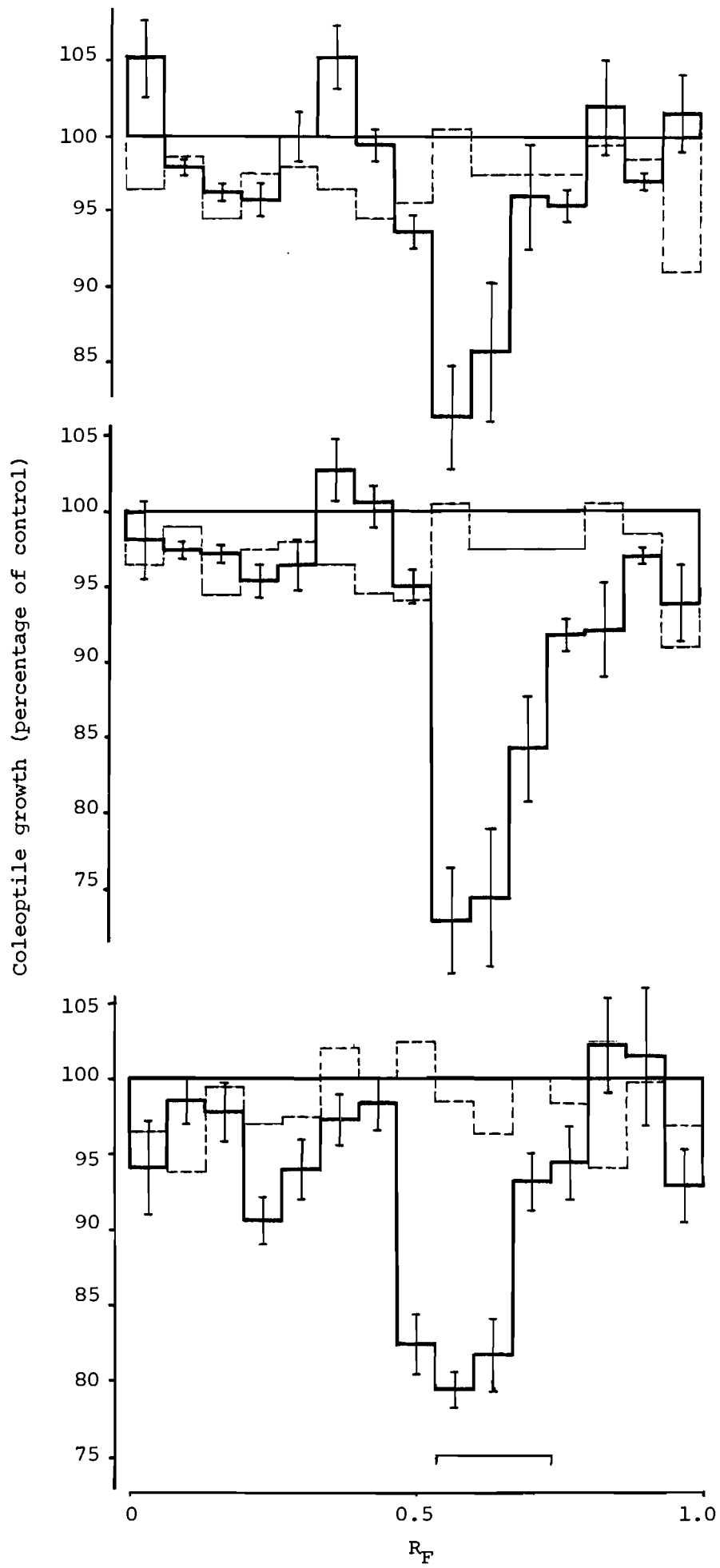


Figure 59. Wheat coleoptile section assay of acidic ether-soluble extracts obtained from apices of *Alnus glutinosa* trees harvested on 14 April 1978 (H2). The equivalent of 0.20 g DW of apical material was assayed and the solvent system was isopropanol:NH₃:H₂O (10:1:1::v:v:v). Other details as in figure 18.

Coleoptile growth (percentage of control)

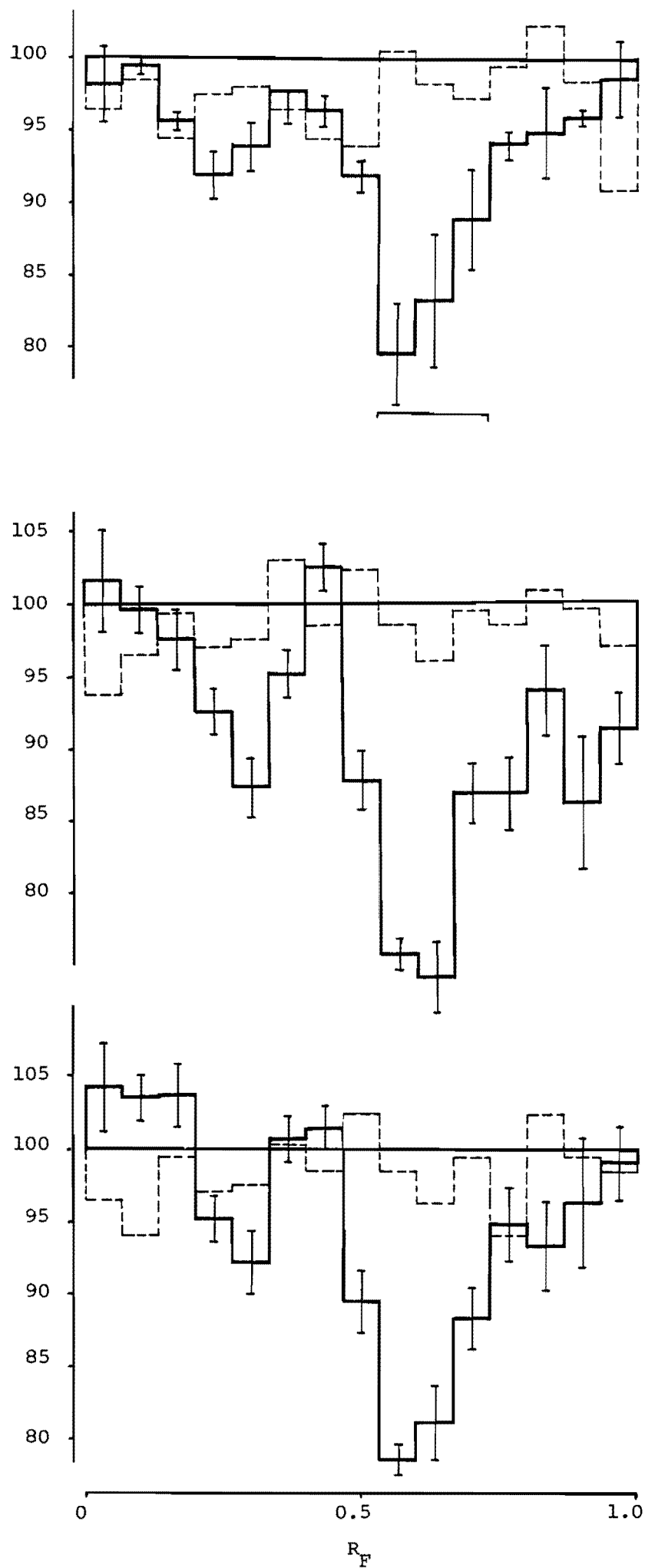


Figure 60. Wheat coleoptile section assay of acidic ether-soluble extracts obtained from apices of *Alnus glutinosa* trees harvested on 2 May 1978 (H3). The equivalent of 0.20 g DW of apical material was assayed and the solvent system was isopropanol:NH₃:H₂O (10:1:1::v:v:v). Other details as in figure 18.

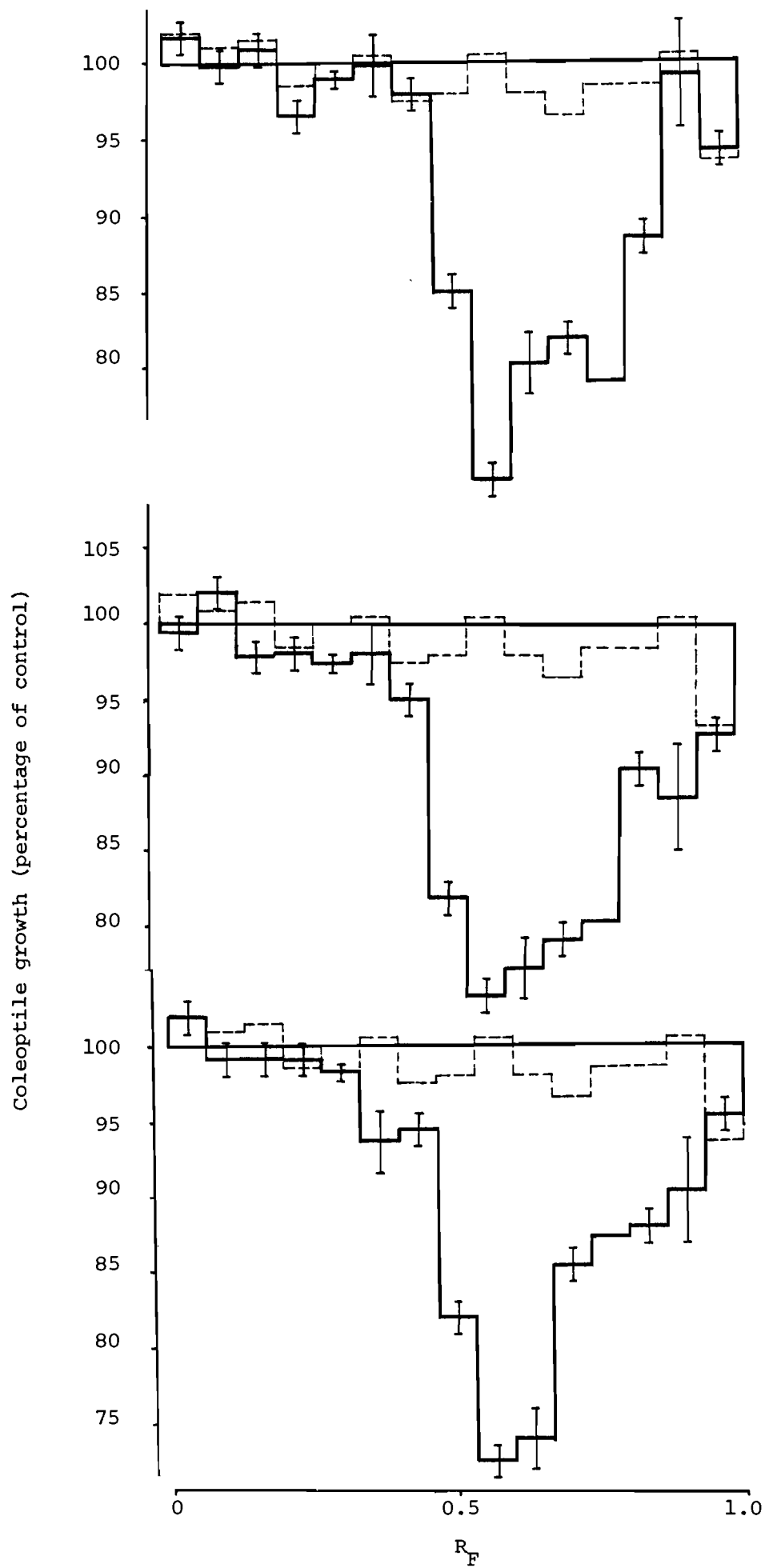


Figure 61. Lettuce hypocotyl assay of acidic ether-soluble extracts obtained from apices of *Alnus glutinosa* trees harvested on 13 March 1978 (H1). The equivalent of 0.20 g DW of apical material was assayed and the solvent system was isopropanol:NH₃:H₂O (10:1:1::v:v:v). Other details as in figure 26.

hypocotyl growth (percentage of control)

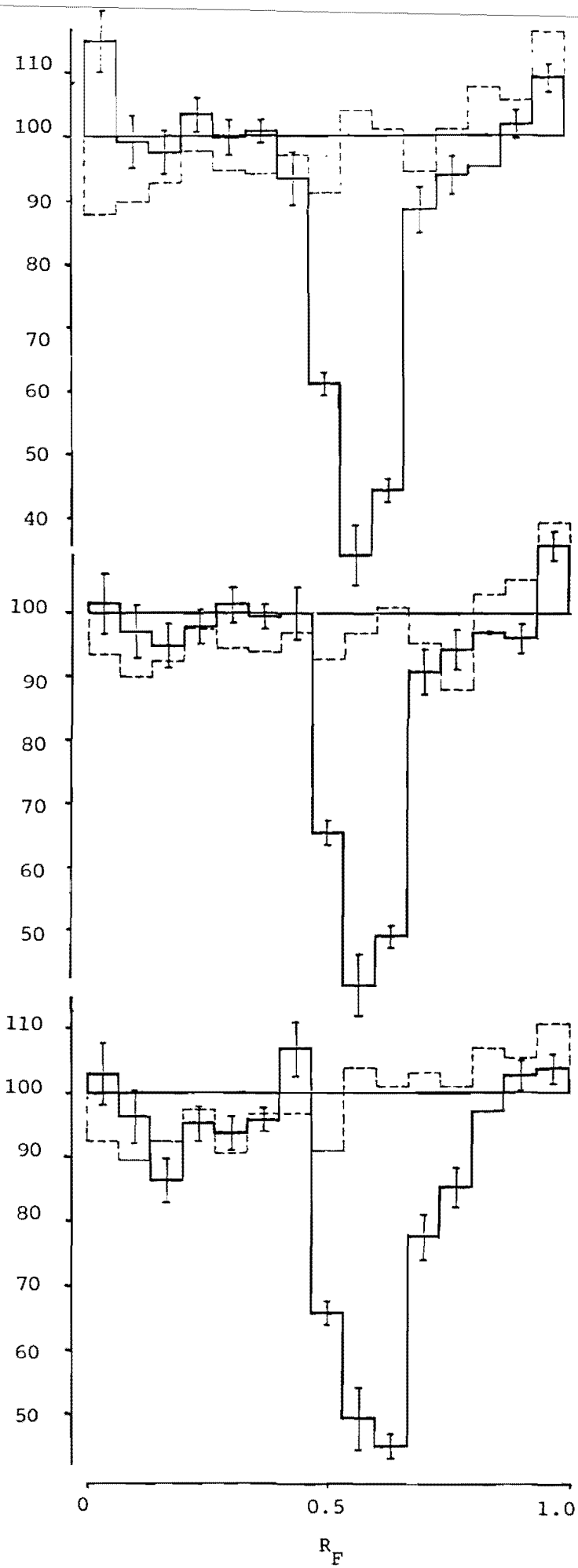


Figure 62. Lettuce hypocotyl assay of acidic ether-soluble extracts obtained from apices of *Alnus glutinosa* trees harvested on 14 April 1978 (H2). The equivalent of 0.20 g DW of apical material was assayed and the solvent system was isopropanol:NH₃:H₂O (10:1:1::v:v:v). Other details as in figure 26.

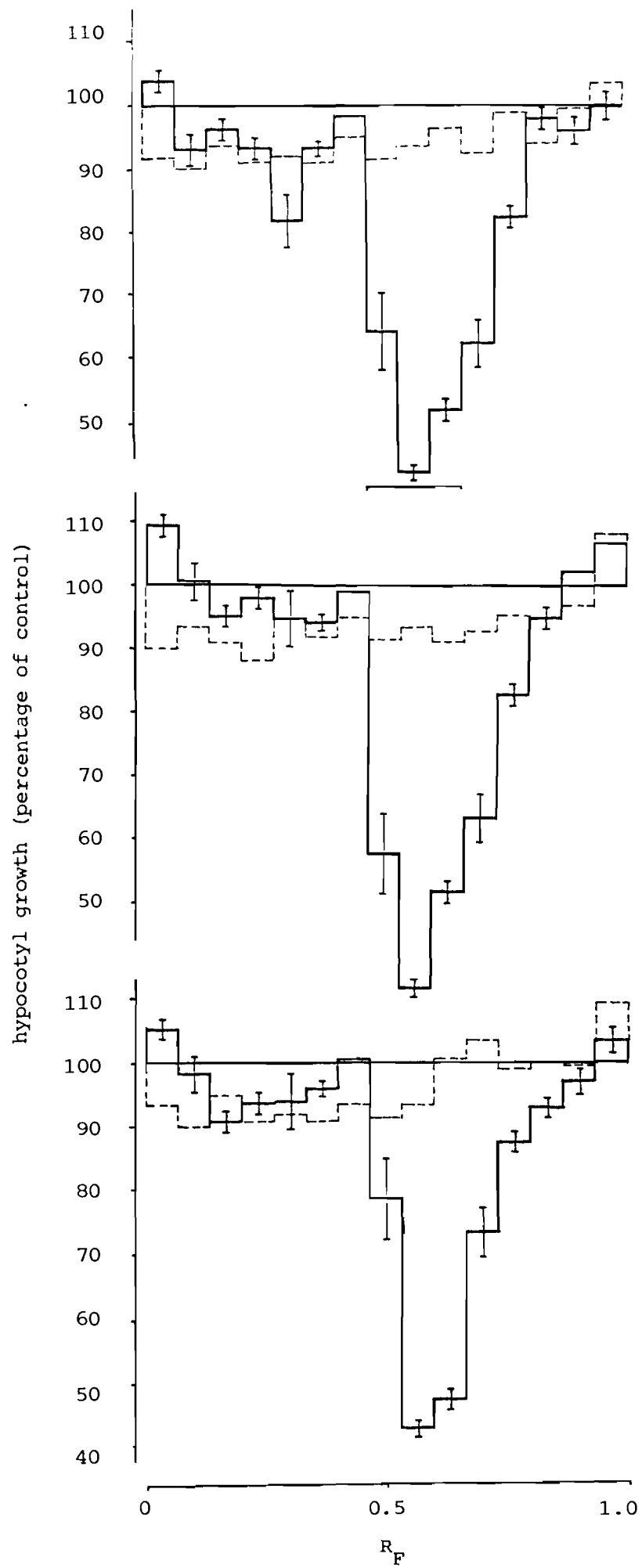


Figure 63. Lettuce hypocotyl assay of acidic ether-soluble extracts obtained from apices of *Alnus glutinosa* trees harvested on 2 May 1978 (H3). The equivalent of 0.20 g DW of apical material was assayed and the solvent system was isopropanol:NH₃:H₂O (10:1:1::v:v:v). Other details as in figure 26.

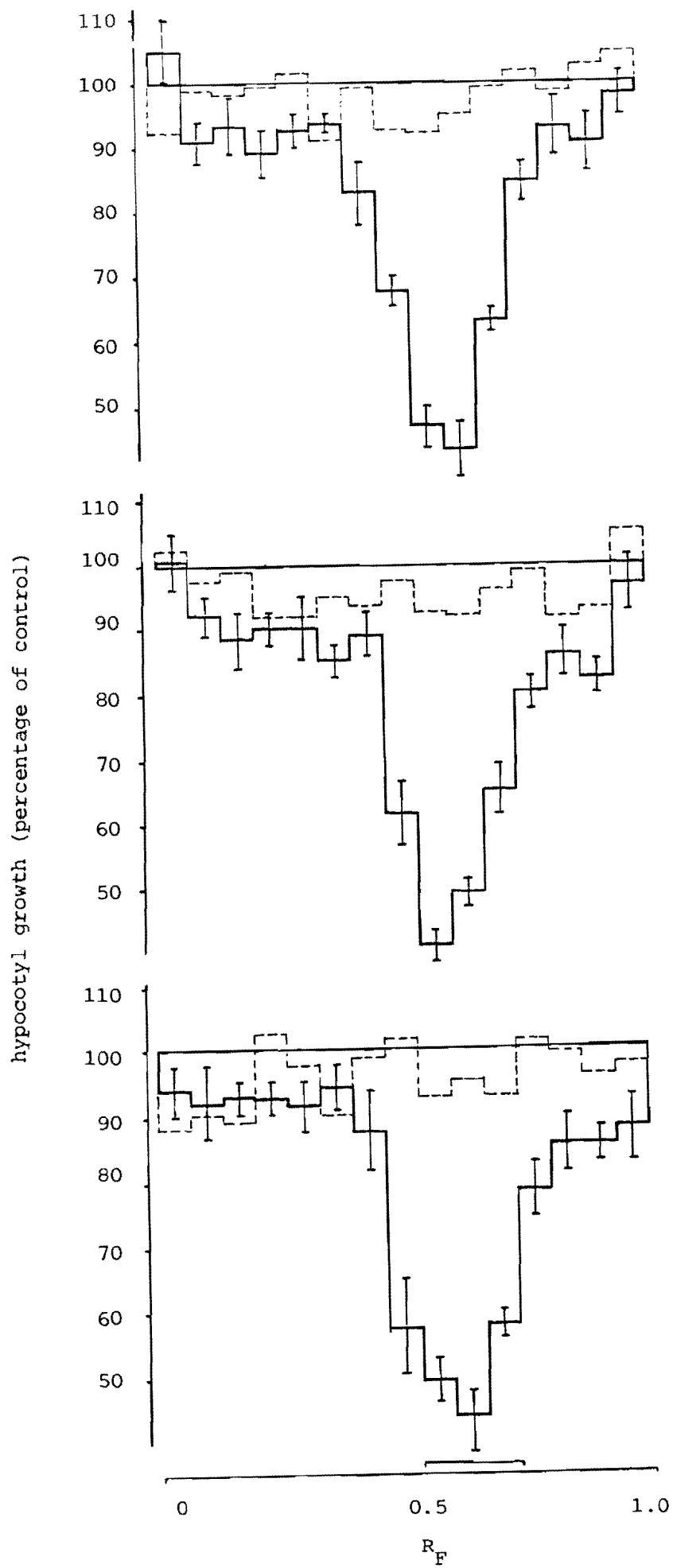


Figure 64. Wheat coleoptile section assay of the serial dilution of an acidic ether-soluble extract obtained from leaves of *Alnus glutinosa* trees harvested on 5 March 1979 (H1). The equivalent of (a) 0.025, (b) 0.125, (c) 0.25 g and (d) 0.30 g DW of leaf material was assayed. Other details as in figure 18.

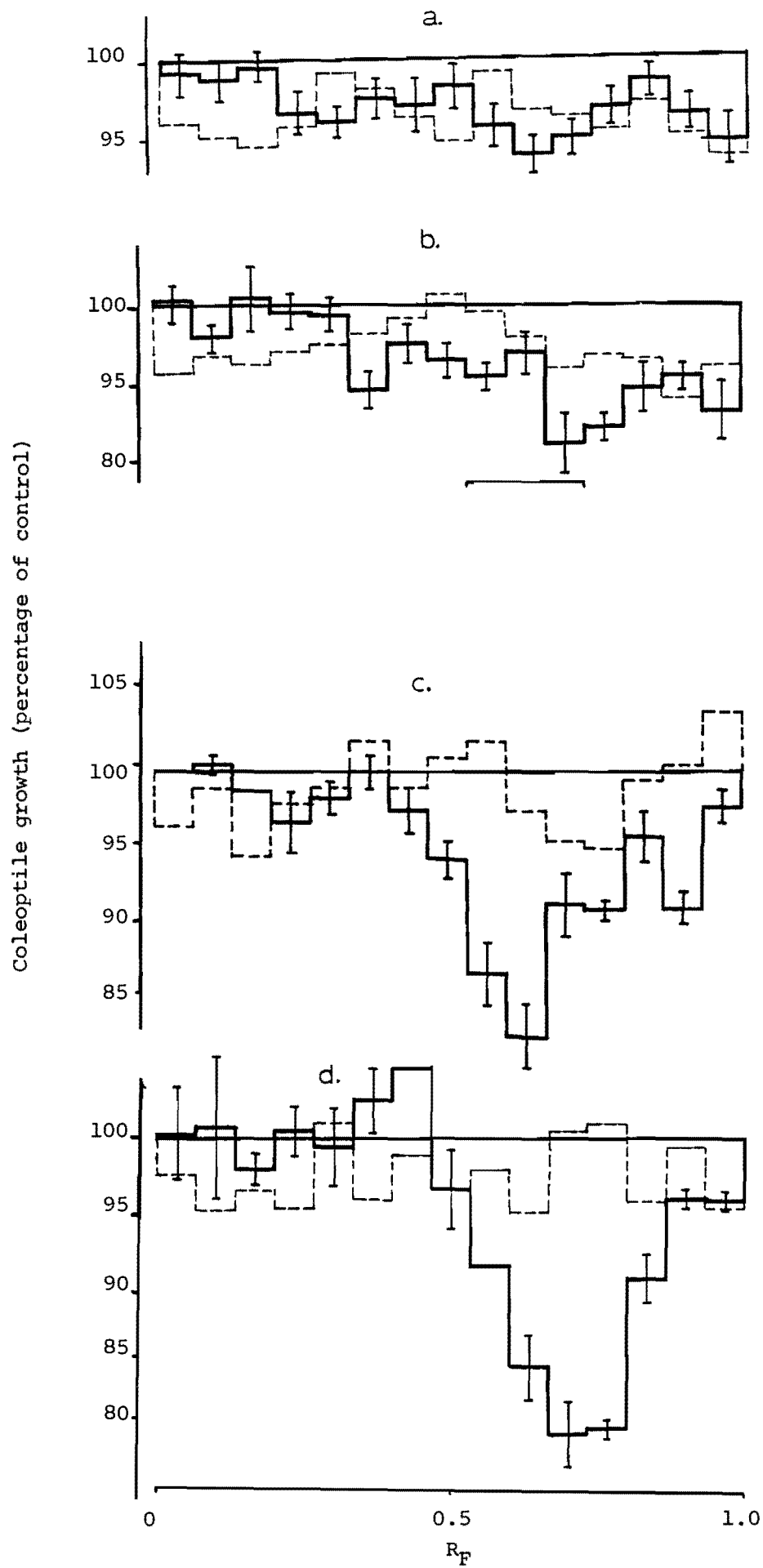


Figure 65. Wheat coleoptile section assay of the serial dilution of an acidic ether-soluble extract obtained from leaves of *Alnus glutinosa* trees harvested on 13 March 1979 (H3). The equivalent of (a) 0.025, (b) 0.125, (c) 0.25 g and (d) 0.30 g DW of leaf material was assayed. Other details as in figure 18.

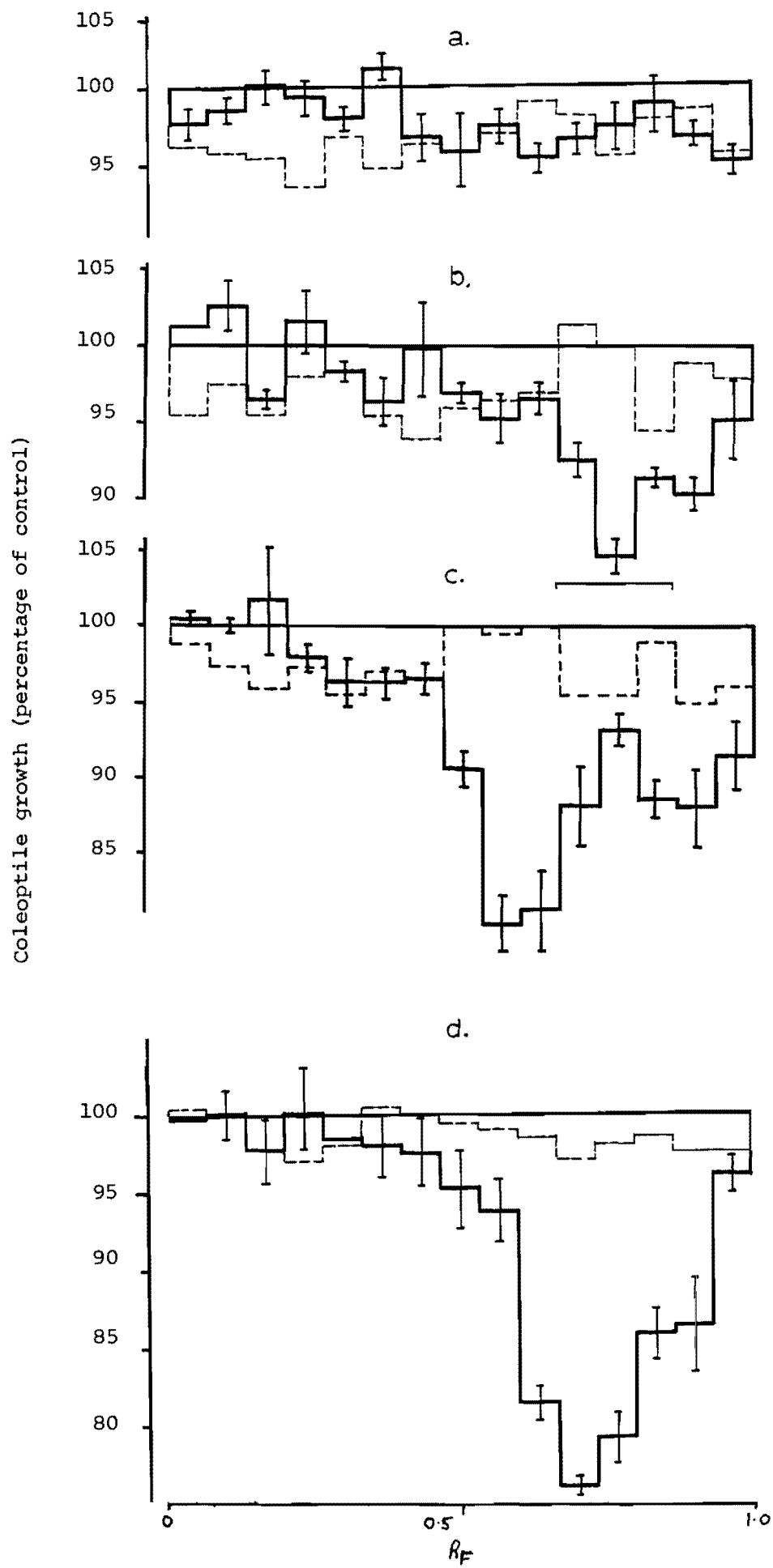


Figure 66. Lettuce hypocotyl assay of the serial dilution of an acidic ether-soluble extract obtained from leaves of *Alnus glutinosa* trees harvested on 5 March 1979 (H1). The equivalent of (a) 0.025, (b) 0.125, (c) 0.25 and (d) 0.30 g DW of leaf material was assayed. Other details as in figure 26.

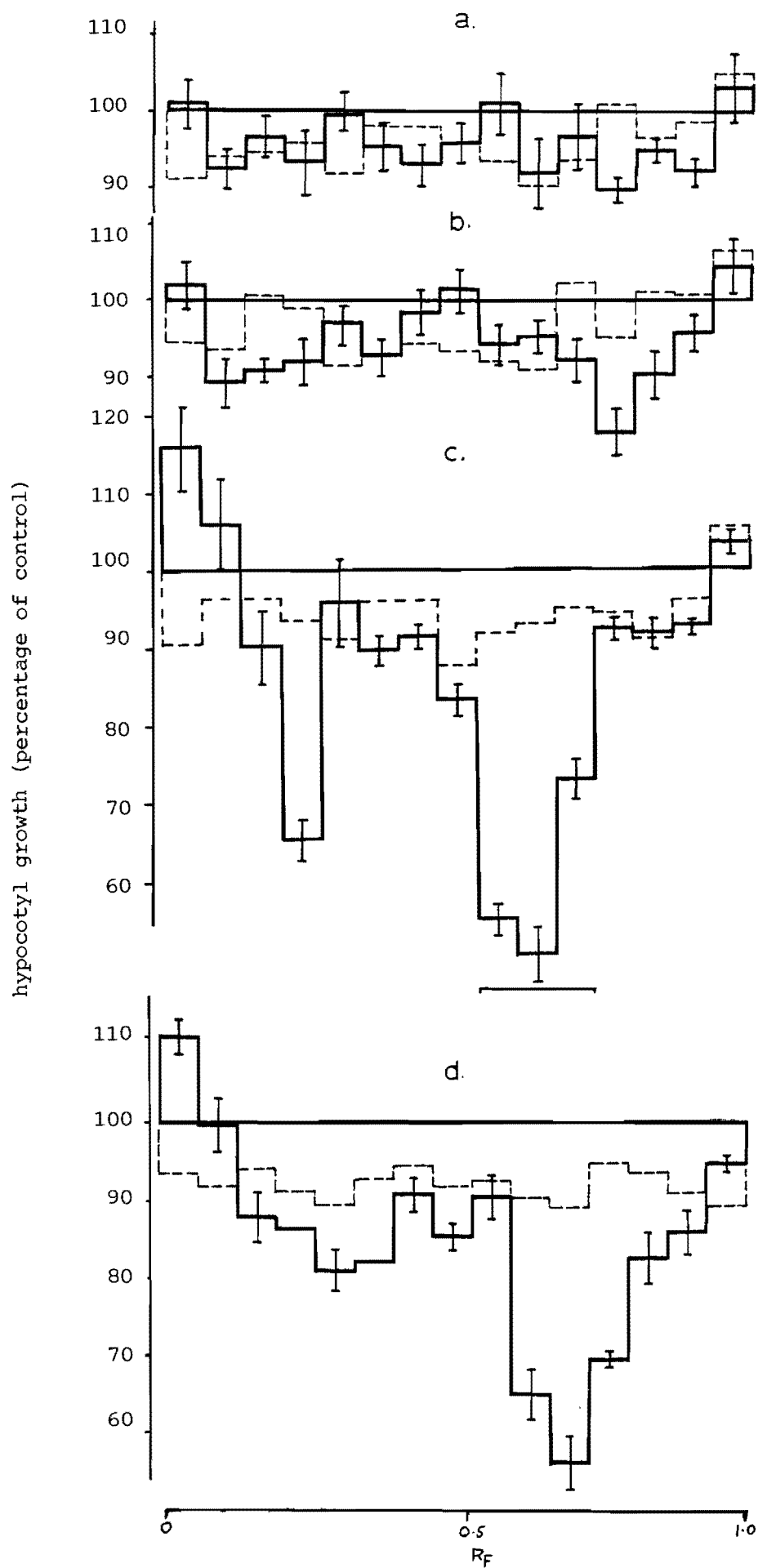


Figure 67. Lettuce hypocotyl assay of the serial dilution of an acidic ether-soluble extract obtained from leaves of *Alnus glutinosa* trees harvested on 4 April 1979 (H3). The equivalent of (a) 0.025, (b) 0.125, (c) 0.25 and (d) 0.30 g DW of apical material was assayed. Other details as in figure 26.

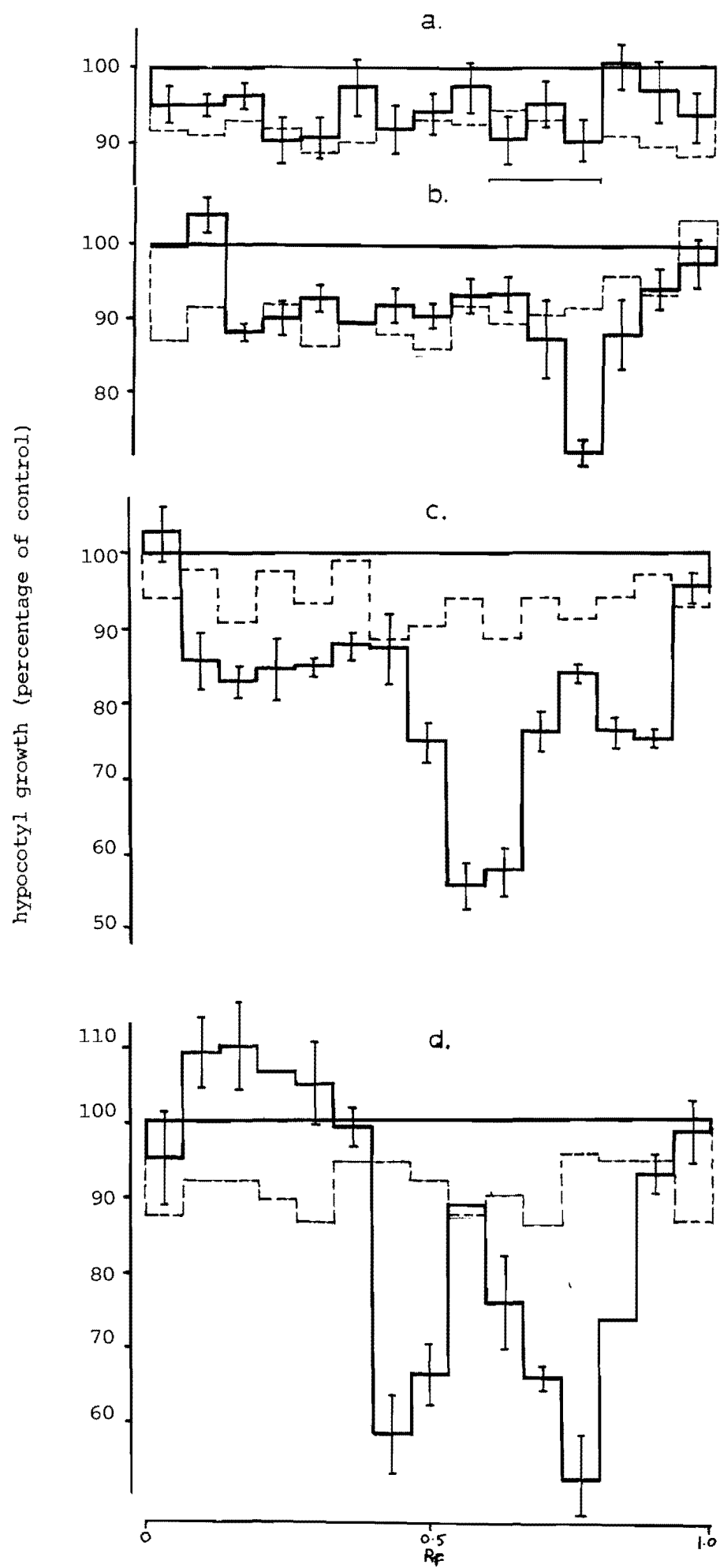
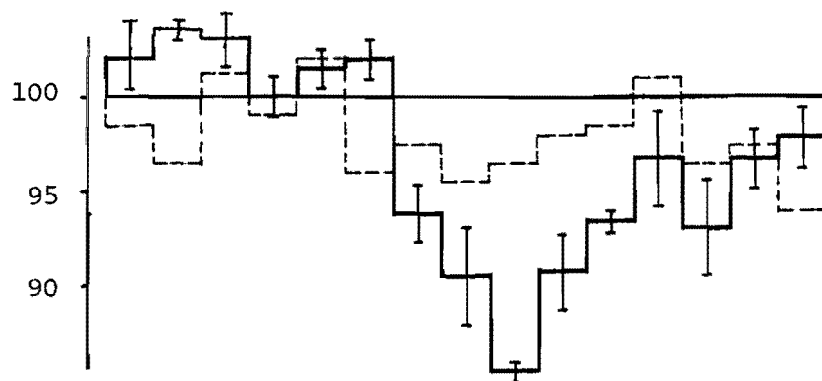


Figure 68. Wheat coleoptile section assay of acidic ether-soluble extracts obtained from leaves of *Alnus glutinosa* trees harvested on 5 March 1979 (H1). The equivalent of 0.25 g DW of leaf material was assayed and the solvent system was isopropanol:NH₃:H₂O (10:1:1::v:v:v). Other details as in figure 18.



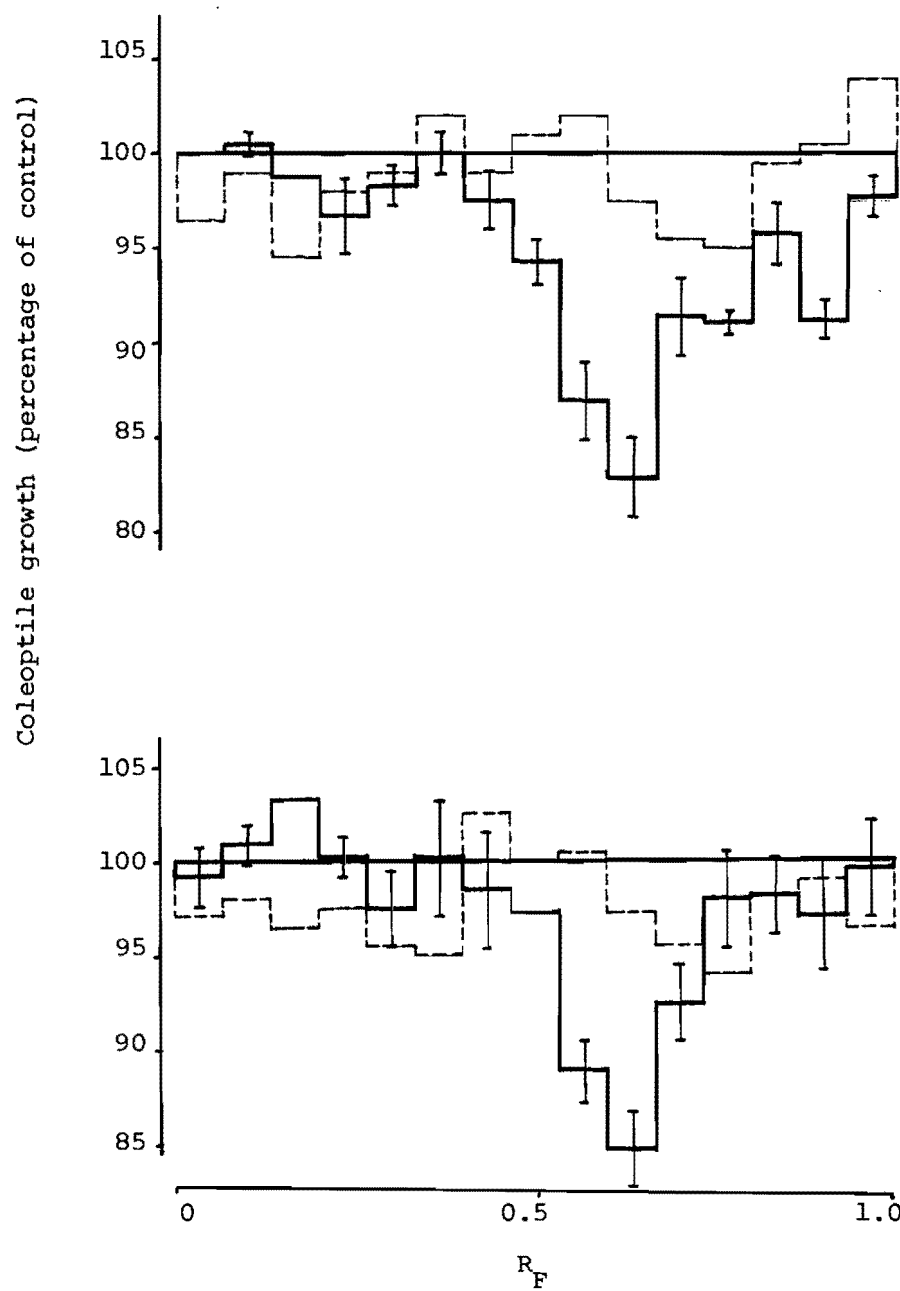


Figure 69. Wheat coleoptile section assay of acidic ether-soluble extracts obtained from leaves of *Alnus glutinosa* trees harvested on 5 March 1979 (H2). The equivalent of 0.25 g DW of leaf material was assayed and the solvent system was isopropanol:NH₃:H₂O (10:1:1::v:v:v). Other details as in figure 18.

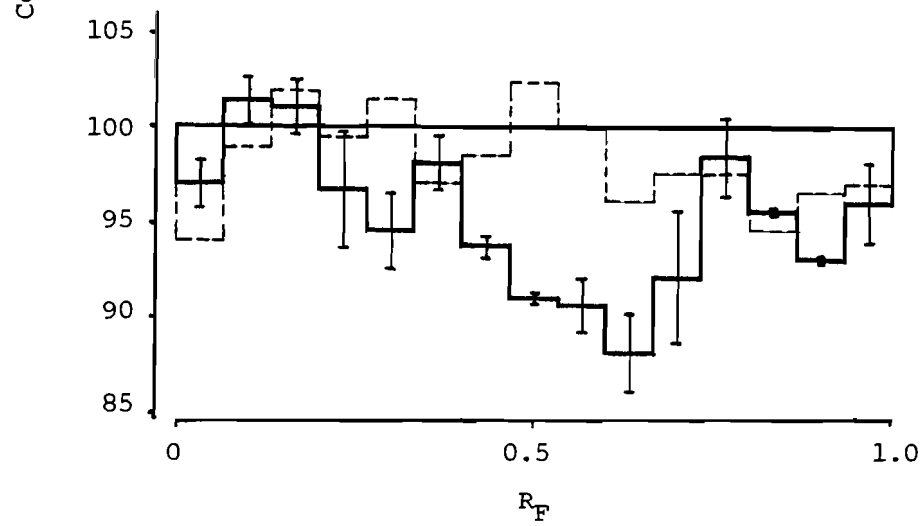
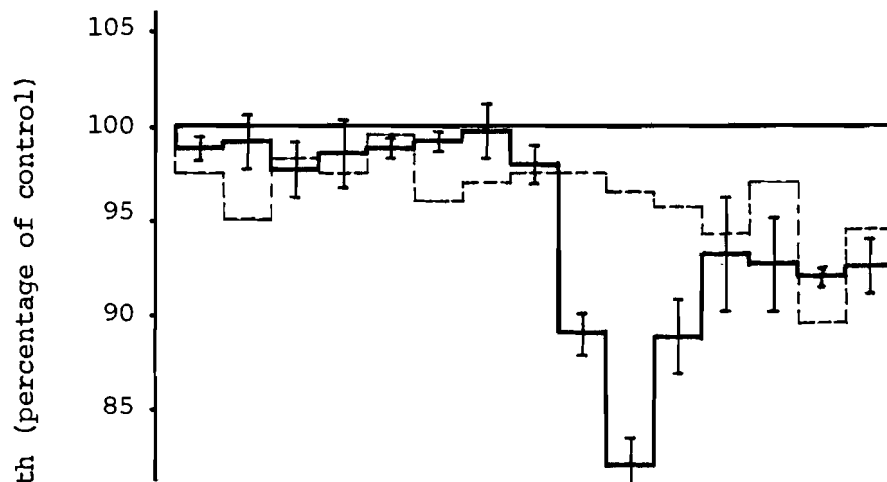
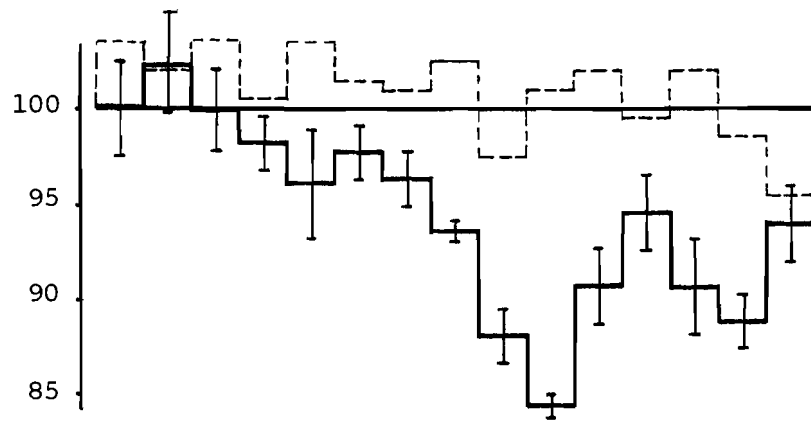


Figure 70. Wheat coleoptile section assay of acidic ether-soluble extracts obtained from leaves of *Alnus glutinosa* trees harvested on 4 April 1979 (H3). The equivalent of 0.25 g DW of leaf material was assayed and the solvent system was isopropanol:NH₃:H₂O (10:1:1::v:v:v). Other details as in figure 18.

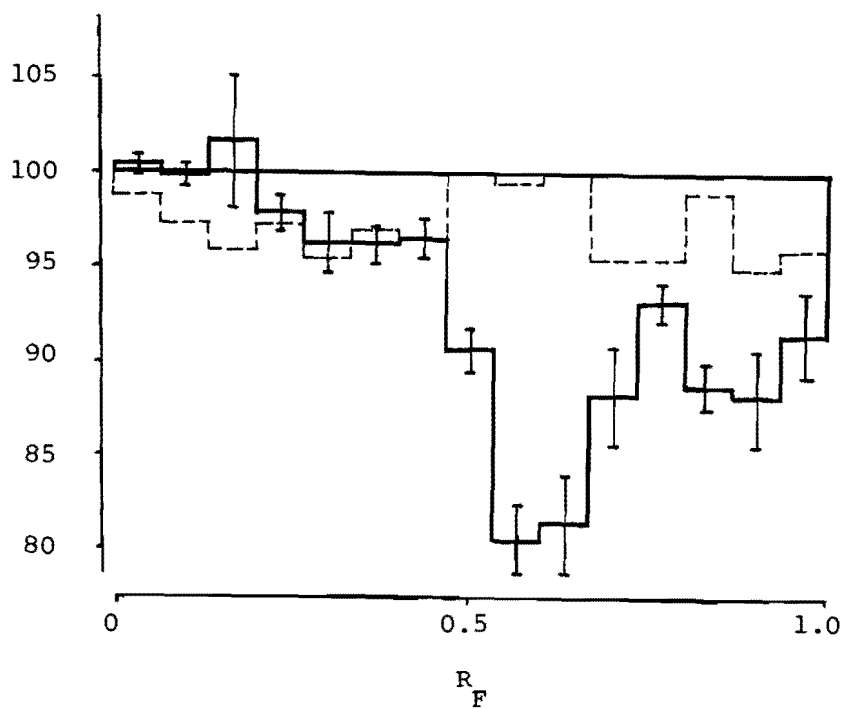
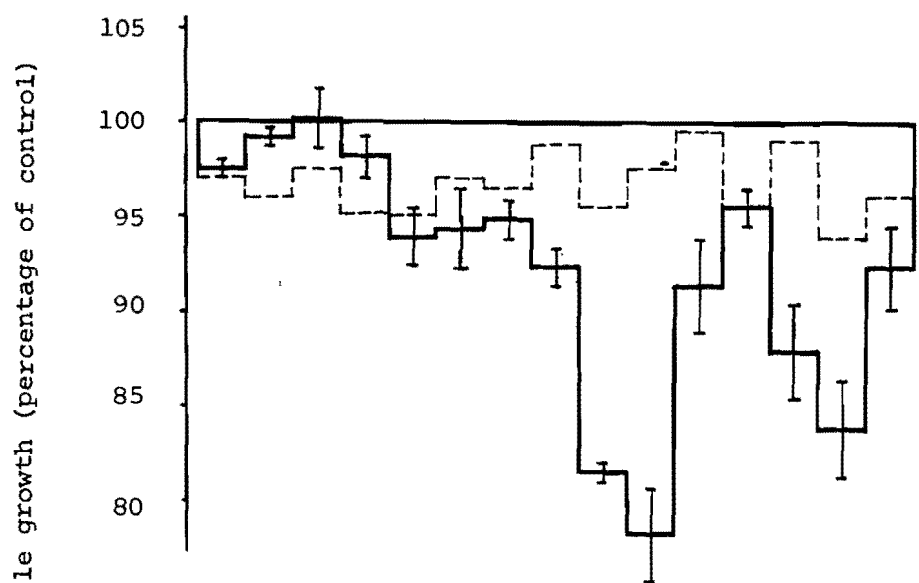
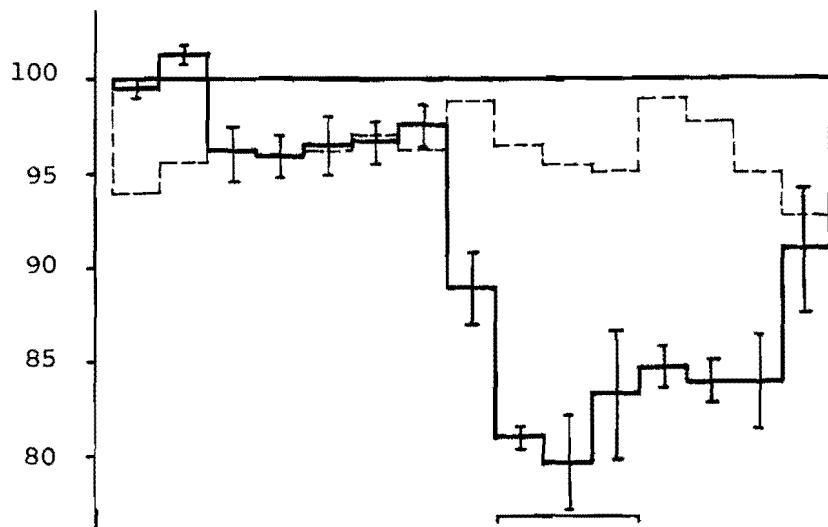


Figure 71. Lettuce hypocotyl assay of acidic ether-soluble extracts obtained from leaves of *Alnus glutinosa* trees harvested on 5 March 1979 (H1). The equivalent of 0.25 g DW of leaf material was assayed and the solvent system was isopropanol:NH₃:H₂O (10:1:1::v:v:v). Other details as in figure 26.

hypocotyl growth (percentage of control)

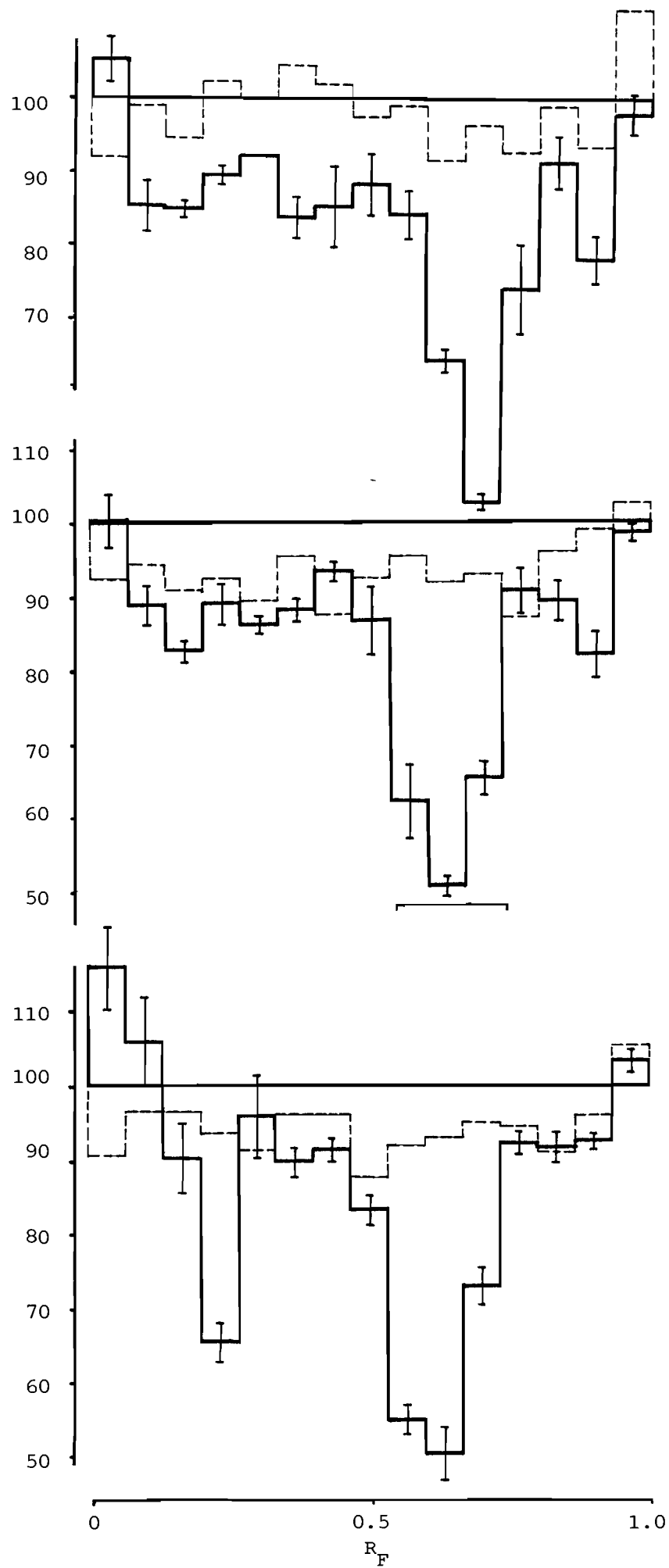


Figure 72. Lettuce hypocotyl assay of acidic ether-soluble extracts obtained from leaves of *Alnus glutinosa* trees harvested on 13 March 1979 (H2). The equivalent of 0.25 g DW of leaf material was assayed and the solvent system was isopropanol:NH₃:H₂O (10:1:1::v:v:v). Other details as in figure 26.

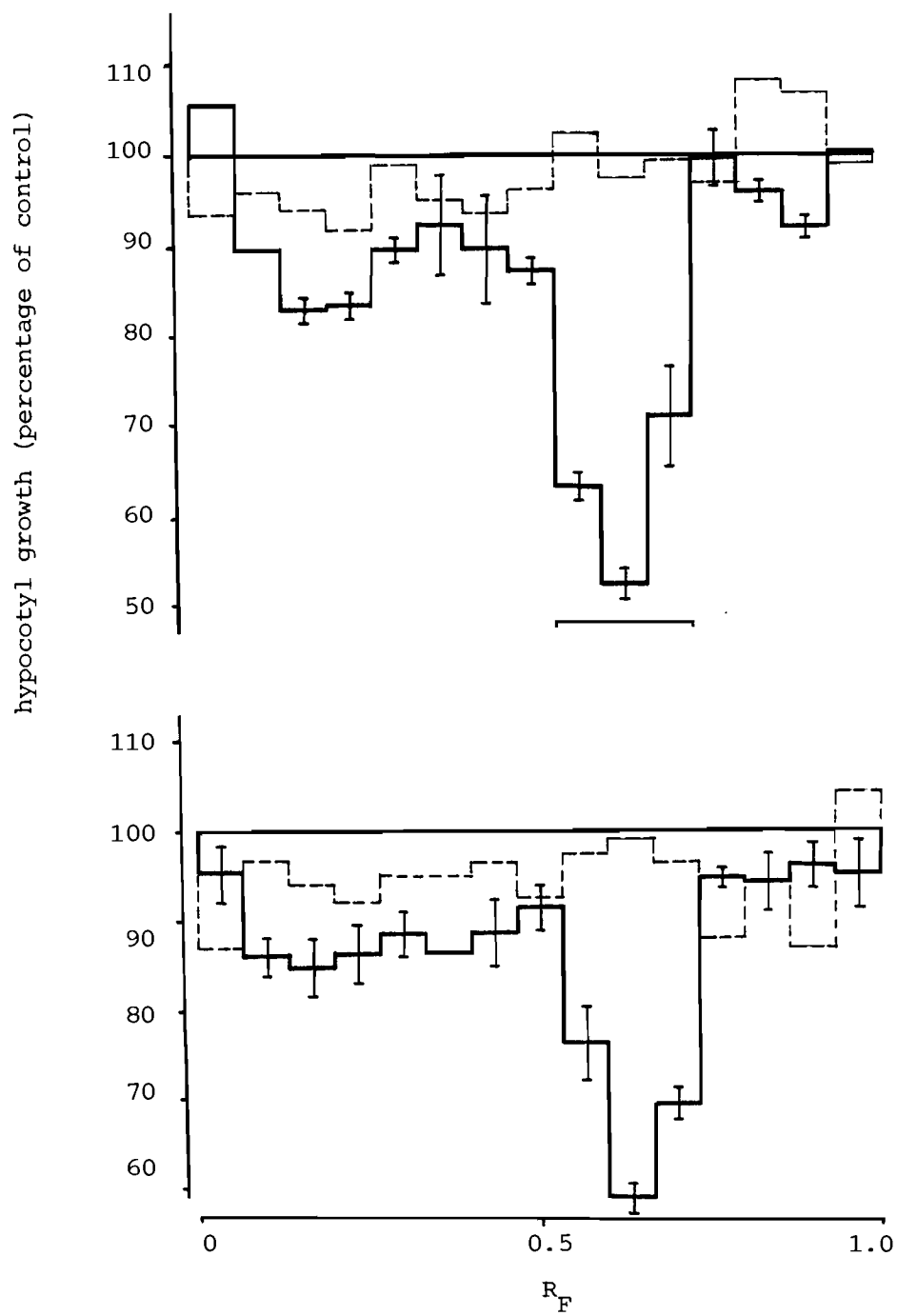
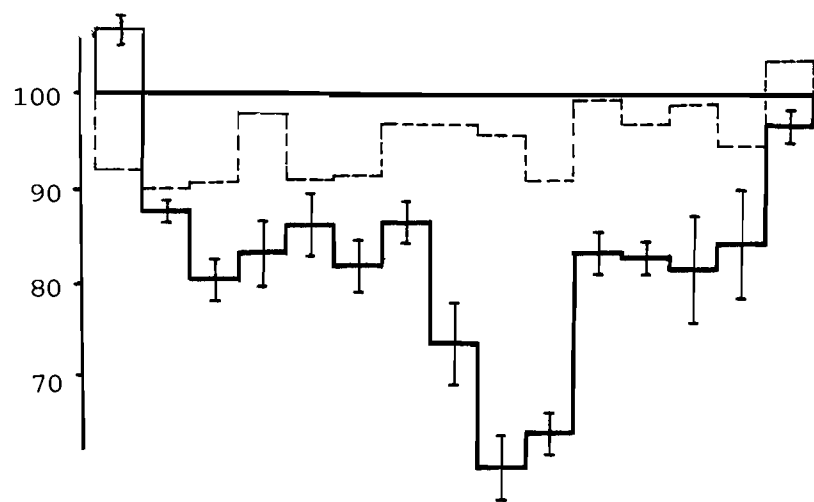


Figure 73. Lettuce hypocotyl assay of acidic ether-soluble extracts obtained from leaves of *Alnus glutinosa* trees harvested on 4 April 1979 (H3). The equivalent of 0.25 g DW of leaf material was assayed and the solvent system was isopropanol:NH₃:H₂O (10:1:1::v:v:v). Other details as in figure 26.

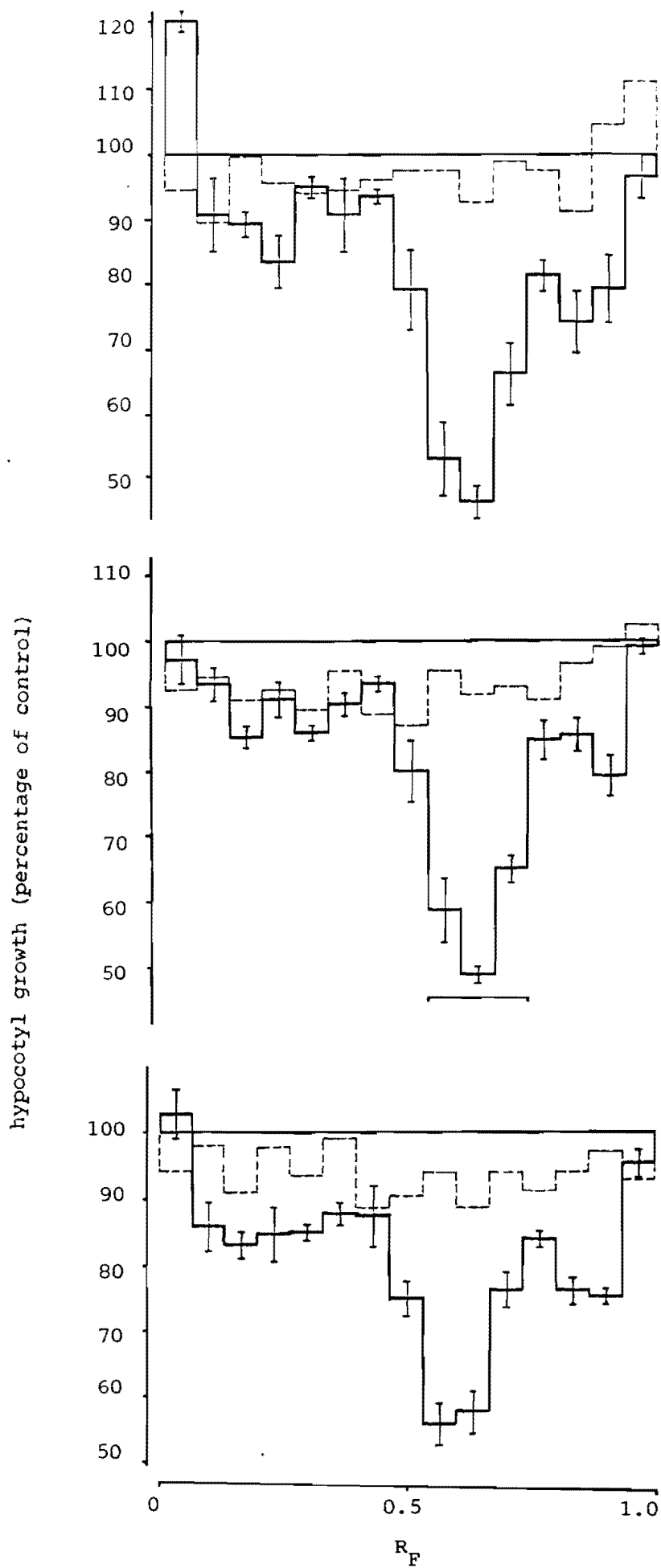


Figure 74. Wheat coleoptile section assay of acidic ether-soluble extracts obtained from apices of *Alnus glutinosa* trees harvested on 5 March 1979 (H1). The equivalent of 0.2 g DW of apical material was assayed and the solvent system was isopropanol:NH₃:H₂O (10:1:1::v:v:v). Other details as in figure 18.

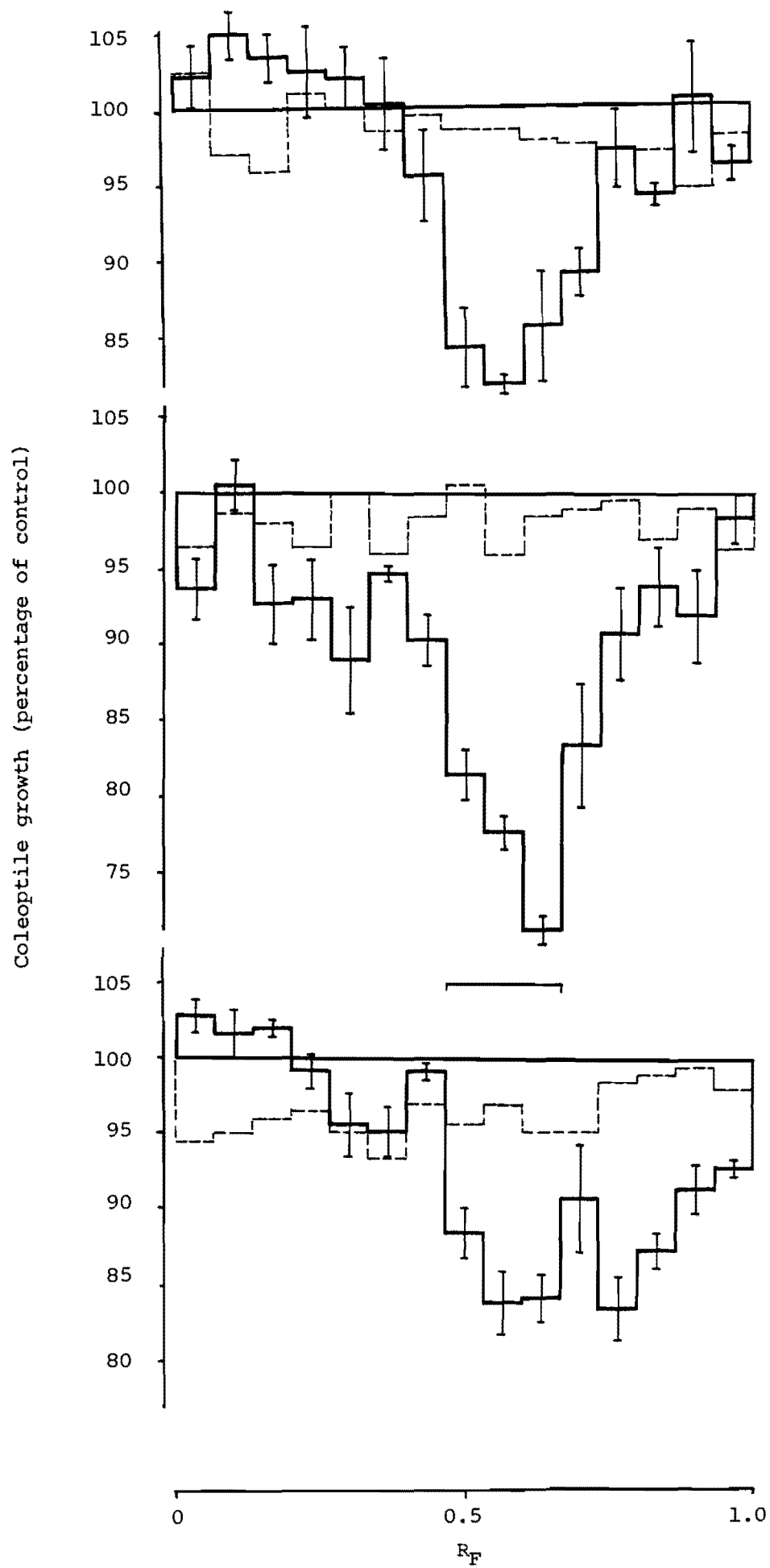


Figure 75. Wheat coleoptile section assay of acidic ether-soluble extracts obtained from apices of *Alnus glutinosa* trees harvested on 13 March 1979 (H2). The equivalent of 0.2 g DW of apical material was assayed and the solvent system was isopropanol:NH₃:H₂O (10:1:1::v:v:v). Other details as in figure 18.

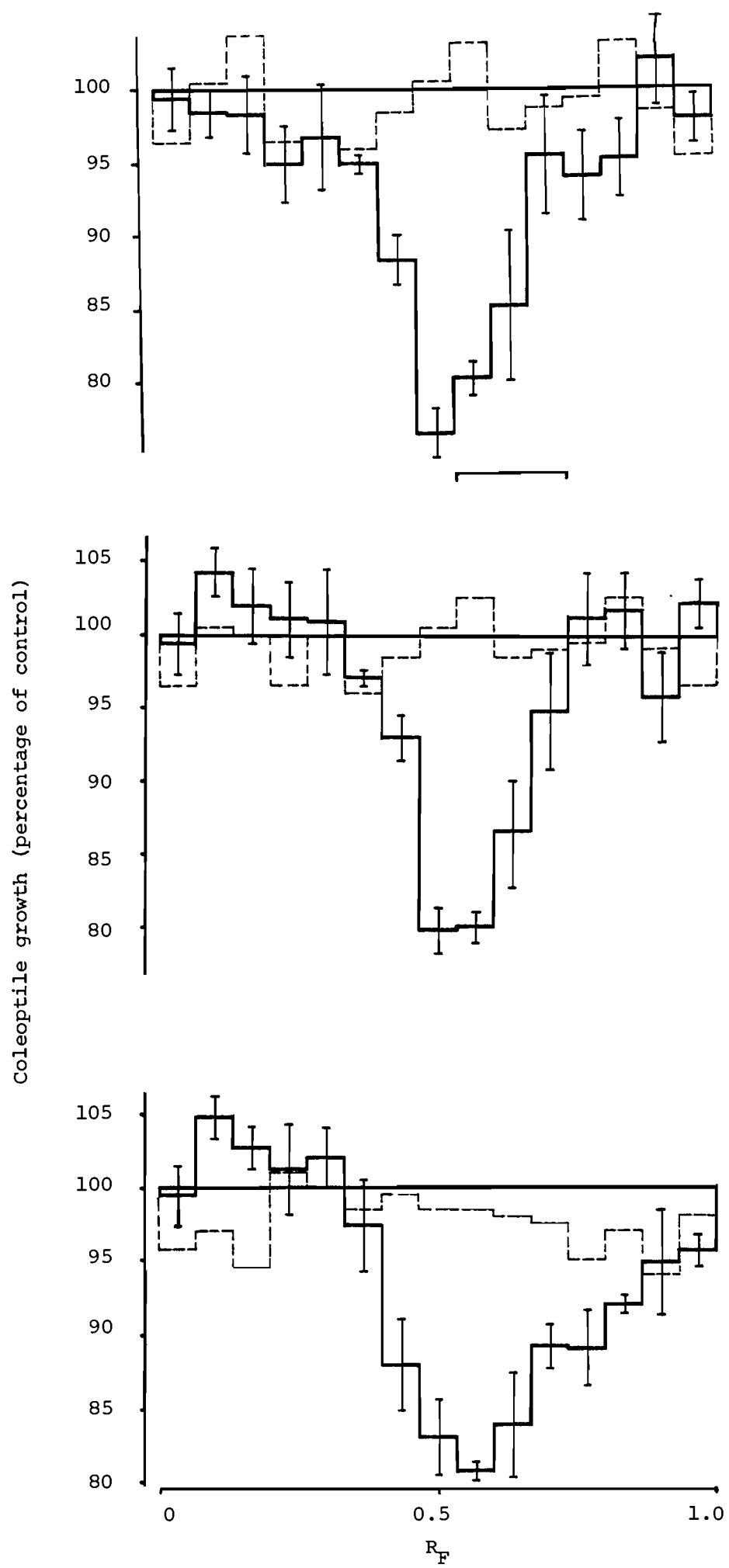


Figure 76. Wheat coleoptile section assay of acidic ether-soluble extracts obtained from apices of *Alnus glutinosa* trees harvested on 4 April 1979 (H3). The equivalent of 0.2 g DW of apical material was assayed and the solvent system was isopropanol:NH₃:H₂O (10:1:1::v:v:v). Other details as in figure 18.

Coleoptile growth (percentage of control)

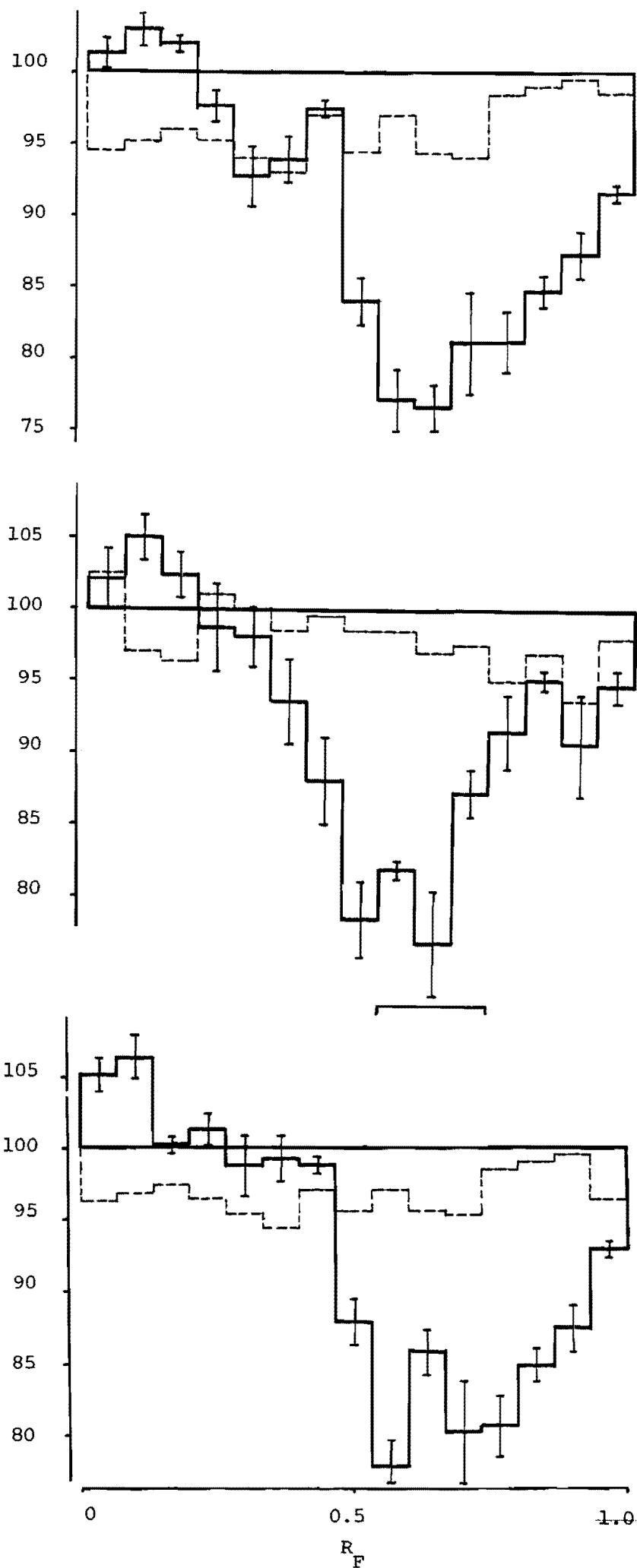


Figure 77. Lettuce hypocotyl assay of acidic ether-soluble extracts obtained from apices of *Alnus glutinosa* trees harvested on 5 March 1979 (H1). The equivalent of 0.2 g DW of apical material was assayed and the solvent system was isopropanol:NH₃:H₂O (10:1:1::v:v:v). Other details as in figure 26.

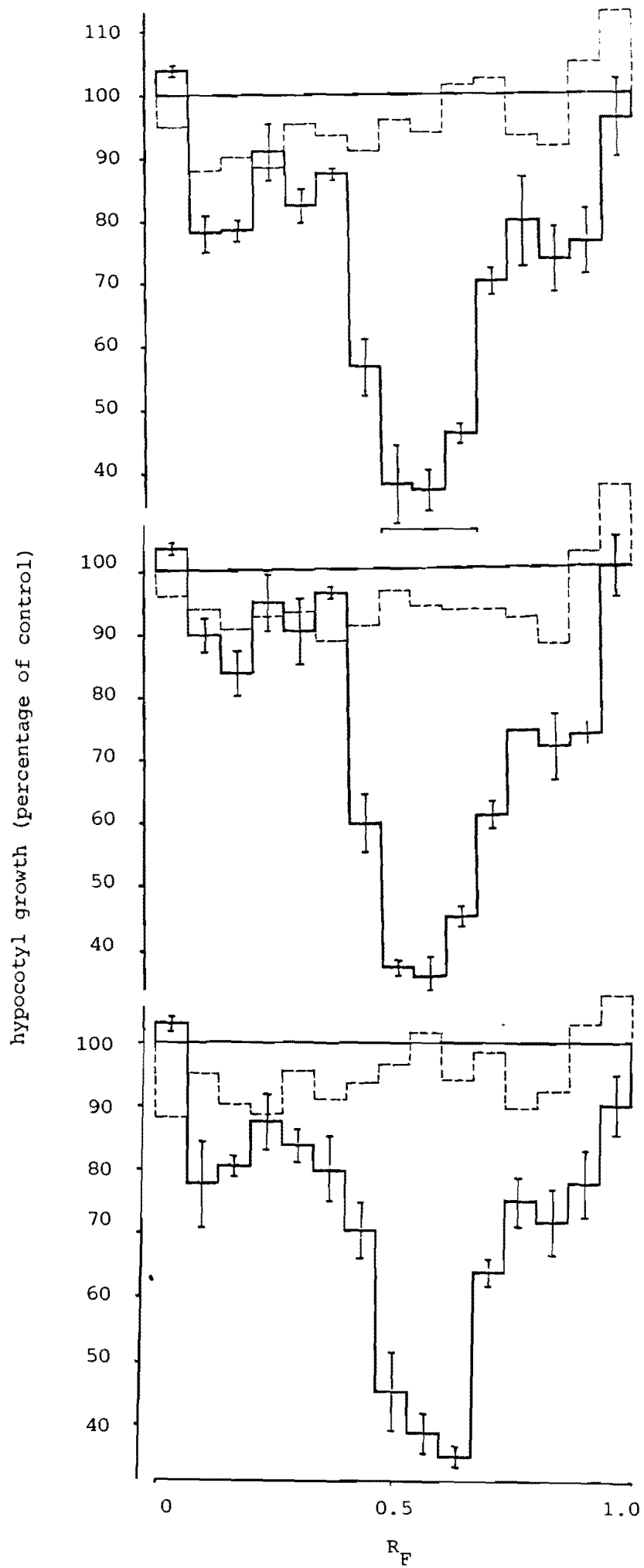


Figure 78. Lettuce hypocotyl assay of acidic ether-soluble extracts obtained from apices of *Alnus glutinosa* trees harvested on 13 March 1979 (H2). The equivalent of 0.2 g DW of apical material was assayed and the solvent system was isopropanol:NH₃:H₂O (10:1:1::v:v:v). Other details as in figure 26.

hypocotyl growth (percentage of control)

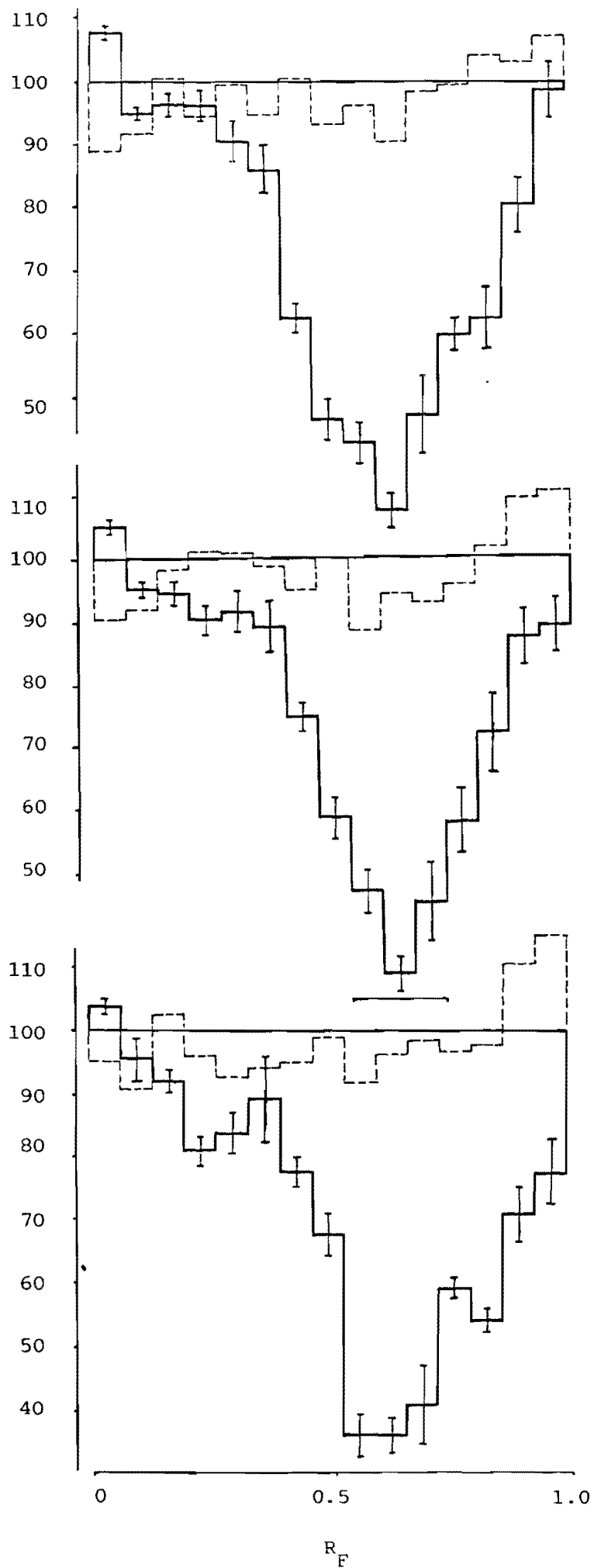


Figure 79. Lettuce hypocotyl assay of acidic ether-soluble extracts obtained from apices of *Alnus glutinosa* trees harvested on 4 April 1979 (H3). The equivalent of 0.2 g DW of apical material was assayed and the solvent system was isopropanol:NH₃:H₂O (10:1:1::v:v:v). Other details as in figure 26.

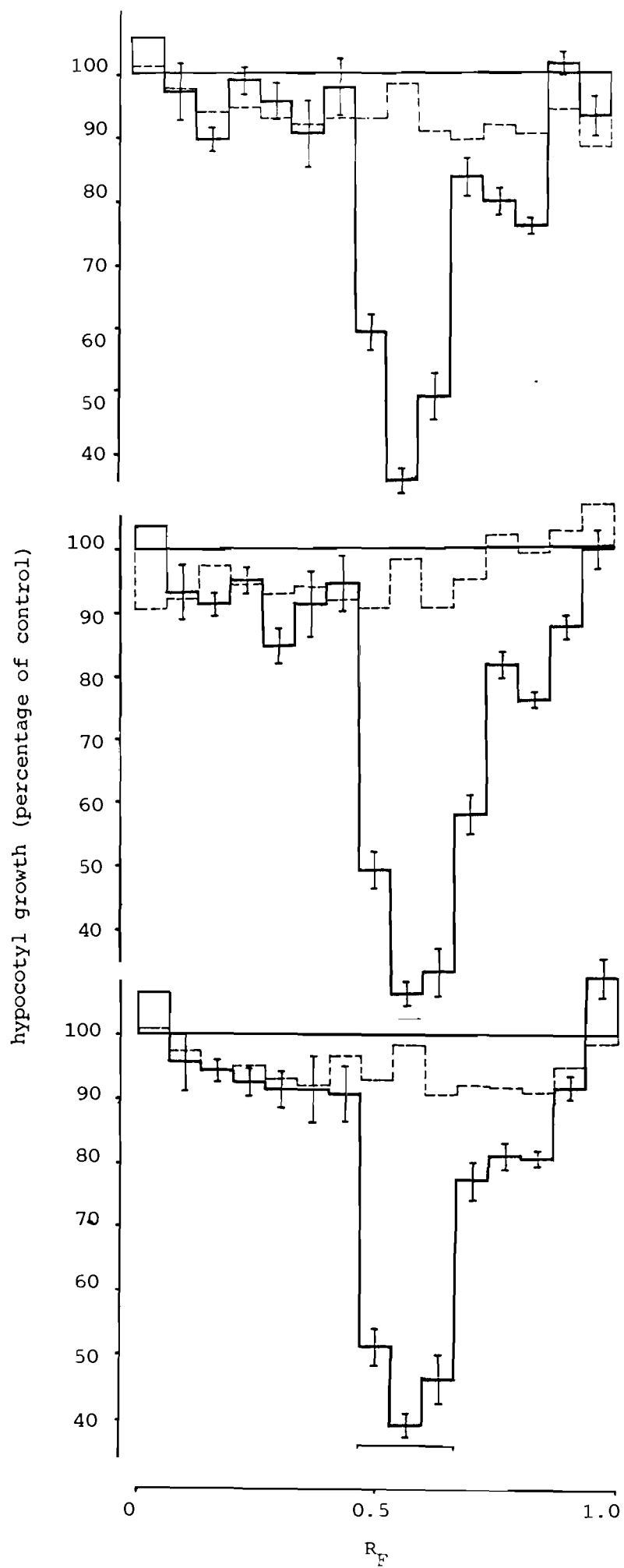


Figure 80. Wheat coleoptile section assay of the serial dilution of an acidic ether-soluble extract obtained from apices of *Alnus glutinosa* trees harvested on 5 March 1979 (H1). The equivalent of (a) 0.02, (b) 0.10, (c) 0.20 and (d) 0.35 g DW of apical material was assayed. Other details as in figure 18.

Coleoptile growth (percentage of control)

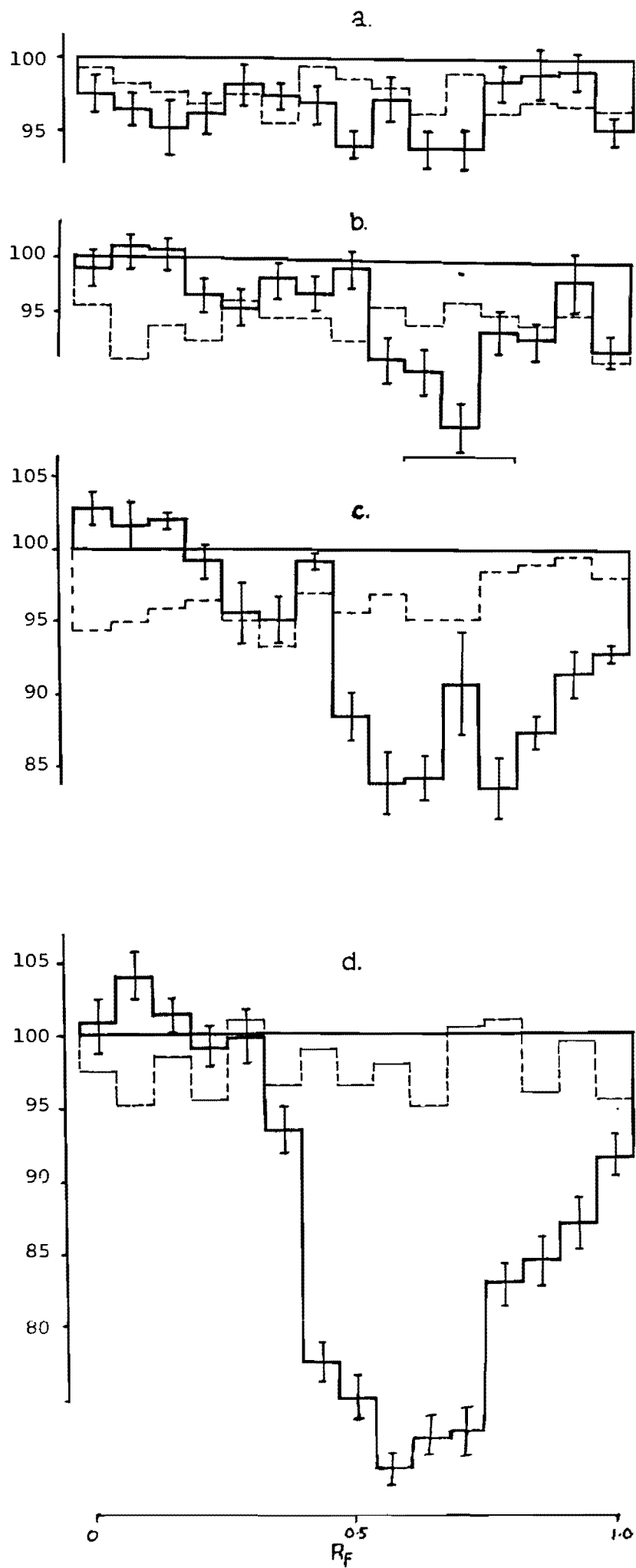


Figure 81. Wheat coleoptile section assay of the serial dilution of an acidic ether-soluble extract obtained from apices of *Alnus glutinosa* trees harvested on 4 April 1979 (H3). The equivalent of (a) 0.02, (b) 0.10, (c) 0.20 and (d) 0.35 g DW of apical material was assayed. Other details as in figure 18.

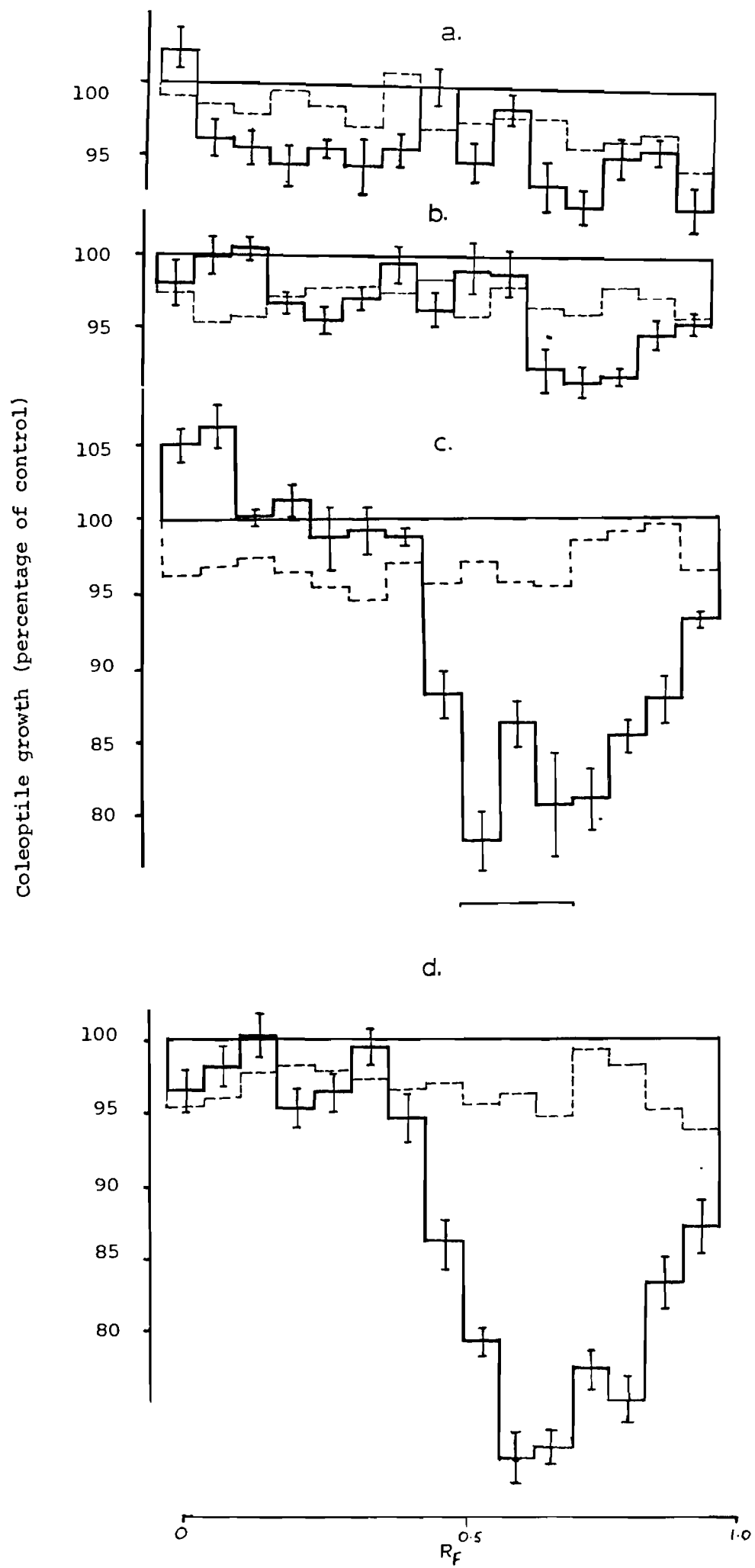


Figure 82. Lettuce hypocotyl assay of the serial dilution of an acidic ether-soluble extract obtained from apices of *Alnus glutinosa* trees harvested on 5 March 1979 (H1). The equivalent of (a) 0.02, (b) 0.10, (c) 0.20 and (d) 0.35 g DW of material was assayed. Other details as in figure 26.

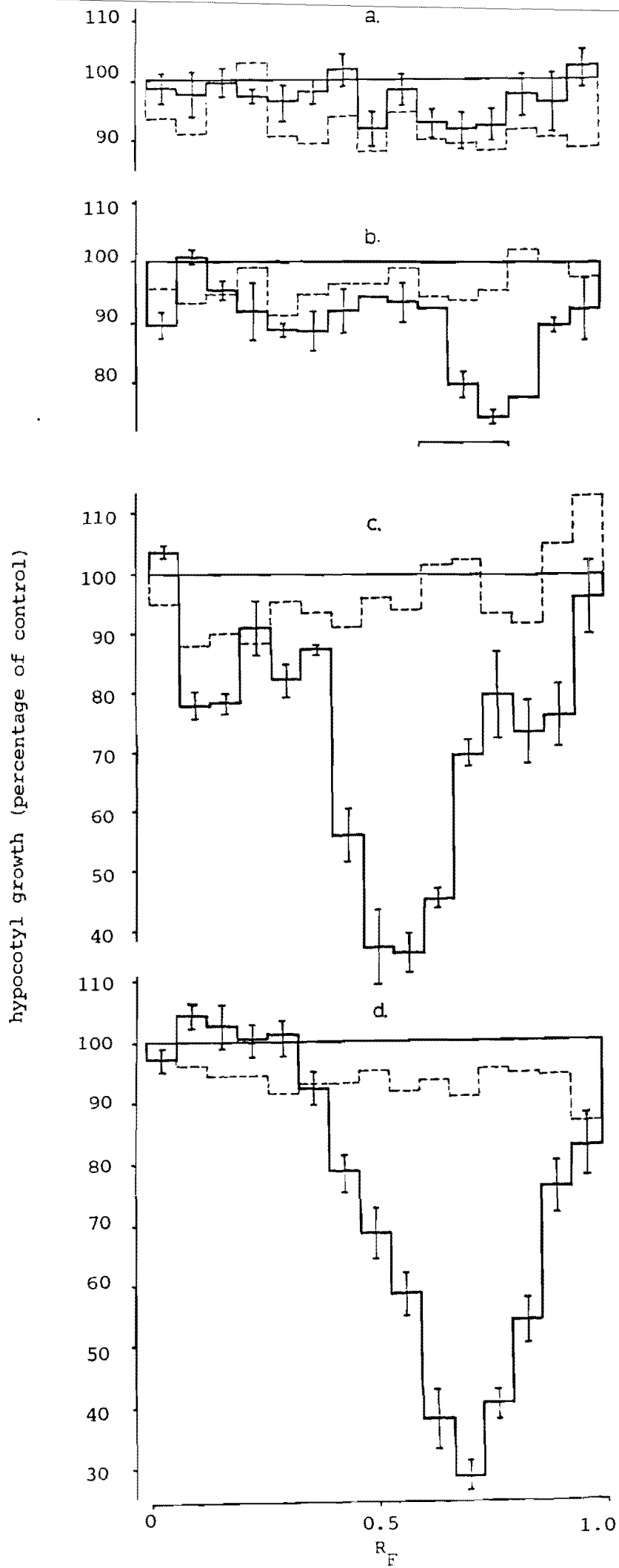


Figure 83. Lettuce hypocotyl assay of the serial dilution of an acidic ether-soluble extract obtained from apices of *Alnus glutinosa* trees harvested on 4 April 1979 (H3). The equivalent of (a) 0.02, (b) 0.10, (c) 0.20 and (d) 0.35 g DW of apical material was assayed. Other details as in figure 26.

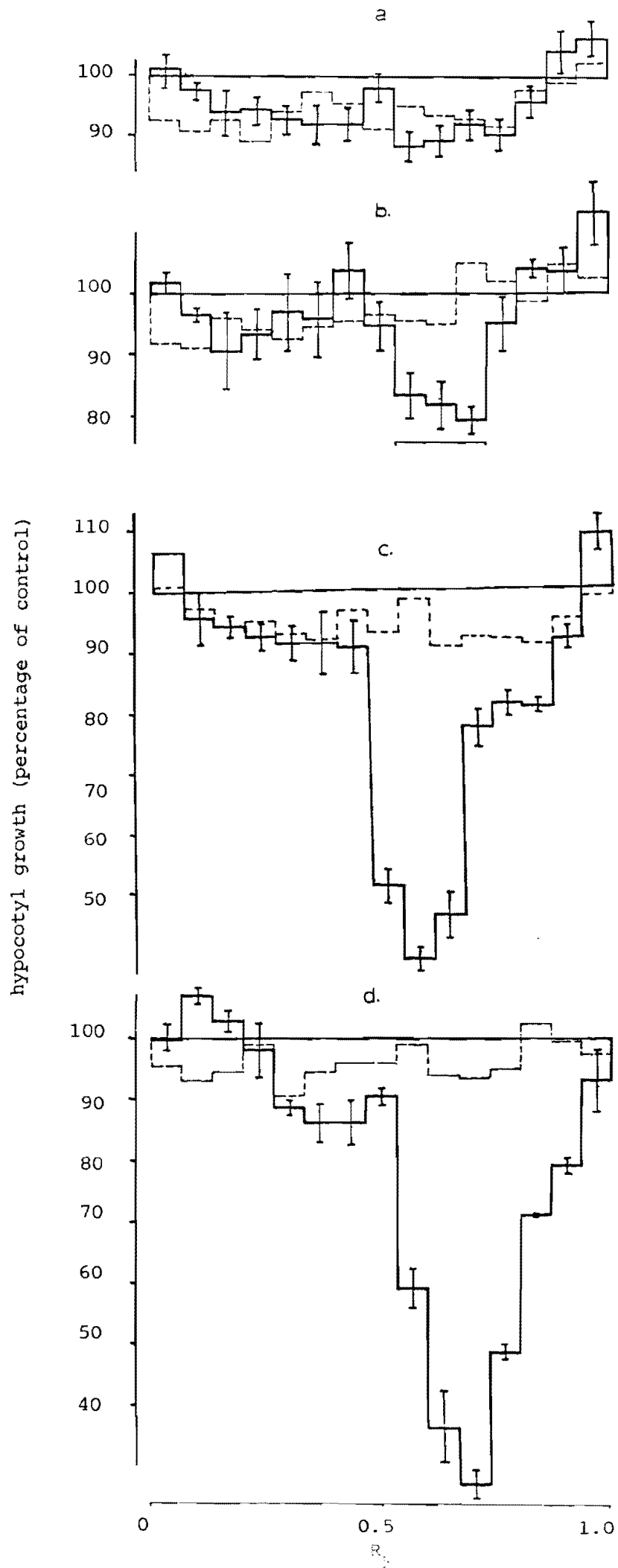


Figure 84. Diagram of a typical chromatograph of the inhibitor β fraction of *Alnus glutinosa* apices as seen under (A) U.V. light, and (B) visible light before and after spray treatment with phenolic reagents. The inhibitor β fraction was isolated from the equivalent of 0.2 g DW of tissue. The R_F zone 0.5 to 0.9 on paper chromatographs developed in isopropanol:NH₃:H₂O (10:1:1::v:v:v) was rechromatographed using n-butanol:NH (5:1::v:v) as solvent.

- (i) no spray treatment
- (ii) treatment with diazotised sulphanilic acid
- (iii) treatment with diazotised p-nitroaniline
- (iv) treatment with 0.2% FeCl₃ (w:v in water)

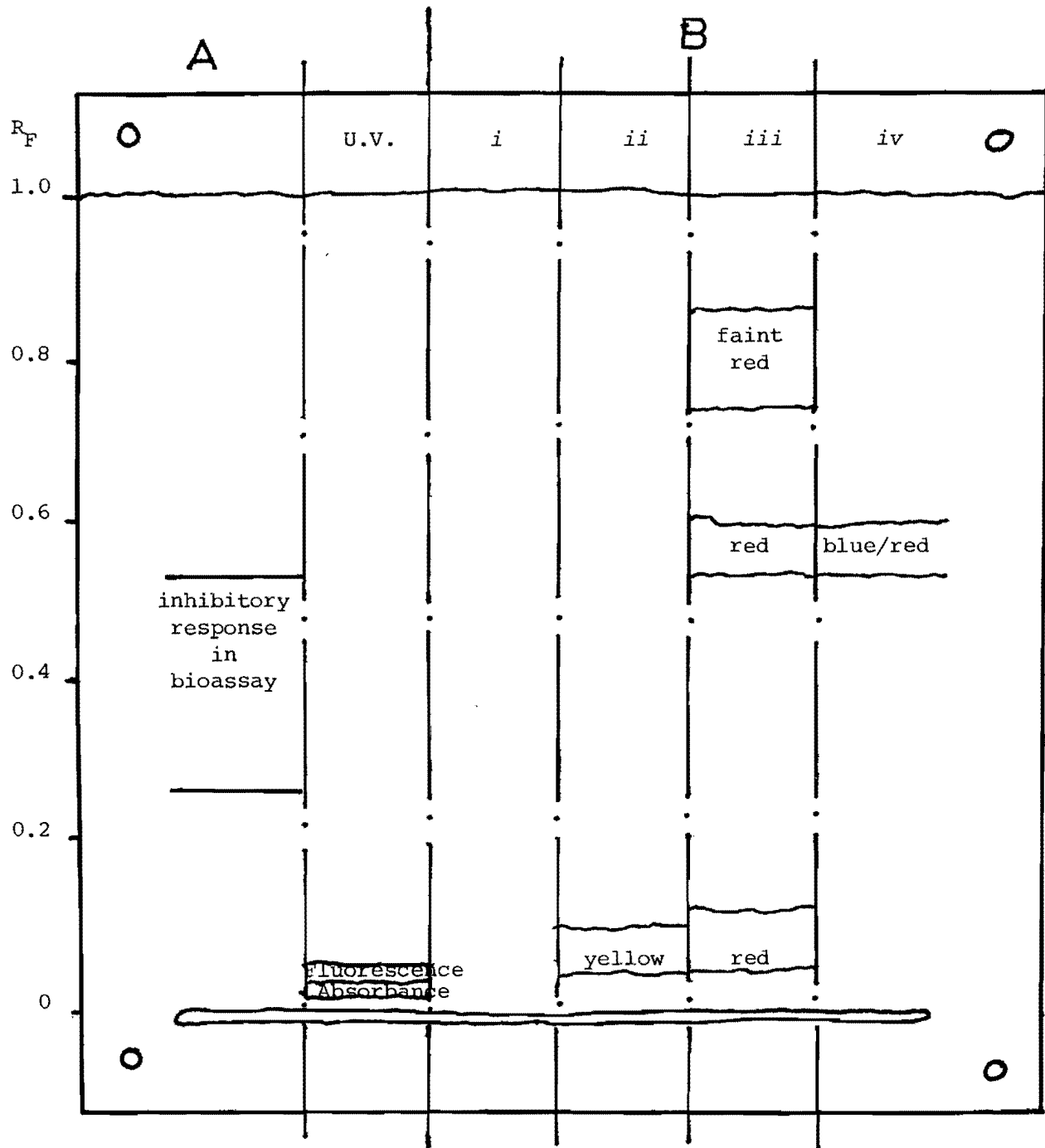


Figure 85. Wheat coleoptile section assay of the inhibitor β fraction of *Alnus glutinosa* apices harvested on (A) 13 March 1978 (H1) and (B) 2 May 1978 (H3). The inhibitor β fraction was isolated from the acidic ether-soluble fraction by paper chromatography with isopropanol:NH₃:H₂O (10:1:1::v:v:v) solvent. The zone R_F 0.5 to 0.9 was rechromatographed using n-butanol:NH₃ (5:1::v:v) as solvent. The equivalent to 0.25 g DW of apical material was assayed. The solid horizontal bar indicates the R_F of authentic ABA and the broken horizontal bars indicate the positions of phenolic-like substances. Other details as in figure 18.

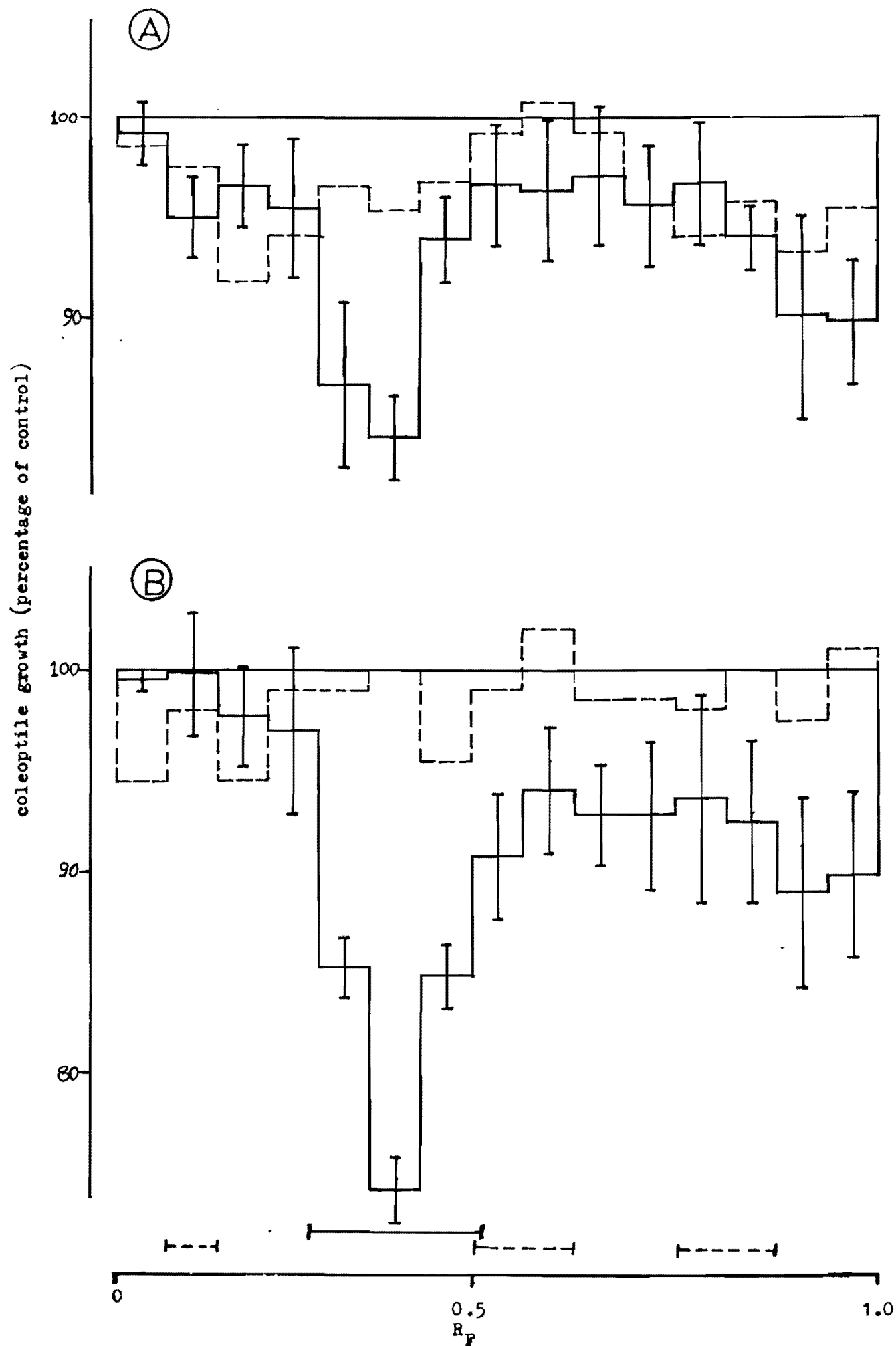


Figure 86. Lettuce hypocotyl assay of the inhibitor β fraction of *Alnus glutinosa* apices harvested on (A) 13 March 1978 (H1) and (B) 2 May 1978 (H3). The inhibitor β fraction was isolated from the acidic ether-soluble fraction by paper chromatography with isopropanol:NH₃:H₂O (10:1:1::v:v:v) solvent. The zone R_F 0.5 to 0.9 was rechromatographed using n-butanol:NH₃ (5:1::v:v) as solvent. The equivalent to 0.25 g DW of apical material was assayed. The solid horizontal bar indicates the R_F of authentic ABA and the broken horizontal bars indicate positions of phenolic-like substances. Other details as in figure 26.

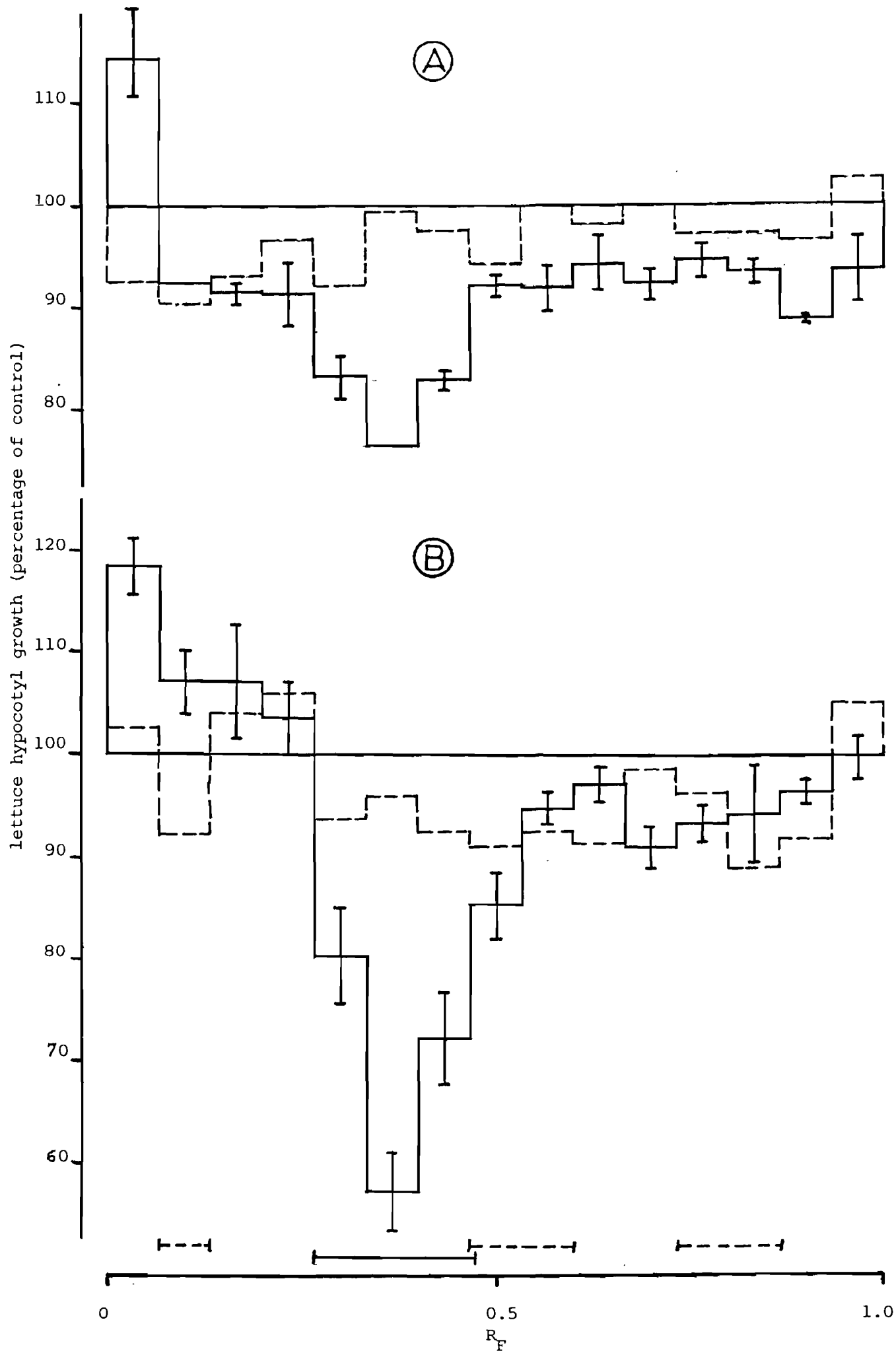


Figure 87. U.V. spectrum of (A) the main inhibitory component(s) present in acidic ether-soluble extracts of dormant *Alnus glutinosa* apices, and (B) the mixed isomers of authentic ABA. The apices were harvested on (a) 13 March 1978 (H1) and (b) 2 May 1978 (H3).

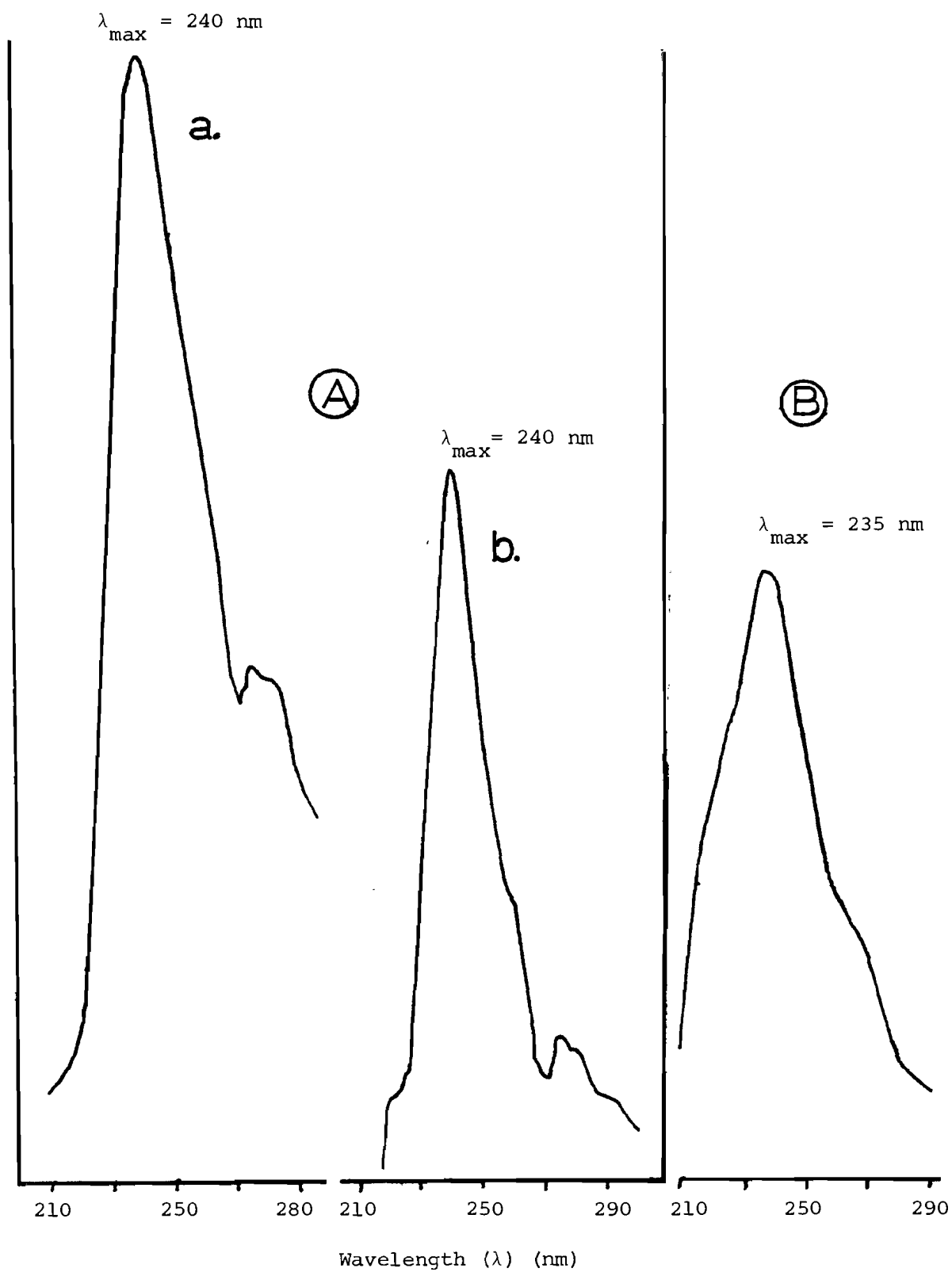


Figure 88. Gas chromatographs of the main inhibitory component(s) present in acidic ether-soluble extracts of dormant *Alnus glutinosa* apices. The chromatographs were obtained using an electron capture detector and 3% OV 17 as the stationary phase. Column and detector temperatures were 195°C and 295°C, respectively. Samples were methylated prior to chromatography. The apices were harvested on (A) 13 March 1978 (H1) and (B) 2 May 1978 (H3), and the inhibitory component(s) was isolated from the inhibitor β fraction by paper chromatography. Arrows indicate peaks corresponding with cis,trans Me-ABA and trans,trans Me-ABA.

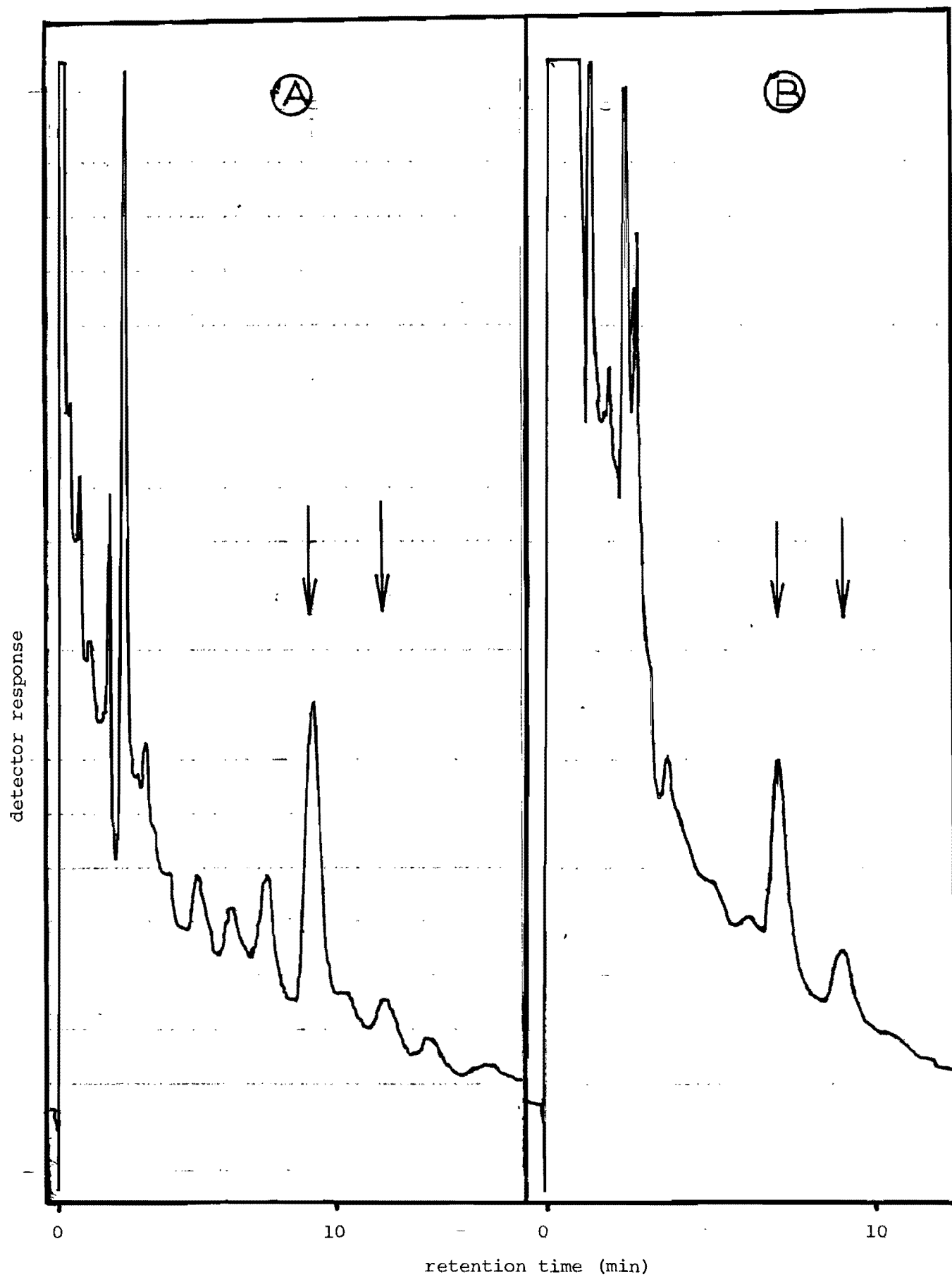


Figure 89. Growth of *Alnus viridis* seedlings under 16 h (LD) and then transfer to 8 h (SD) photoperiods. Plants were maintained at 10°C night and 14°C day (equivalent to the high light intensity period). Arrow indicates the start of the SD treatment. Each value represents the percentage increase in height and is the mean of 10 replicates (LD's) or 7 replicates to day 7 and 4 replicates after day 15 of SD treatment. Each replicate consists of two plants and the vertical bars represent twice the S.E.'s of the means.

Leaves and apices were harvested from three replicates on day 0, day 7 and day 15 of SD treatment.

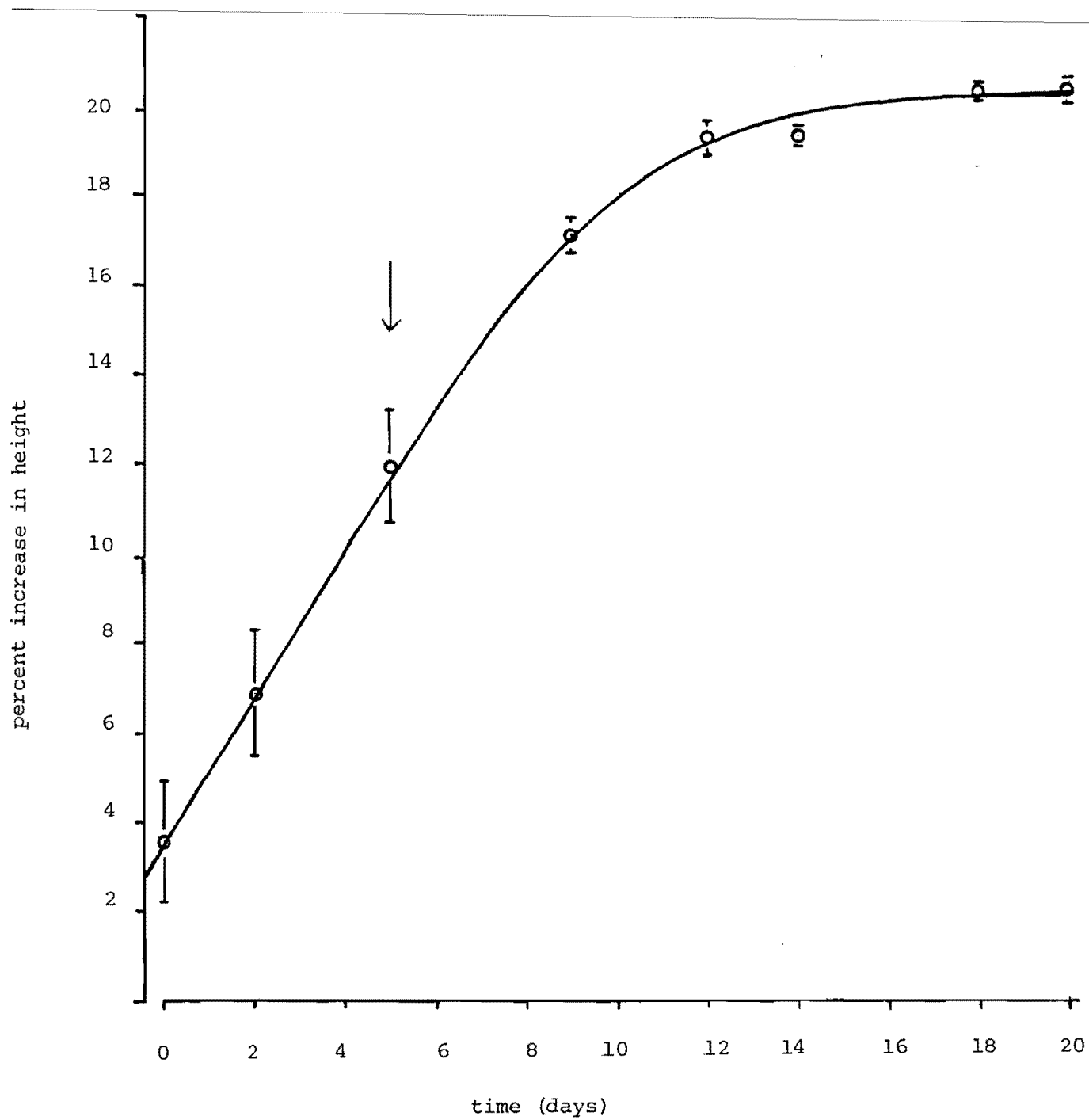


Figure 90. Wheat coleoptile section assay of acidic ether-soluble extracts obtained from leaves of *Alnus viridis* seedlings harvested on day 0 of SD treatment (H1). The equivalent of 0.25 g DW of leaf material was assayed and the solvent system was isopropanol:NH₃:H₂O (10:1:1::v:v:v). Other details as in figure 18.

Coleoptile growth (percentage of control)

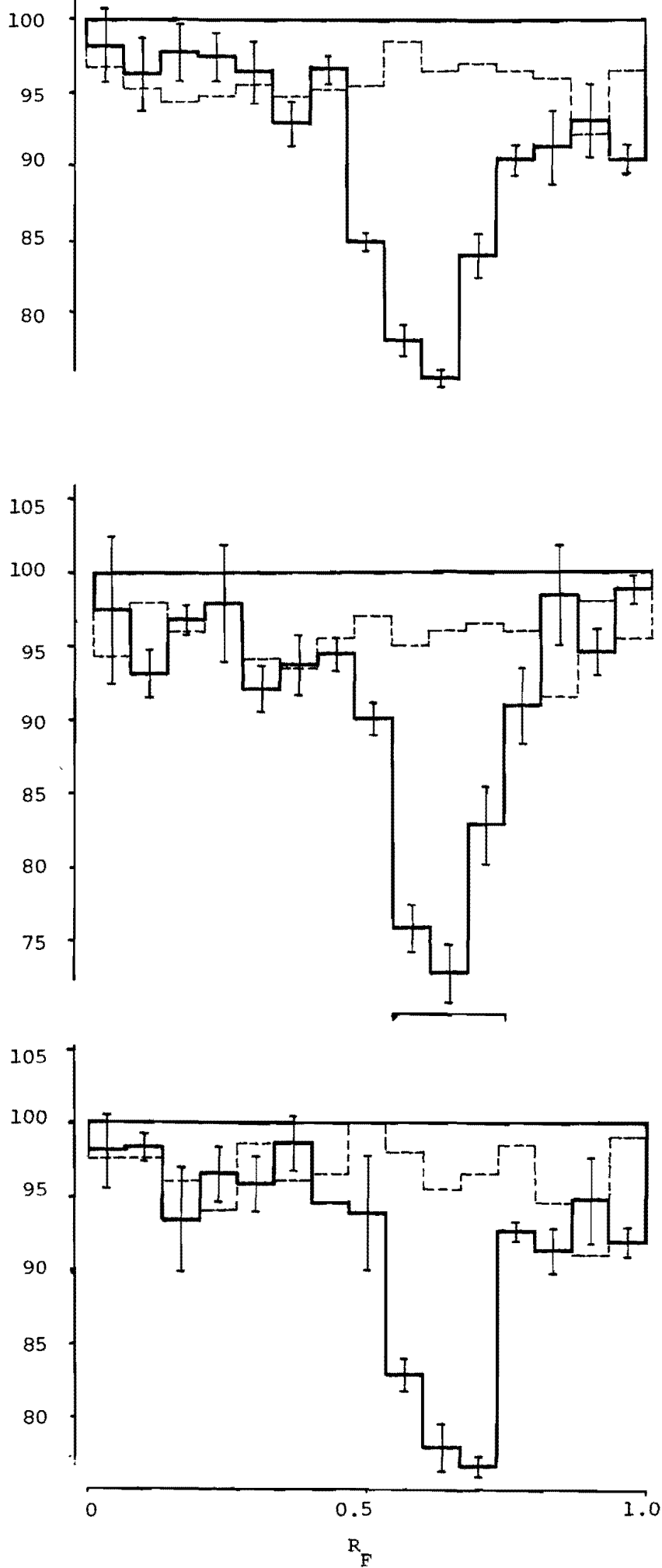


Figure 91. Wheat coleoptile section assay of acidic ether-soluble extracts obtained from leaves of *Alnus viridis* seedlings harvested on day 7 of SD treatment (H2). The equivalent of 0.25 g DW of leaf material was assayed and the solvent system was isopropanol:NH₃:H₂O (10:1:1::v:v:v). Other details as in figure 18.

Coleoptile growth (percentage of control)

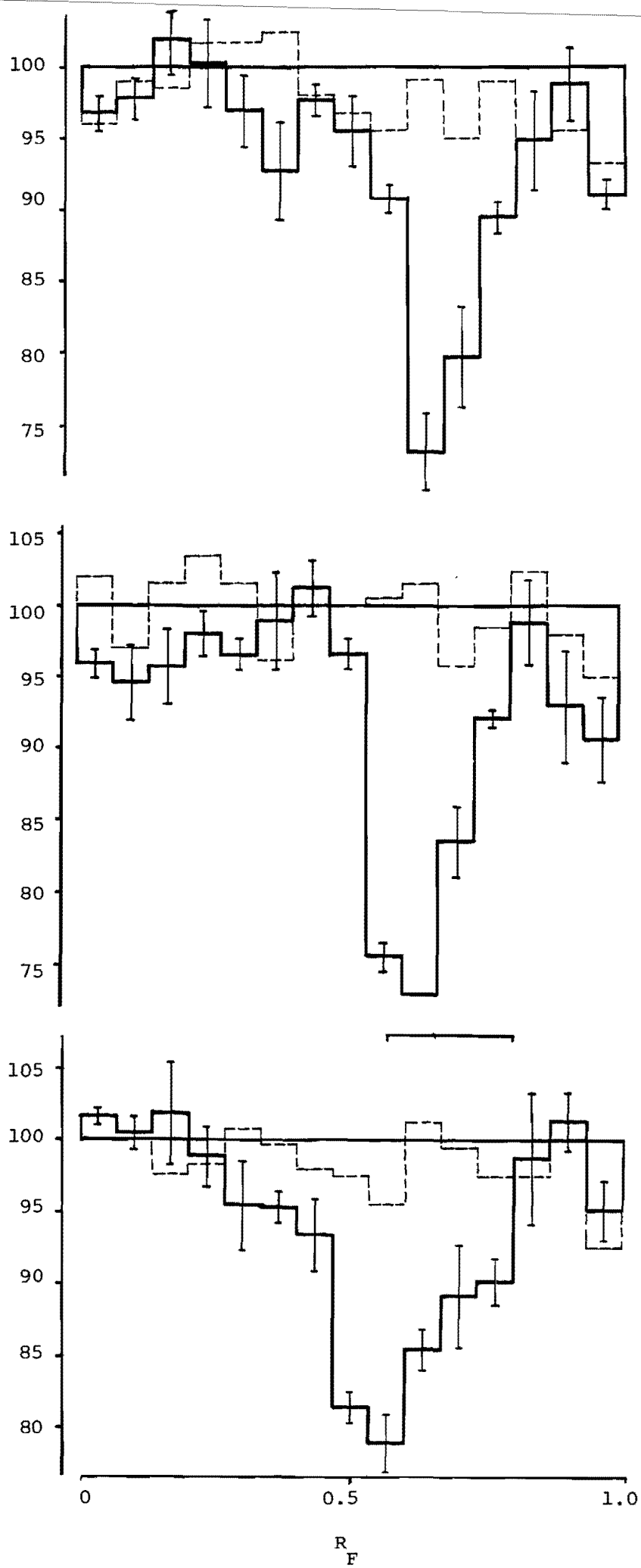


Figure 92. Wheat coleoptile section assay of acidic ether-soluble extracts obtained from leaves of *Alnus viridis* seedlings harvested on day 15 of SD treatment (H3). The equivalent of 0.25 g DW of leaf material was assayed and the solvent system was isopropanol:NH₃:H₂O (10:1:1::v:v:v). Other details as in figure 18.

Coleoptile growth (percentage of control)

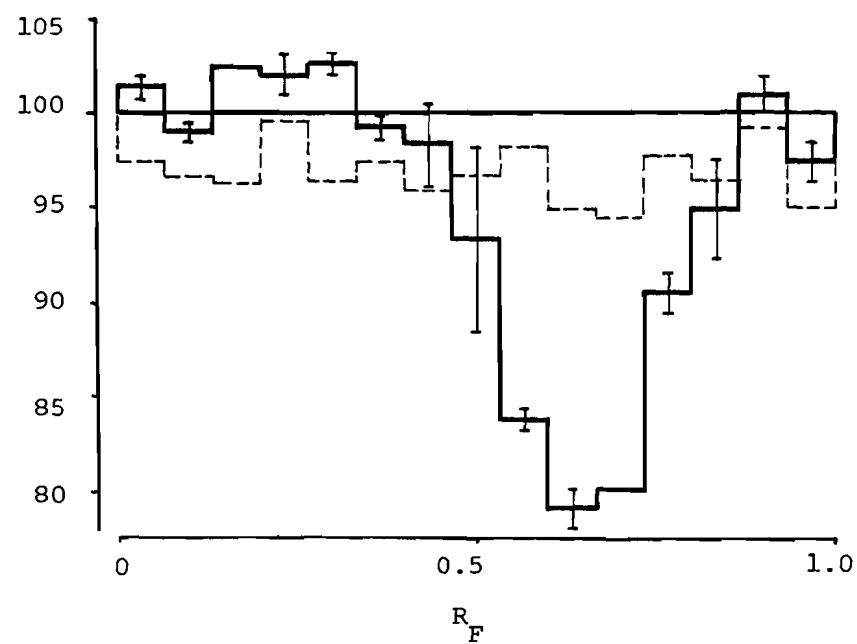
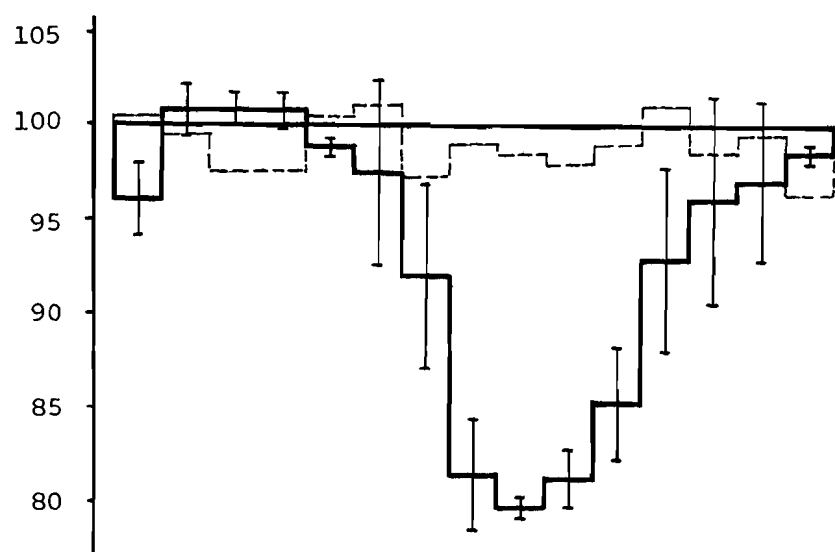
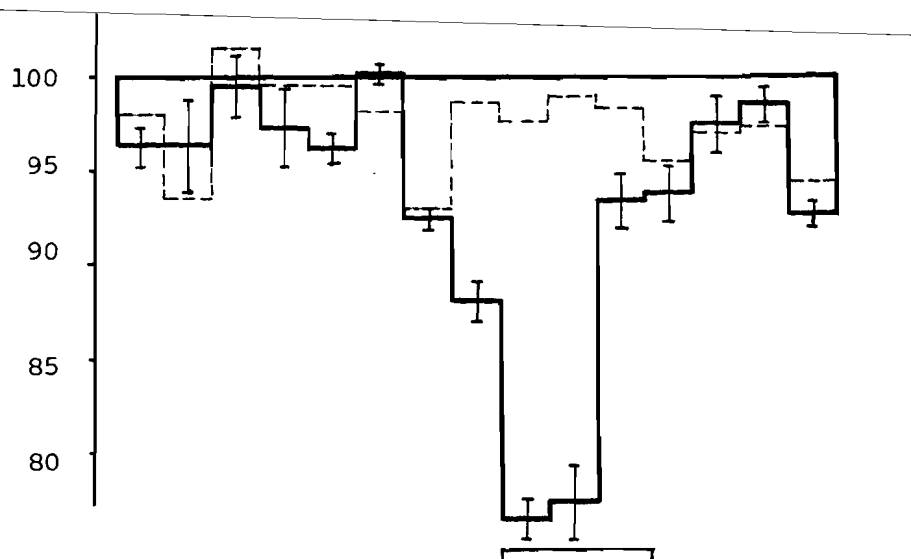


Figure 93. Lettuce hypocotyl assay of acidic ether-soluble extracts obtained from leaves of *Alnus viridis* seedlings harvested on day 0 of SD treatment (H1). The equivalent of 0.25 g DW of leaf material was assayed and the solvent system was isopropanol:NH₃:H₂O (10:1:1::v:v:v). Other details as in figure 26.

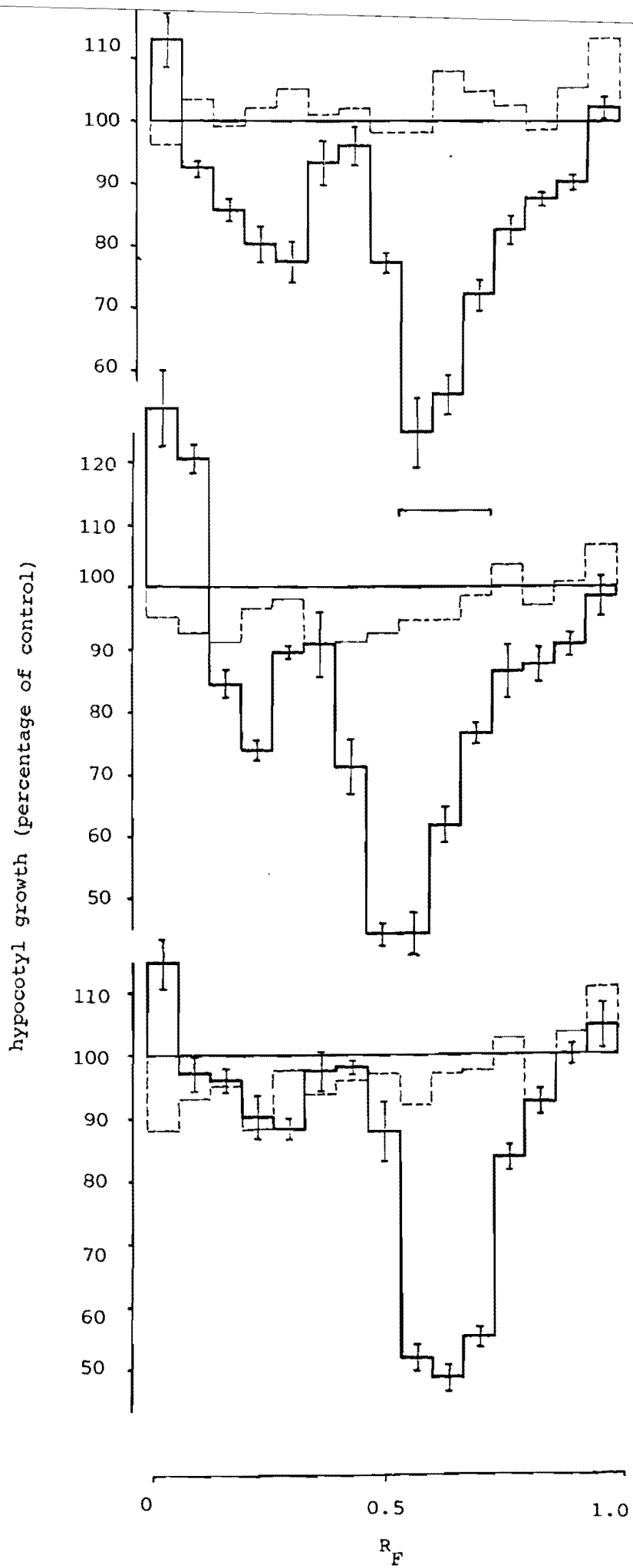


Figure 94. Lettuce hypocotyl assay of acidic ether-soluble extracts obtained from leaves of *Alnus viridis* seedlings harvested on day 7 of SD treatment (H2). The equivalent of 0.25 g DW of leaf material was assayed and the solvent system was isopropanol:NH₃:H₂O (10:1:1::v:v:v). Other details as in figure 26.

hypocotyl growth (percentage of control)

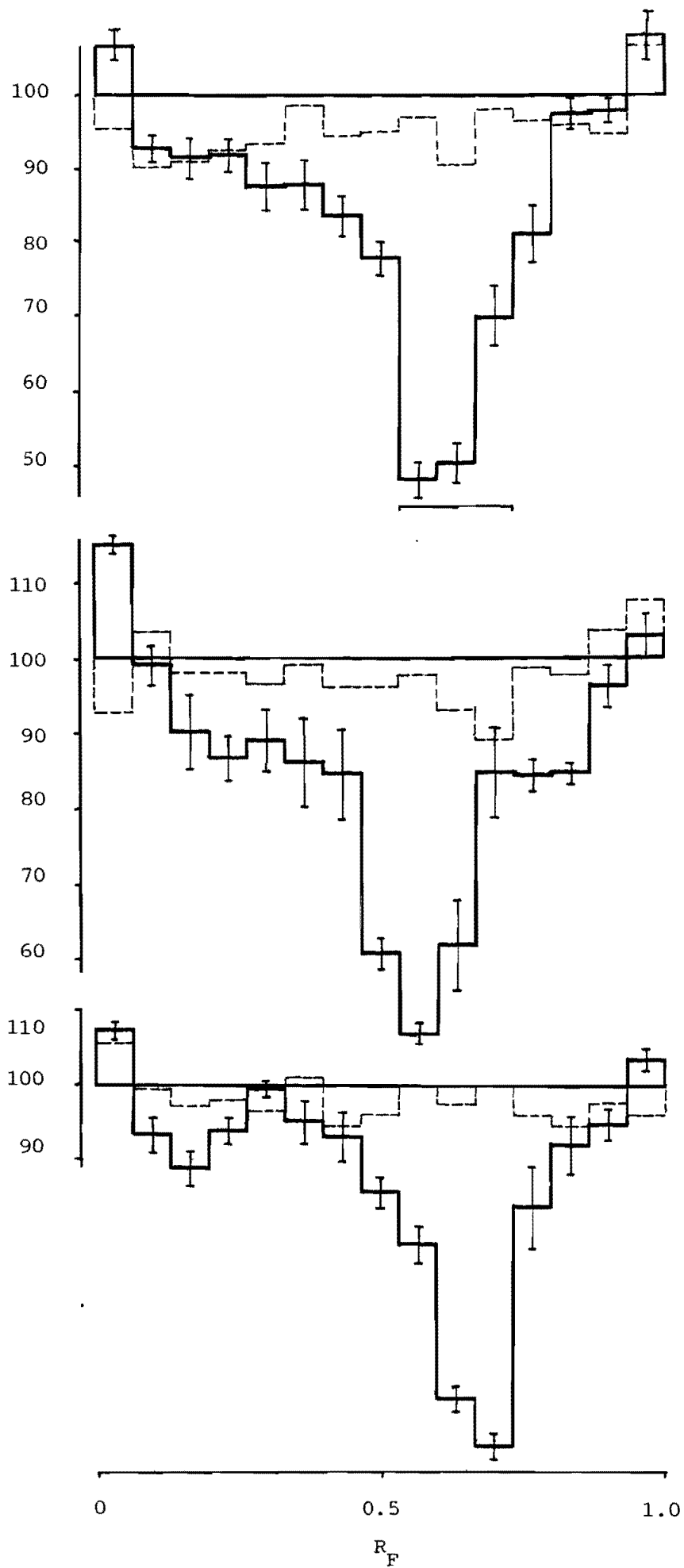


Figure 95. Lettuce hypocotyl assay of acidic ether-soluble extracts obtained from leaves of *Alnus viridis* seedlings harvested on day 15 of SD treatment (H3). The equivalent of 0.25 g DW of leaf material was assayed and the solvent system was isopropanol:NH₃:H₂O (10:1:1::v:v:v). Other details as in figure 26.

hypocotyl growth (percentage of control)

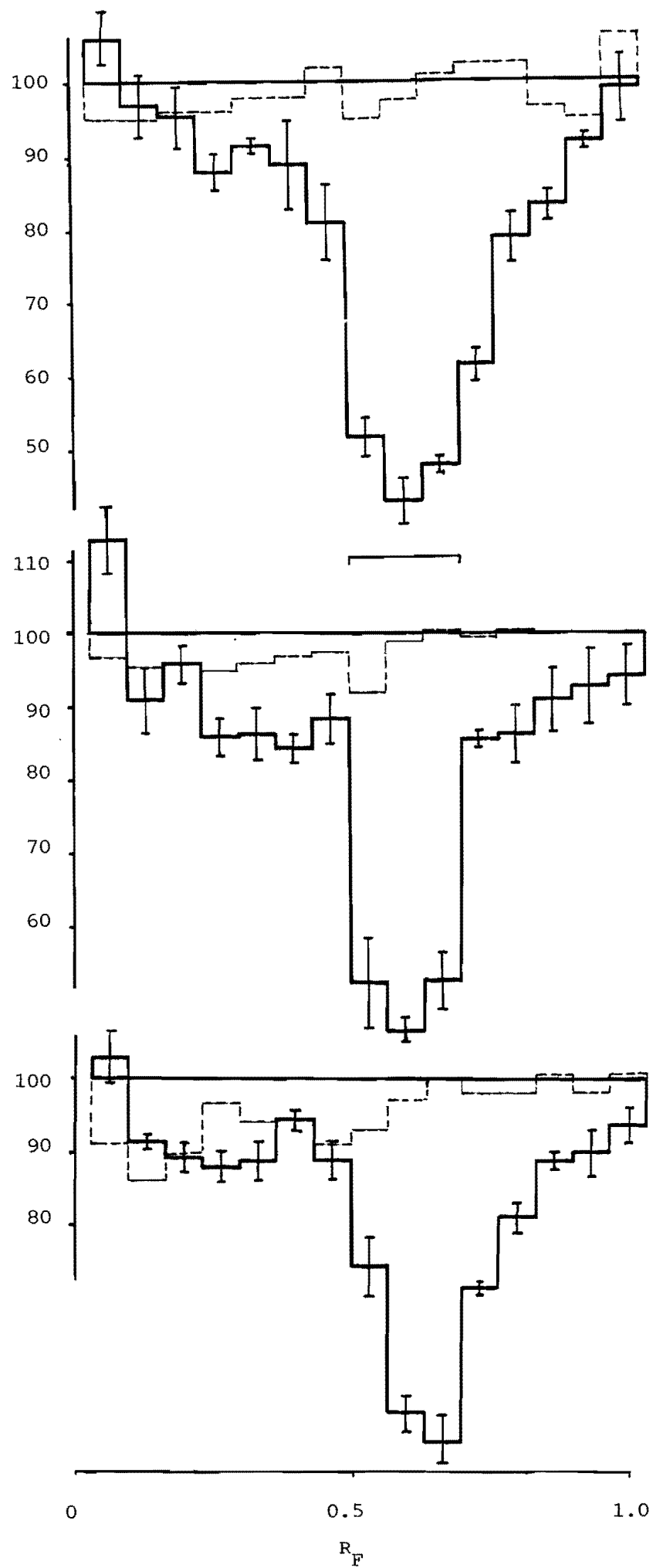


Figure 96. Wheat coleoptile section assay of the serial dilution of an acidic ether-soluble extract obtained from leaves of *Alnus viridis* seedlings harvested on day 0 of SD treatment (H1). The equivalent of (a) 0.025, (b) 0.125, (c) 0.25 and (d) 0.5 g DW of leaf material was assayed. Other details as in figure 18.

Coleoptile growth (percentage of control)

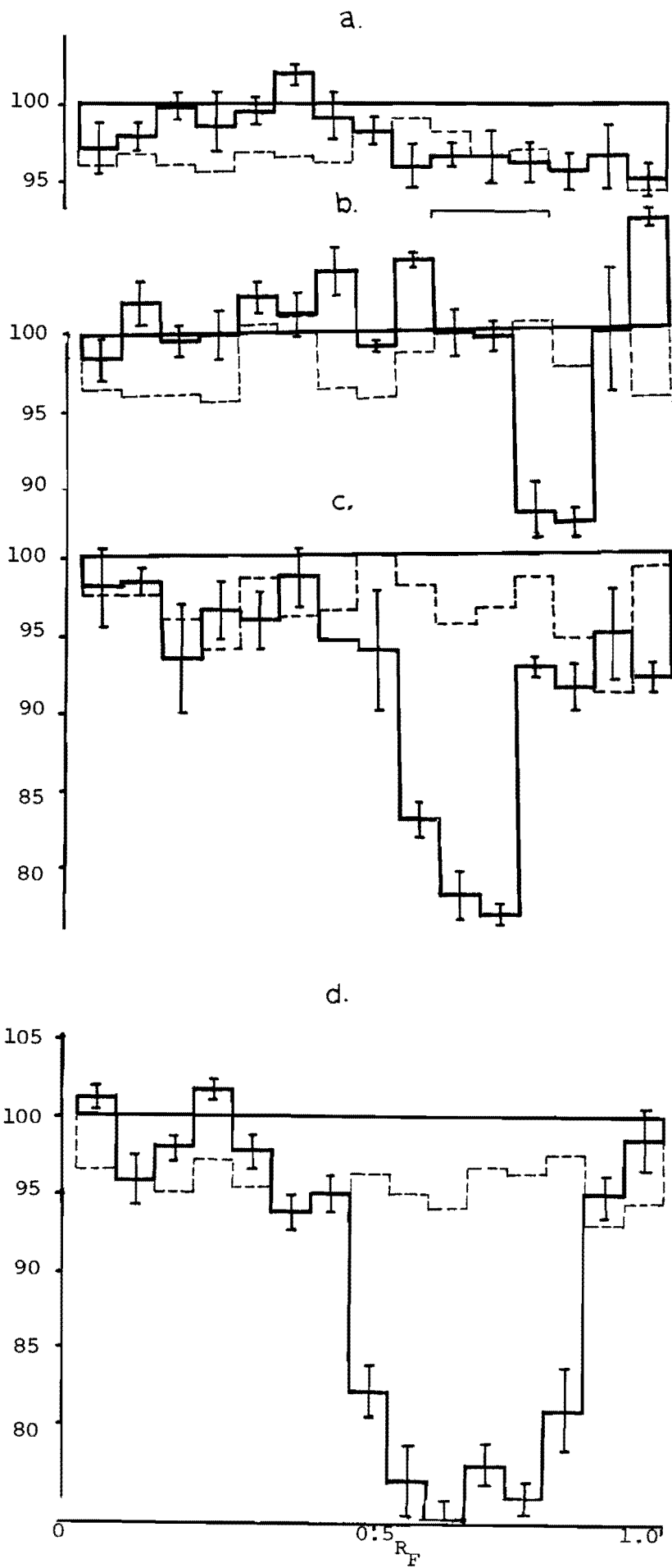


Figure 97. Wheat coleoptile section assay of the serial dilution of an acidic ether-soluble extract obtained from leaves of *Alnus viridis* seedlings harvested on day 15 of SD treatment (H3). The equivalent of (a) 0.025, (b) 0.125, (c) 0.25 and (d) 0.50 g DW of leaf material was assayed. Other details as in figure 18.

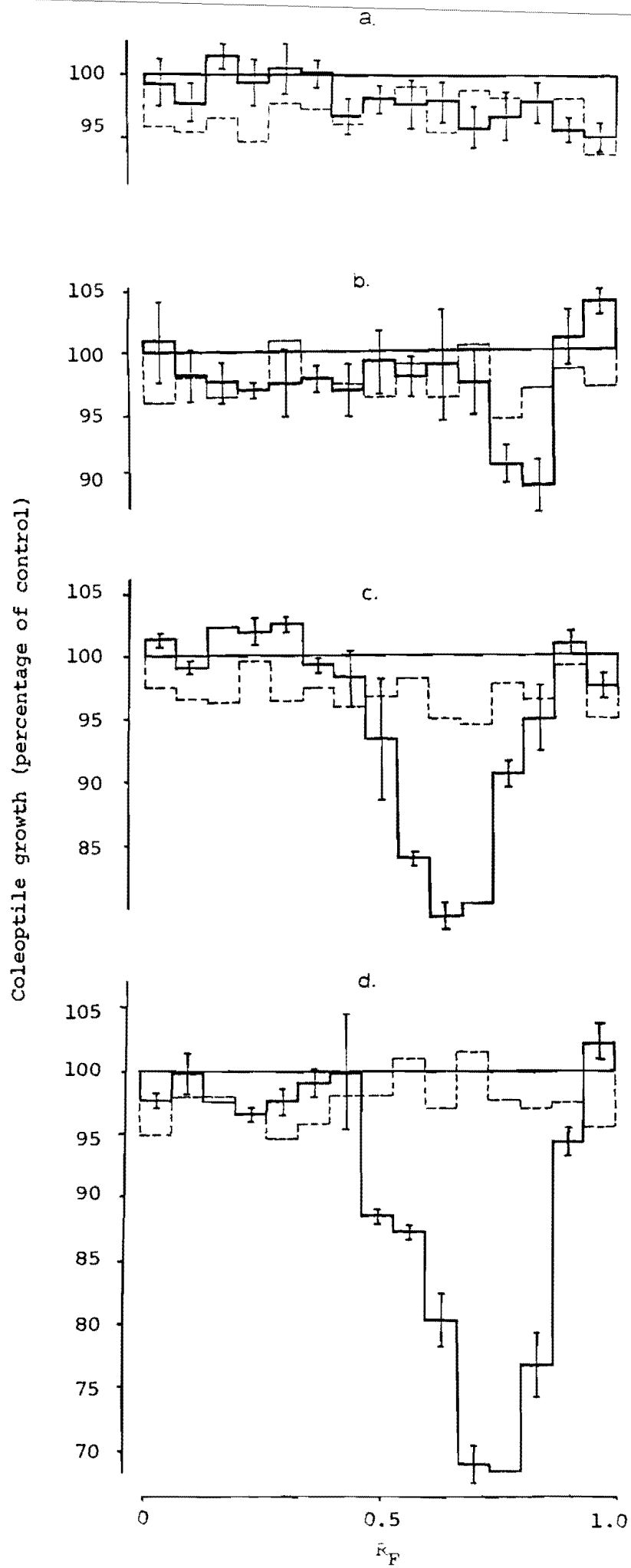


Figure 98. Lettuce hypocotyl assay of the serial dilution of an acidic ether-soluble extract obtained from leaves of *Alnus viridis* seedlings harvested on day 0 of SD treatment (H1). The equivalent of (a) 0.025, (b) 0.125, (c) 0.25 and (d) 0.50 g DW of leaf material was assayed. Other details as in figure 26.

hypocotyl growth (percentage of control)

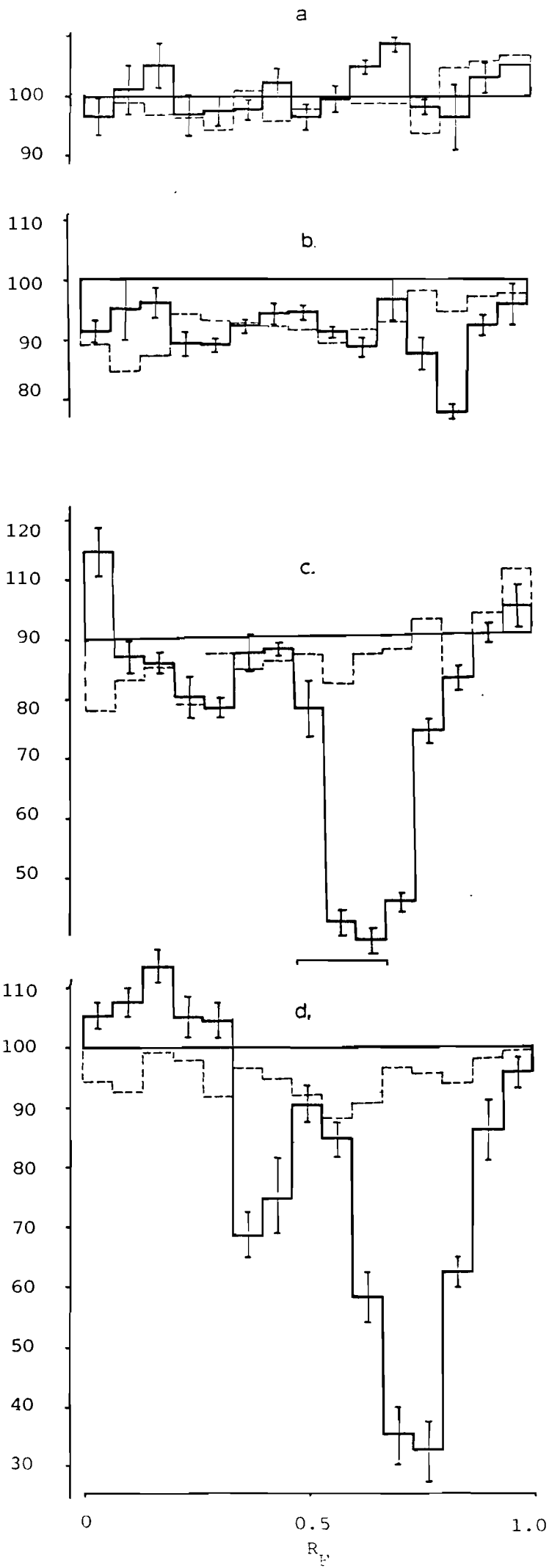


Figure 99. Lettuce hypocotyl assay of the serial dilution of an acidic ether-soluble extract obtained from leaves of *Alnus viridis* seedlings harvested on day 15 of SD treatment (H3). The equivalent of (a) 0.025, (b) 0.125, (c) 0.25 and (d) 0.50 g DW of leaf material was assayed. Other details as in figure 26.

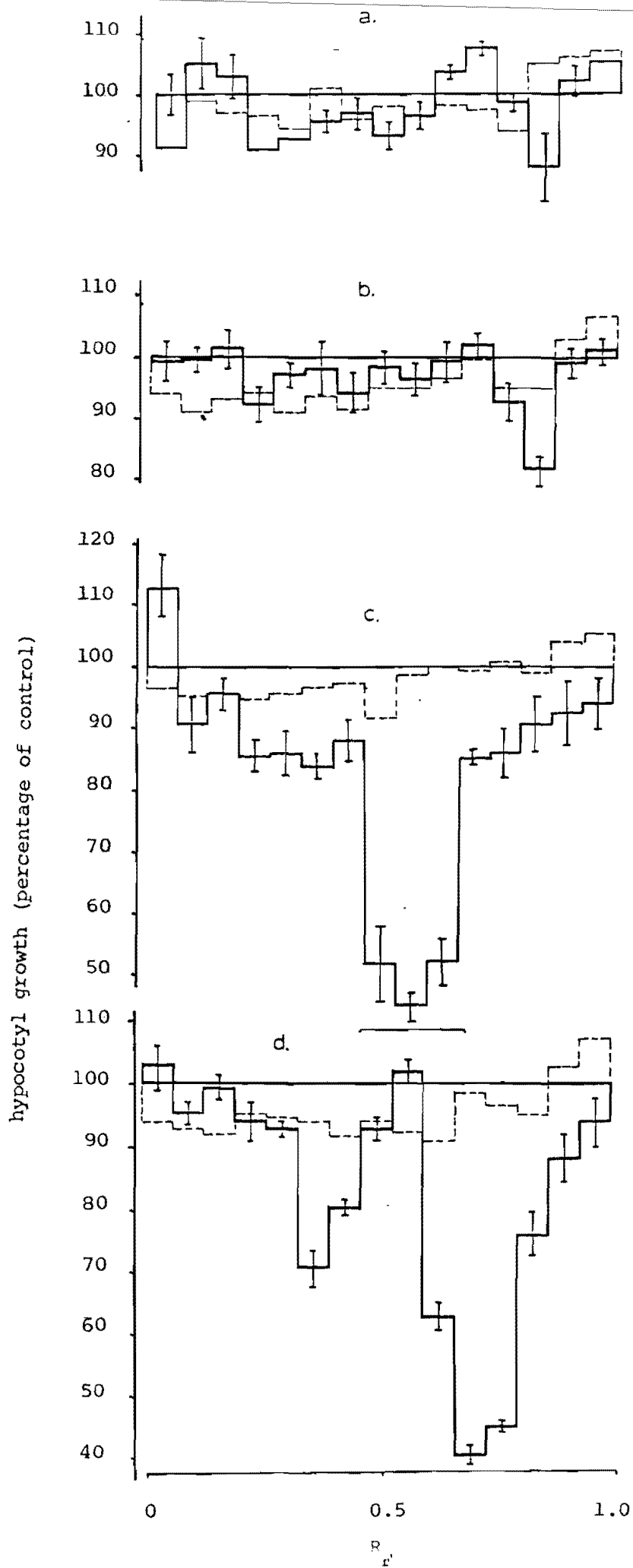


Figure 100. Wheat coleoptile section assay of acidic ether-soluble extracts obtained from apices of *Alnus viridis* seedlings harvested on day 0 of SD treatment (H1). The equivalent of 0.20 g DW of apical material was assayed and the solvent system was isopropanol:NH₃:H₂O (10:1:1::v:v:v). Other details as in figure 18.

Coleoptile growth (percentage of control)

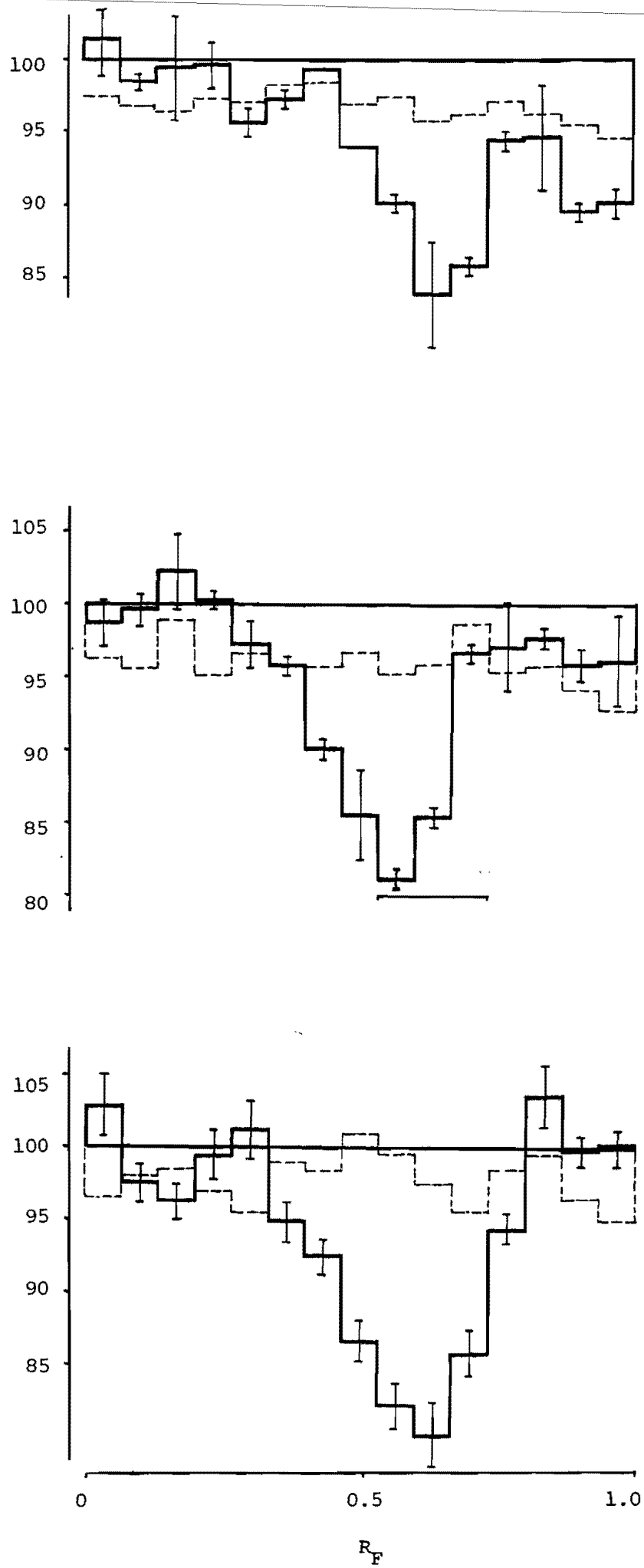


Figure 101. Wheat coleoptile section assay of acidic ether-soluble extracts obtained from apices of *Alnus viridis* seedlings harvested on day 7 of SD treatment (H2). The equivalent of 0.20 g DW of apical material was assayed and the solvent system was isopropanol:NH₃:H₂O (10:1:1::v:v:v). Other details as in figure 18.

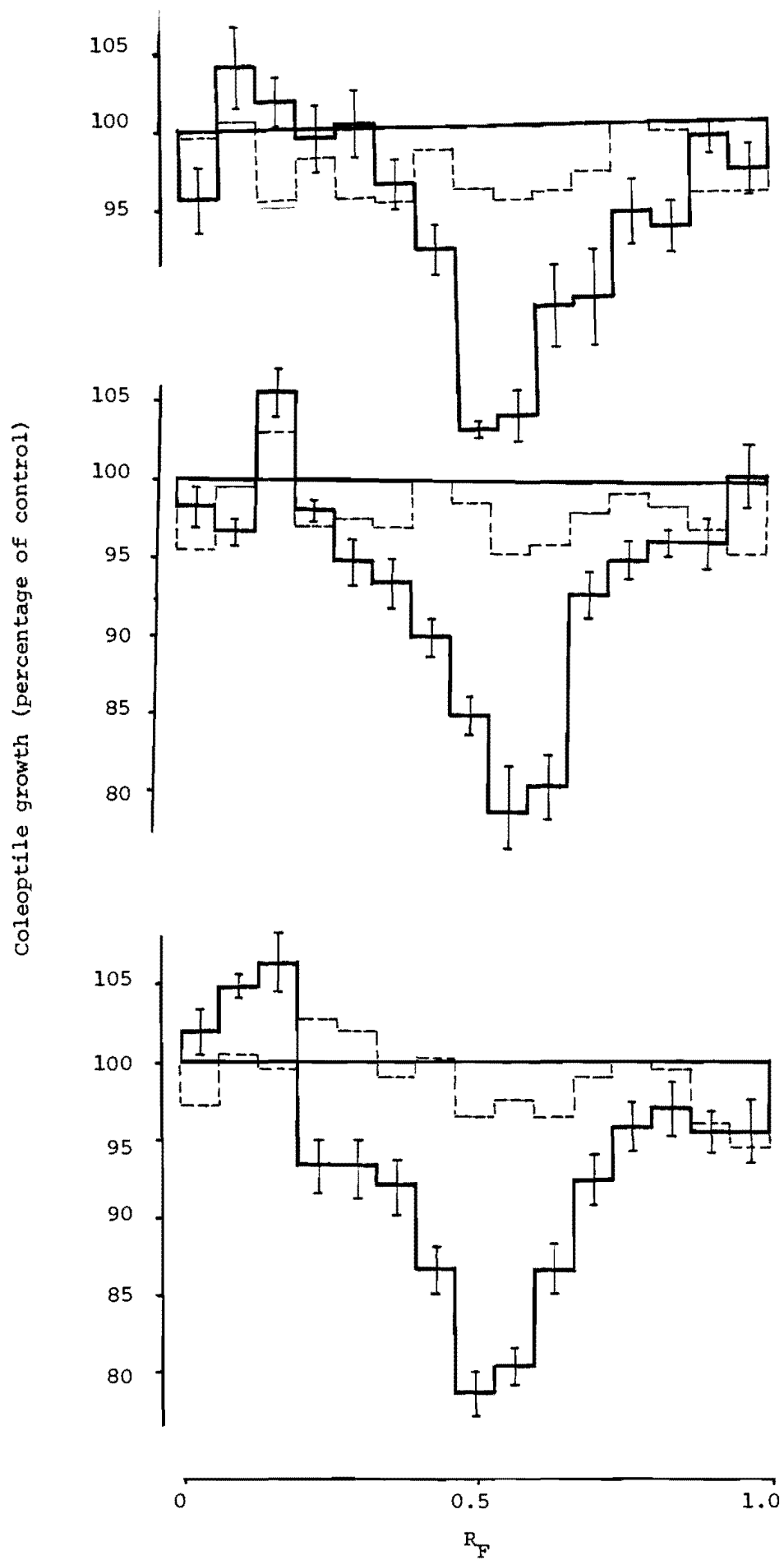


Figure 102. Wheat coleoptile section assay of acidic ether-soluble extracts obtained from apices of *Alnus viridis* seedlings harvested on day 15 of SD treatment (H3). The equivalent of 0.20 g DW of apical material was assayed and the solvent system was isopropanol:NH₃:H₂O (10:1:1::v:v:v). Other details as in figure 18.

Coleoptile growth (percentage of control)

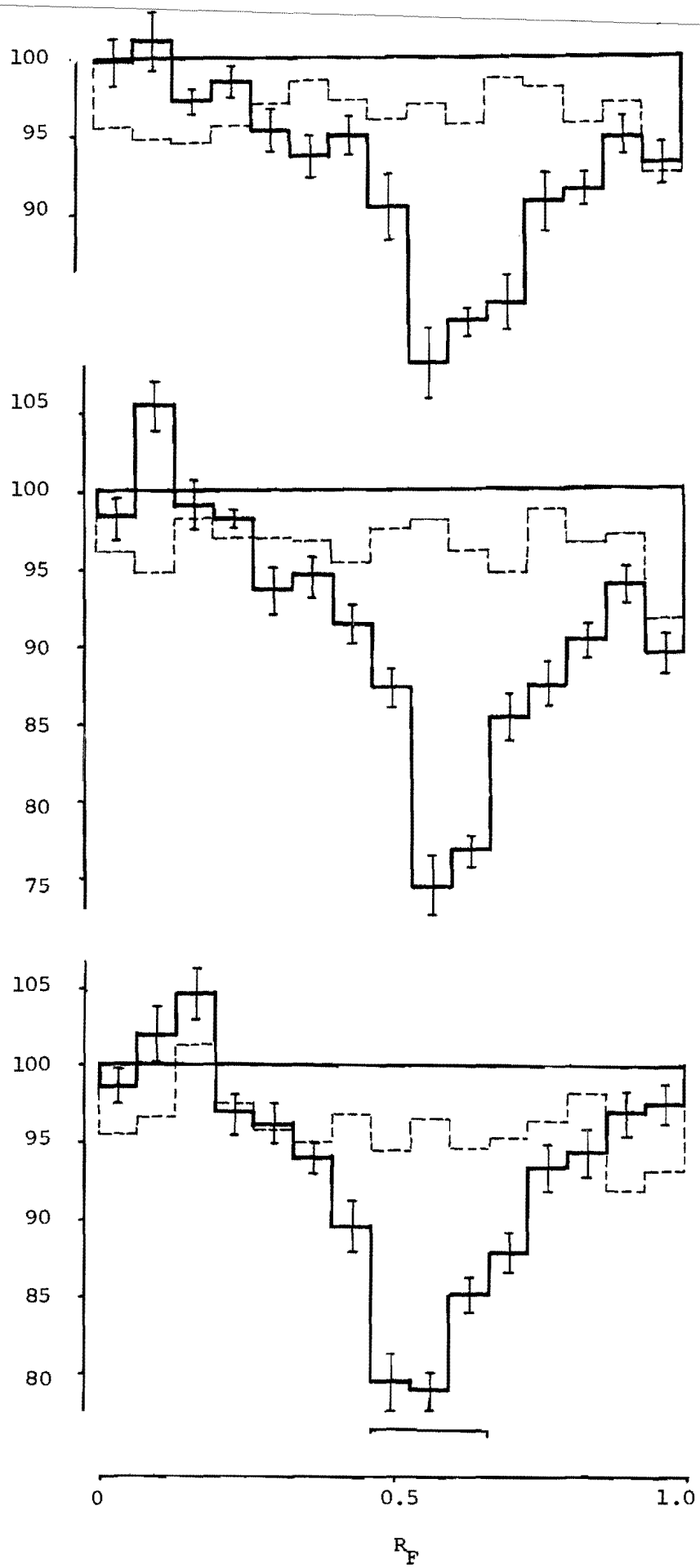


Figure 103. Lettuce hypocotyl assay of acidic ether-soluble extracts obtained from apices of *Alnus viridis* seedlings harvested on day 0 of SD treatment (H1). The equivalent of 0.2 g DW of apical material was assayed and the solvent system was isopropanol:NH₃:H₂O (10:1:1::v:v:v). Other details as in figure 26.

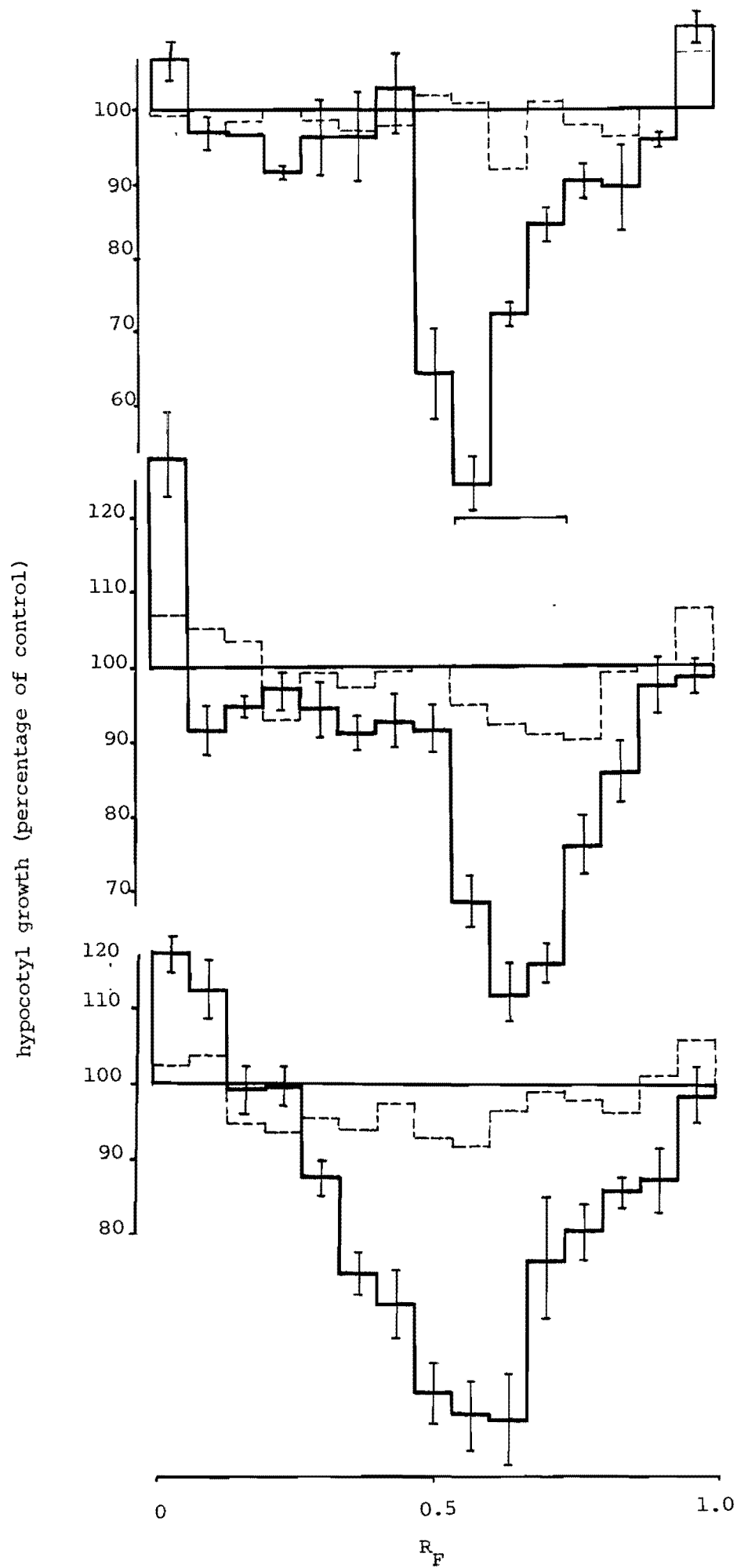


Figure 104. Lettuce hypocotyl assay of acidic ether-soluble extracts obtained from apices of *Alnus viridis* seedlings harvested on day 7 of SD treatment (H2). The equivalent of 0.2 g DW of apical material was assayed and the solvent system was isopropanol:NH₃:H₂O (10:1:1::v:v:v). Other details as in figure 26.

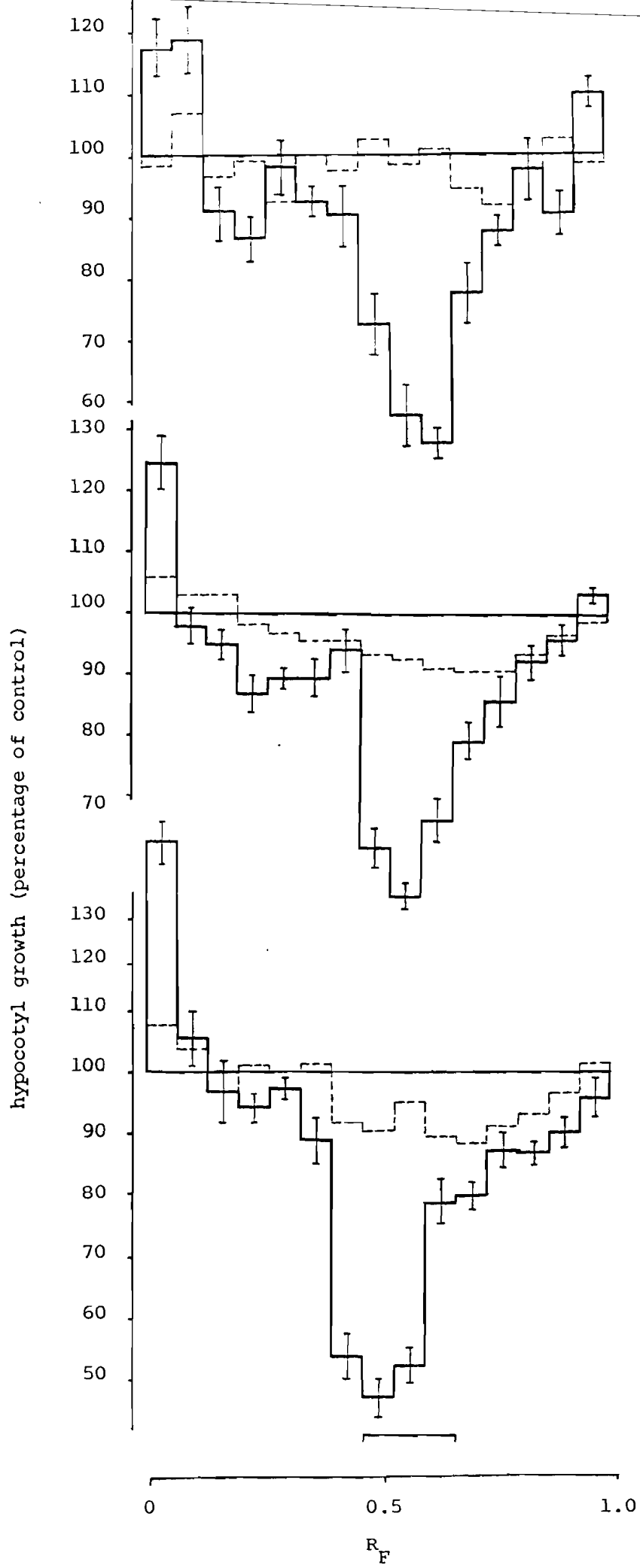


Figure 105. Lettuce hypocotyl assay of acidic ether-soluble extracts obtained from apices of *Alnus viridis* seedlings harvested on day 15 of SD treatment (H3). The equivalent of 0.2 g DW of apical material was assayed and the solvent system was isopropanol:NH₃:H₂O (10:1:1::v:v:v). Other details as in figure 26.

hypocotyl growth (percentage of control)

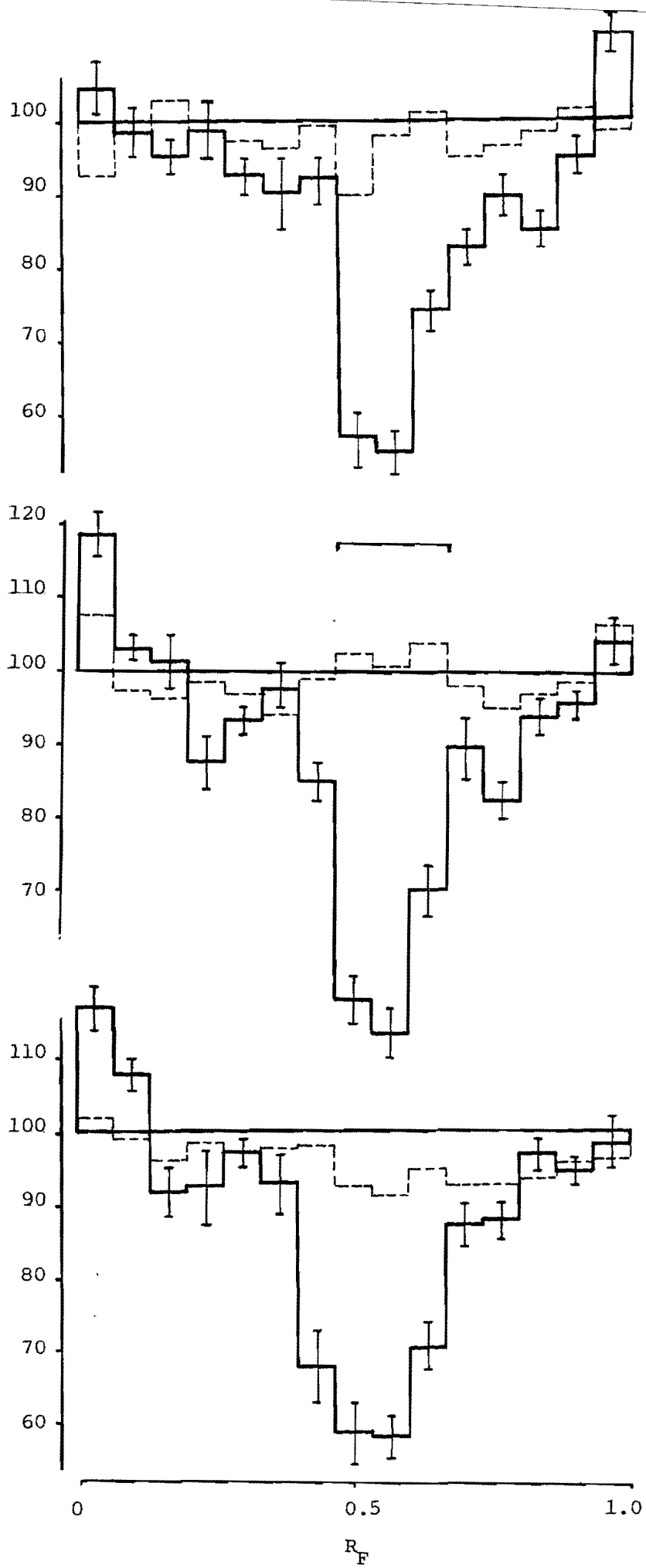


Figure 106. Lettuce hypocotyl assay of acidic ether-soluble extracts obtained from apices of *Alnus viridis* seedlings on (a) day 0 (H1), (b) day 7 (H2), and (c) day 15 (H3) of SD treatment. At each harvest date, the remainder of three replicate extracts after assays previous were combined and the equivalent of 0.14 g DW of apical material was assayed. Other details as in figure 26.

hypocotyl growth (percentage of control)

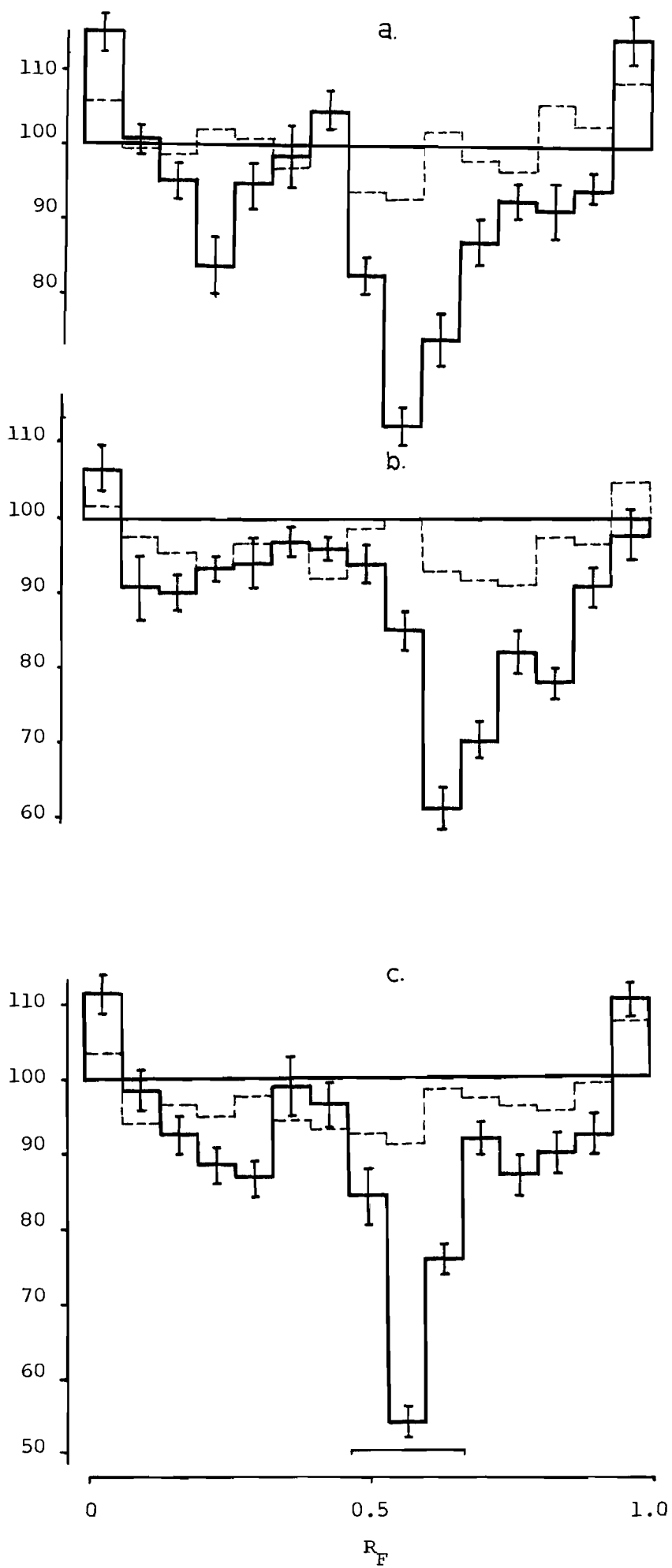
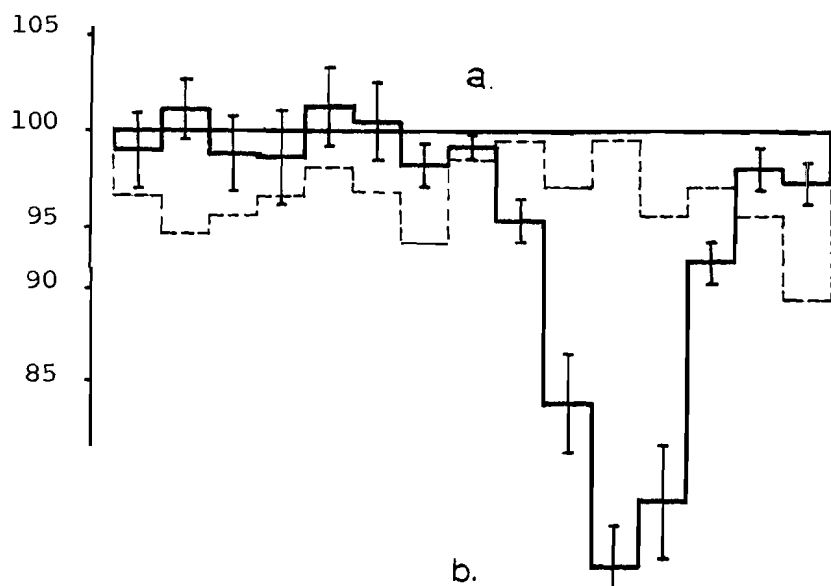
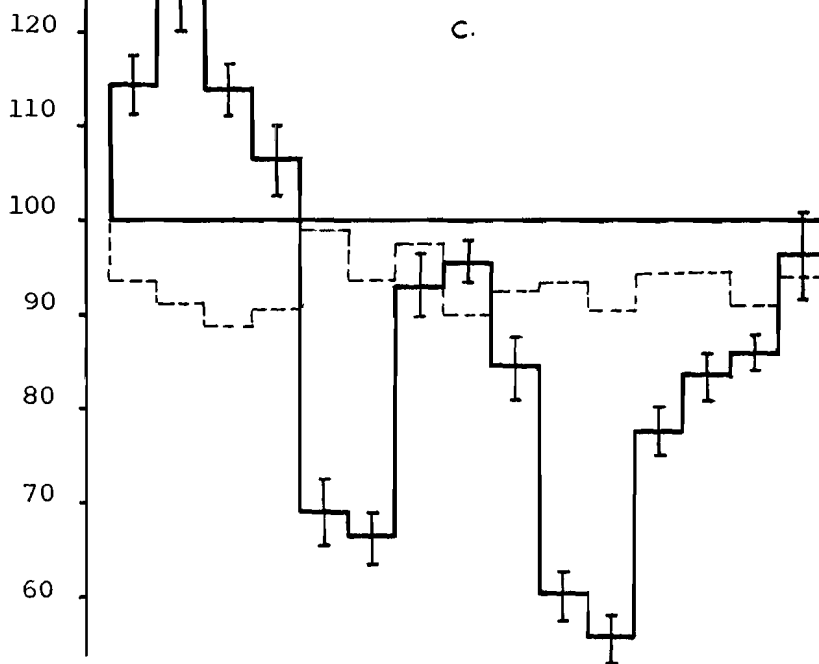
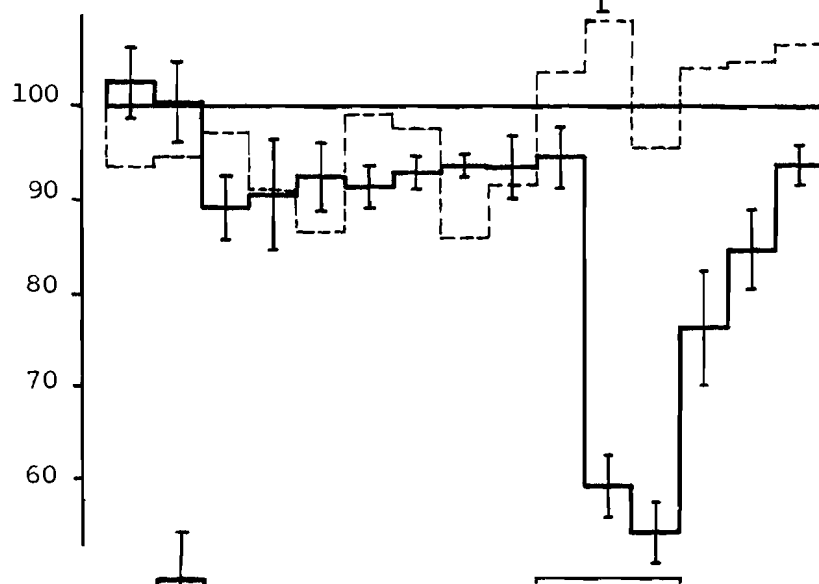


Figure 107. Bioassay of acidic ether-soluble extracts obtained from (a,b) leaves and (c) apices of *Alnus viridis* seedlings harvested on day 28 of SD treatment (H4). Leaf extracts were assayed using (a) wheat coleoptile sections and (b) lettuce hypocotyls and the apical extract (c) using lettuce hypocotyls. The equivalent of 0.25 g DW of leaf and 0.19 g DW of apical material were assayed and the chromatographs were developed in isopropanol:NH₃:H₂O (10:1:1::v:v:v). Other details as in figures 18 and 26.

coleoptile growth (% of control)



hypocotyl growth (percentage of control)



0 0.5 1.0
 R_F

Figure 108. Extension growth of *Alnus viridis* seedlings with intact (●) and pruned (▲) root systems, maintained under LD's (16 h photoperiod) at 20°C. Arrows indicate the start of the treatments. The experiment was repeated three times using five plants (a, b) and six plants (c). The equivalent of approx. one-third (a and b) and one-sixth (c) of the root system was left undisturbed. The vertical bars show one S.E. above and/or below the lines.

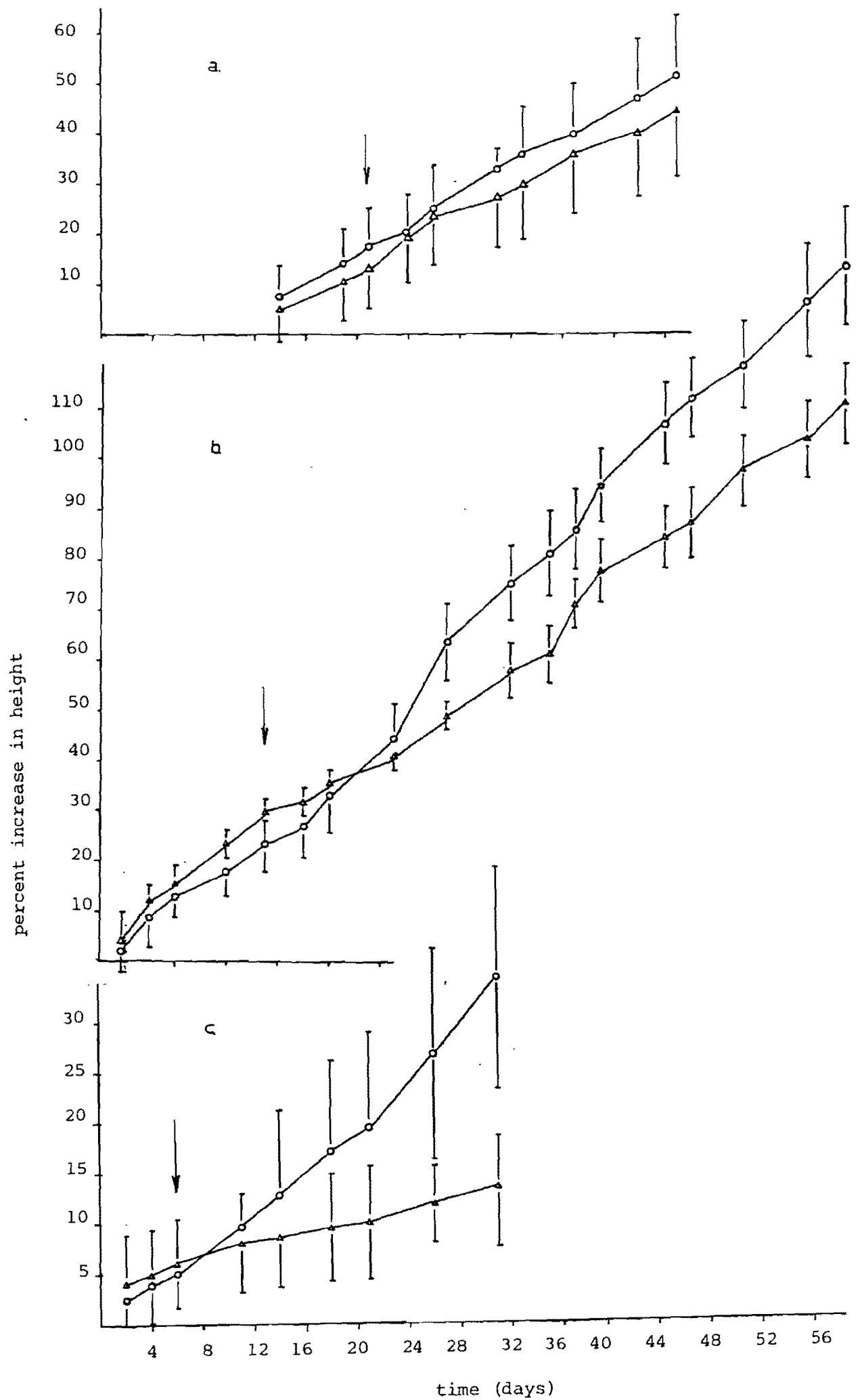


Figure 109. Effect of growth regulator application on the extension growth of *Alnus viridis* seedlings maintained under LD's (16 h photoperiod) and 14°C day (equivalent to the high light intensity period) and 10°C night temperatures. Daily spray applications of (b) ABA, (c) CCC, (d) ABA + CCC, and (e) C9 were made to the leaves and apices, and the youngest fully-expanded leaf was partially immersed in the test solution. Plants treated with water solvent (a) served as controls, and the arrows indicate the start of the treatments. Each value represents percent increase in growth and is the mean \pm S.E. of three plants.

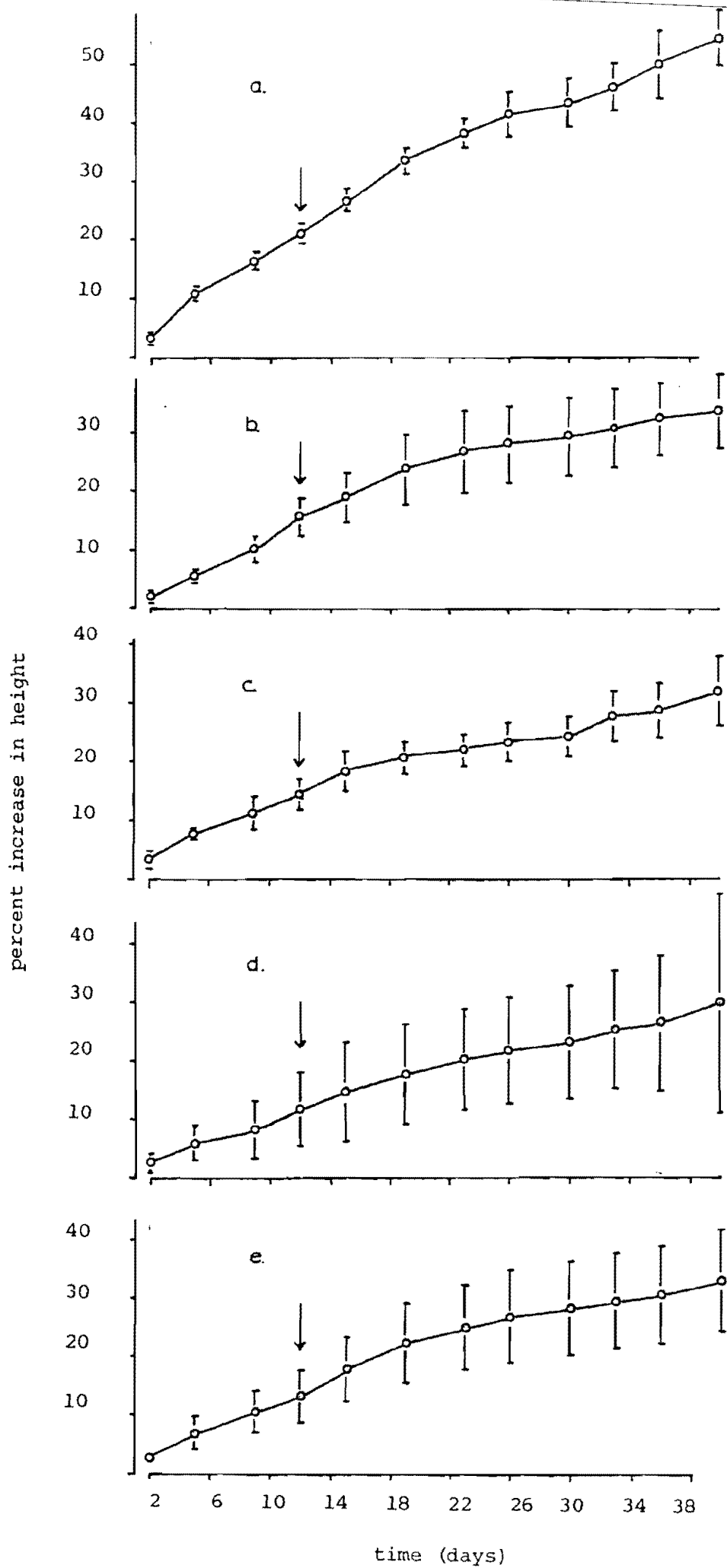


Figure 110. Effect of growth regulator application on extension growth of *Alnus viridis* seedlings maintained under LD's (16 h photoperiod) at 20°C. Daily spray applications of ABA (Δ), CCC (\square) and ABA + CCC (\bullet) were made to the leaves and apices, and the youngest fully expanded leaf was partially immersed in the test solution. Plants treated with water solvent (\circ) served as controls, and the arrows indicate the start of the treatments. Each value represents percent increase in growth and is the mean \pm S.E. of five plants.

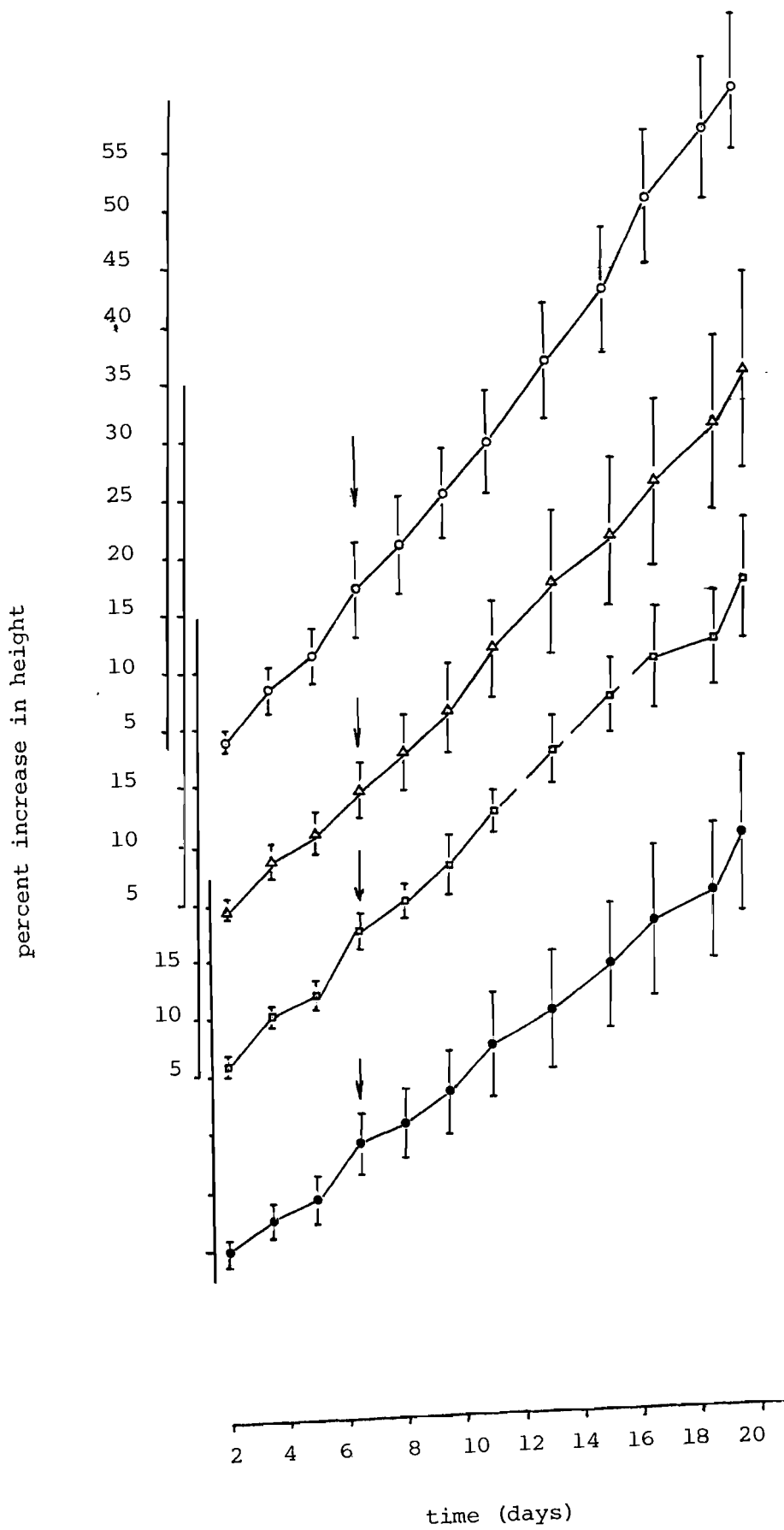


Figure 111. Effect of growth regulator application on the extension growth of *Alnus viridis* seedlings maintained under LD's (16 h photoperiod) at 20°C. Daily applications of (b) ABA, (c) CCC, and (d) ABA + CCC were made to the leaves and apices. Plants treated with water solvent (a) served as controls and the arrows indicate the start of the treatments. Each value represents percent increase in growth and is the mean \pm S.E. of six plants.

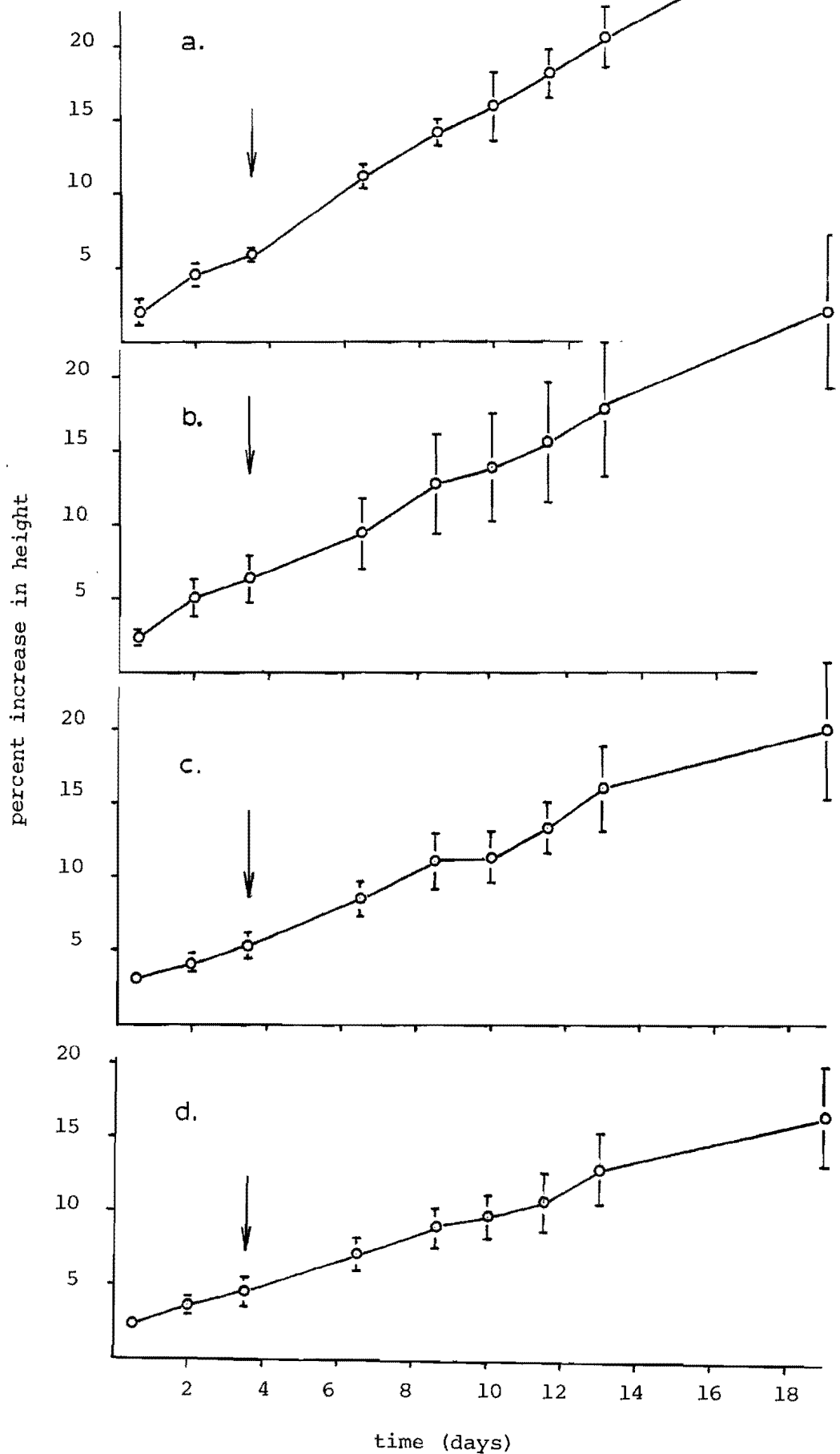


Figure 112. Effect of short chain fatty acid application on the extension growth of *Alnus viridis* seedlings maintained under LD's (16 h photoperiod) at 20°C. Daily spray applications of (b) C5, (c) C9, and (d) C10 were made to the leaves and apices and for the seedlings treated with C9 and C10, the youngest fully-expanded leaf was partially immersed in the test solutions. Plants treated with water solvent (a) served as controls and the arrows indicate the start of the treatments. Each value represents percent increase in growth and is the mean \pm S.E. of five plants.

percent increase in height

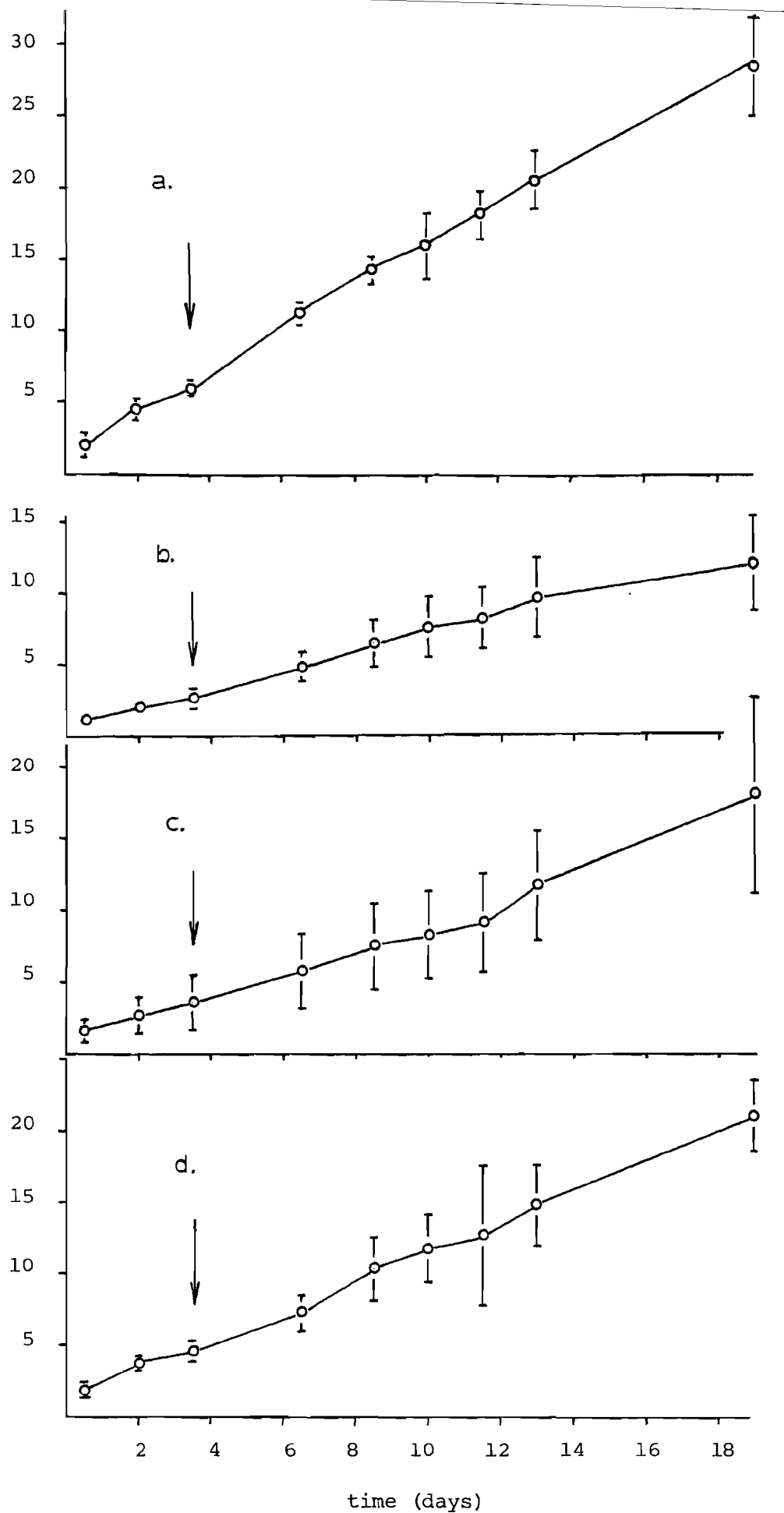
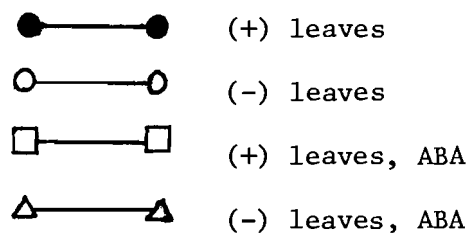


Figure 113. The effect on (a) bud burst and (b) the number of leaves retained after growth regulator application to the base of isolated, growing shoots of *Alnus viridis*. The values represent the number of growing buds or the number of leaves retained as a function of position on the stem with the leaves retained (●, □) or removed (○, △) at the beginning of the experiment. Each treatment comprised of 10 shoots. The number in brackets indicates the number of growing shoots which had added one or more nodes.



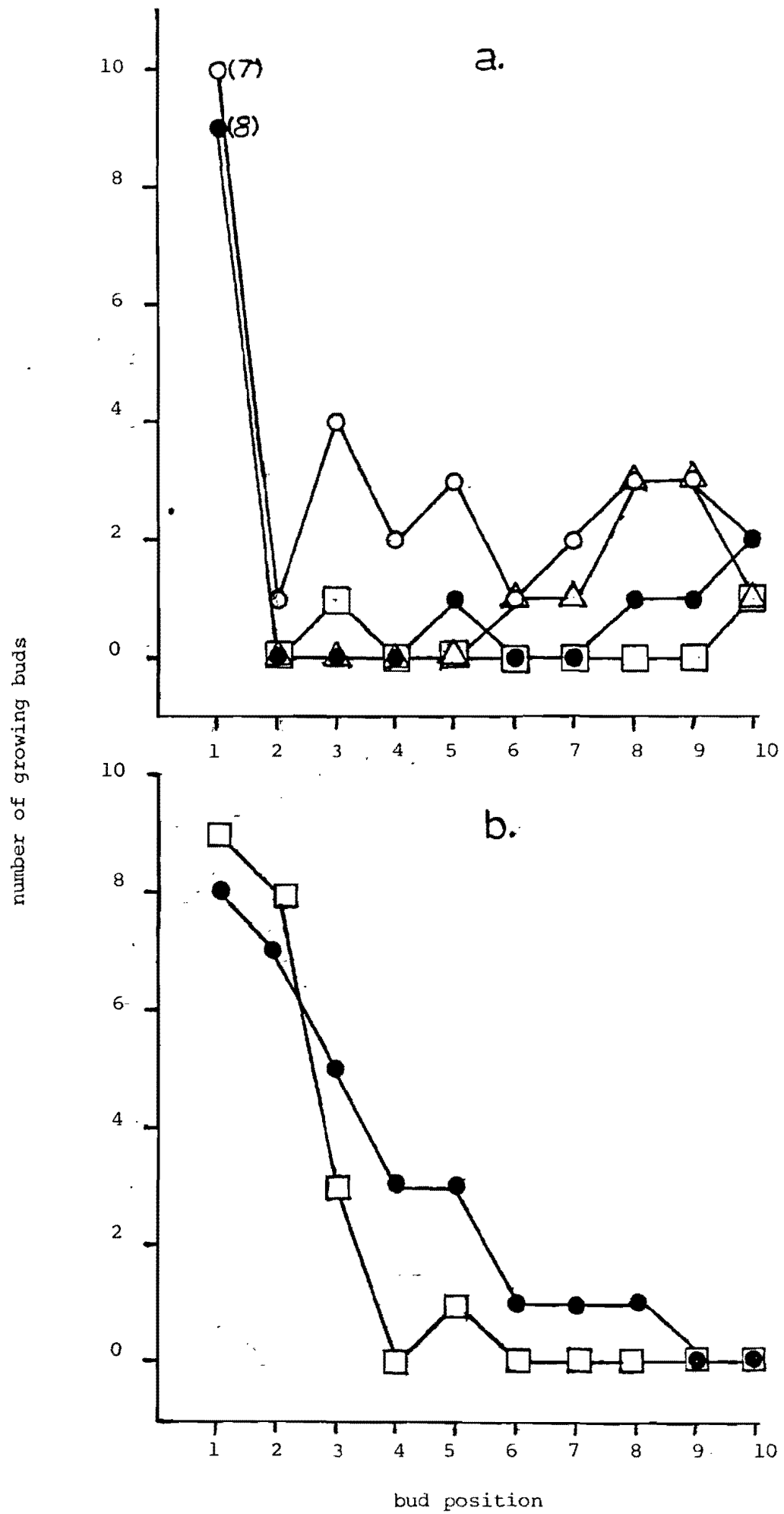
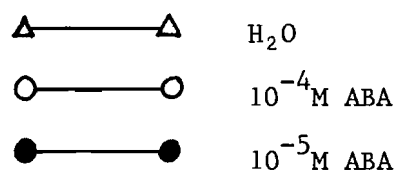


Figure 114. The effect on bud burst of abscisic acid application to the base of isolated dormant shoots of *Populus nigra* "Italica" (late winter harvest). The values represent the percent bud burst as a function of position on the stem. Each value is the mean of four replicates, each comprising 10 shoots. The vertical bars indicate the least significant differences at the (a) 5% and (b) 1% level.



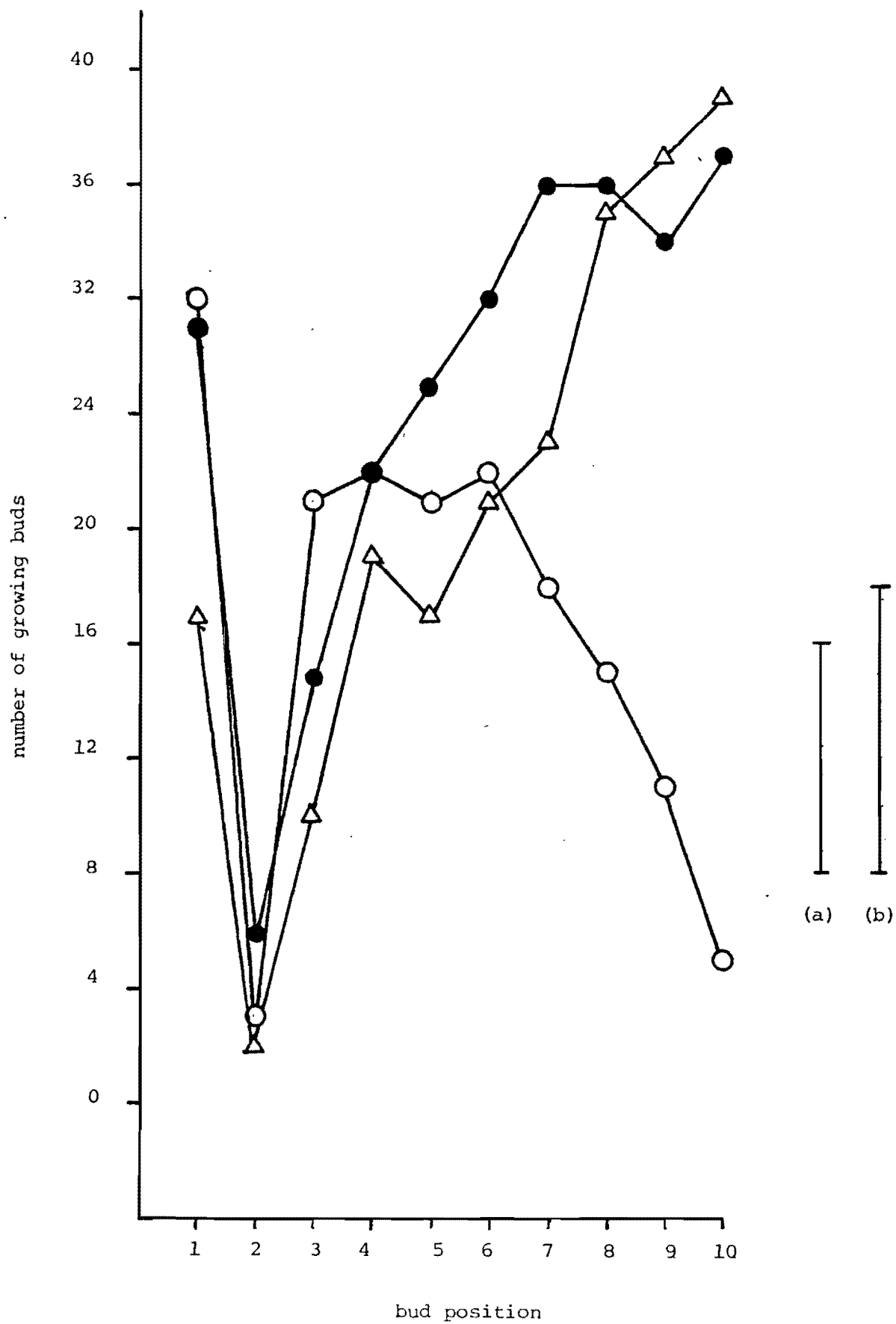
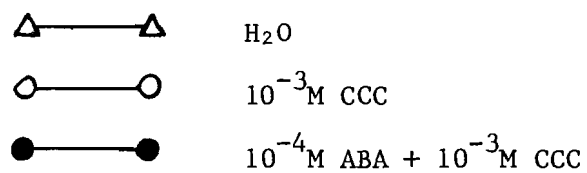


Figure 115. The effect on bud burst of the growth regulator application to the base of isolated dormant shoots of *Populus nigra* "Italica" (late winter harvest). The values represent the percent bud burst as a function of position on the stem. Each value is the mean of four replicates, each comprising 10 shoots. The vertical bars indicate the least significant difference at the (a) 5% and (b) 1% level.



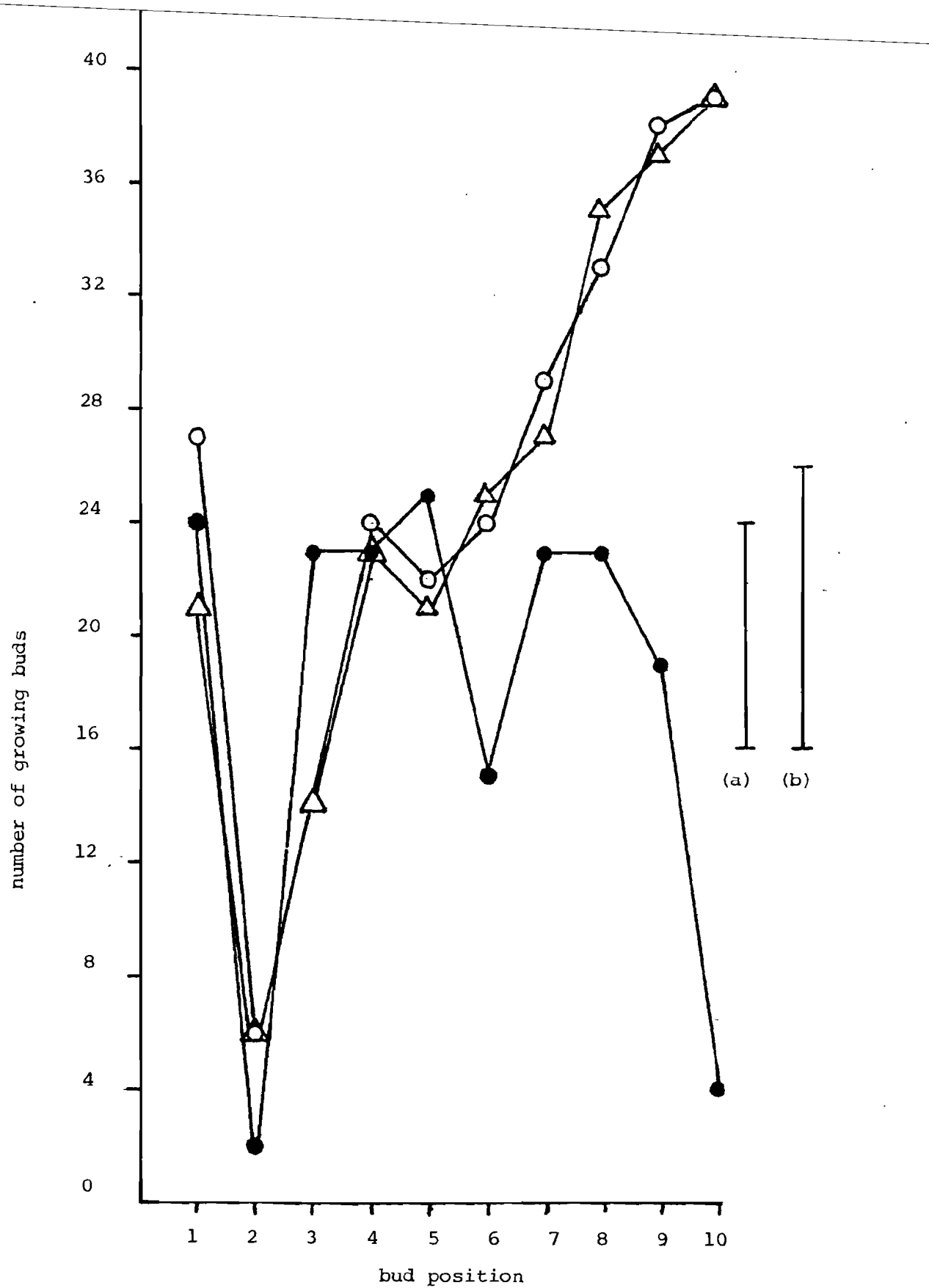
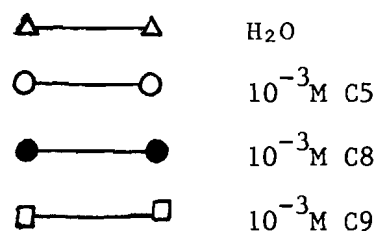


Figure 116. The effect on bud burst of short chain fatty acid application to the base of isolated, dormant shoots of *Populus nigra* "Italica" (late winter harvest). The values represent the percent bud burst as a function of position on the stem. Each value is the mean of four replicates, each comprising 10 shoots. The vertical bars indicate the least significant difference at the (a) 5% and (b) 1% level.



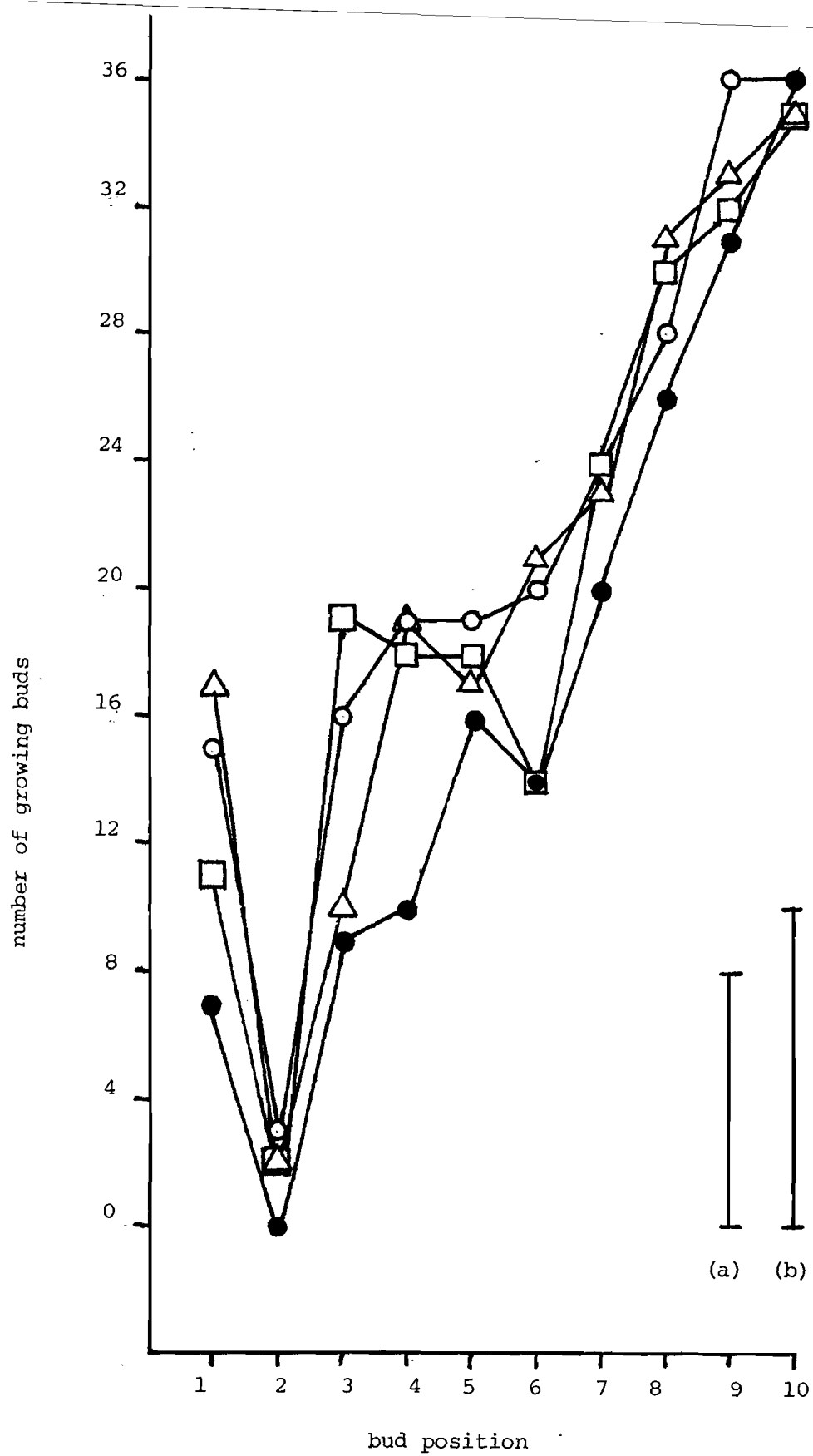
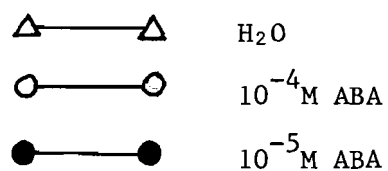


Figure 117. The effect on bud burst of abscisic acid application to the base of isolated dormant shoots of *Alnus viridis* (late winter harvest). The values represent percent bud burst as a function of position on the stem. Each value is the mean of four replicates, each comprising 10 shoots. Vertical bars indicate the least significant difference at the (a) 5% and (b) 1% level.



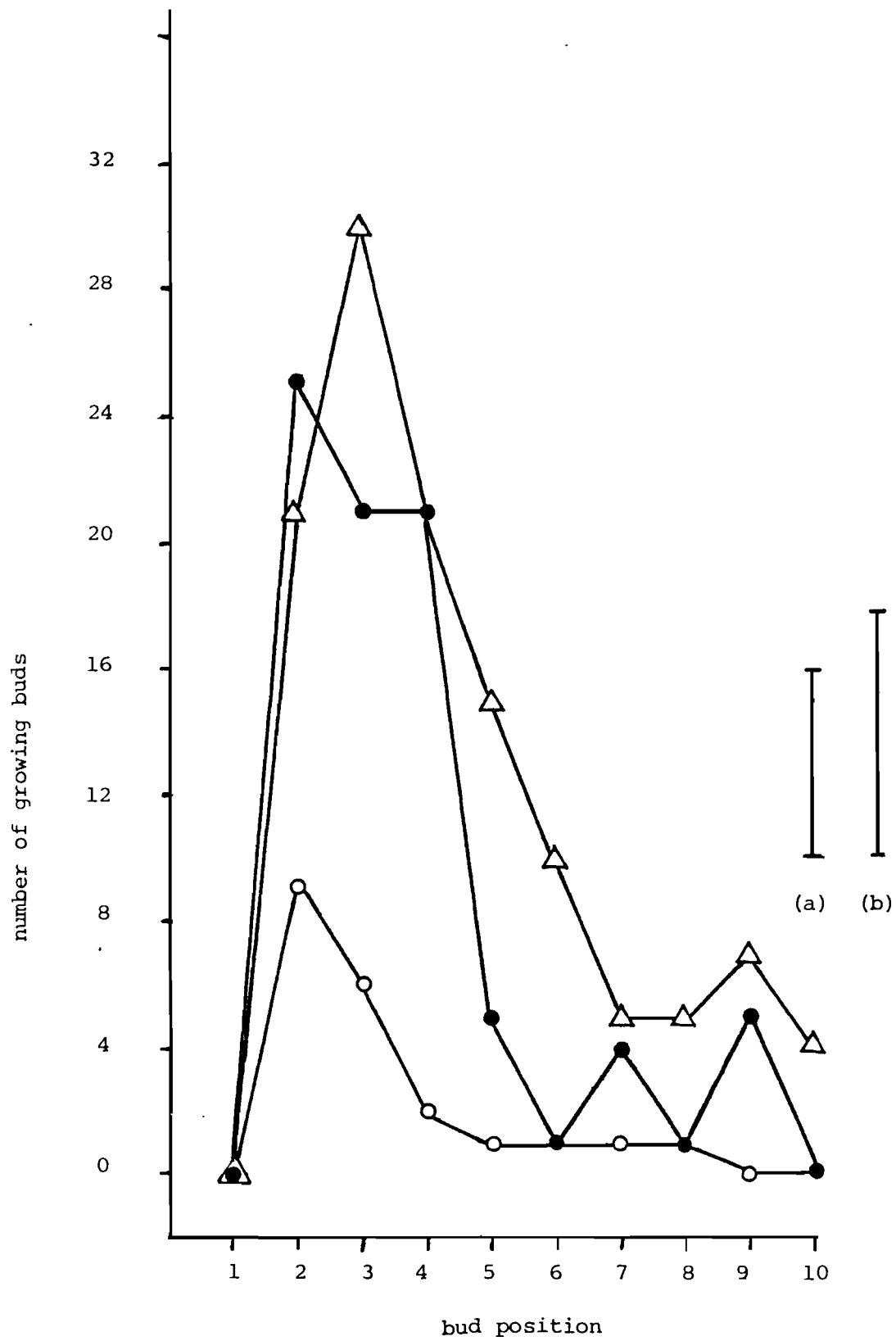
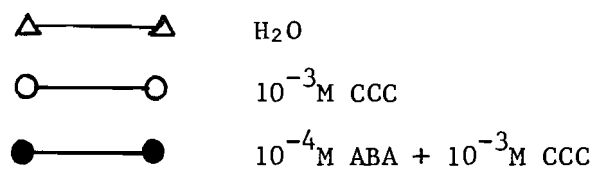


Figure 118. The effect on bud burst of the growth regulator application to the base of isolated dormant shoots of *Alnus viridis* (late winter harvest). The values represent percent bud burst as a function of position on the stem. Each value is the mean of four replicates, each comprising 10 shoots. Vertical bars indicate the least significant difference at the (a) 5% and (b) 1% level.



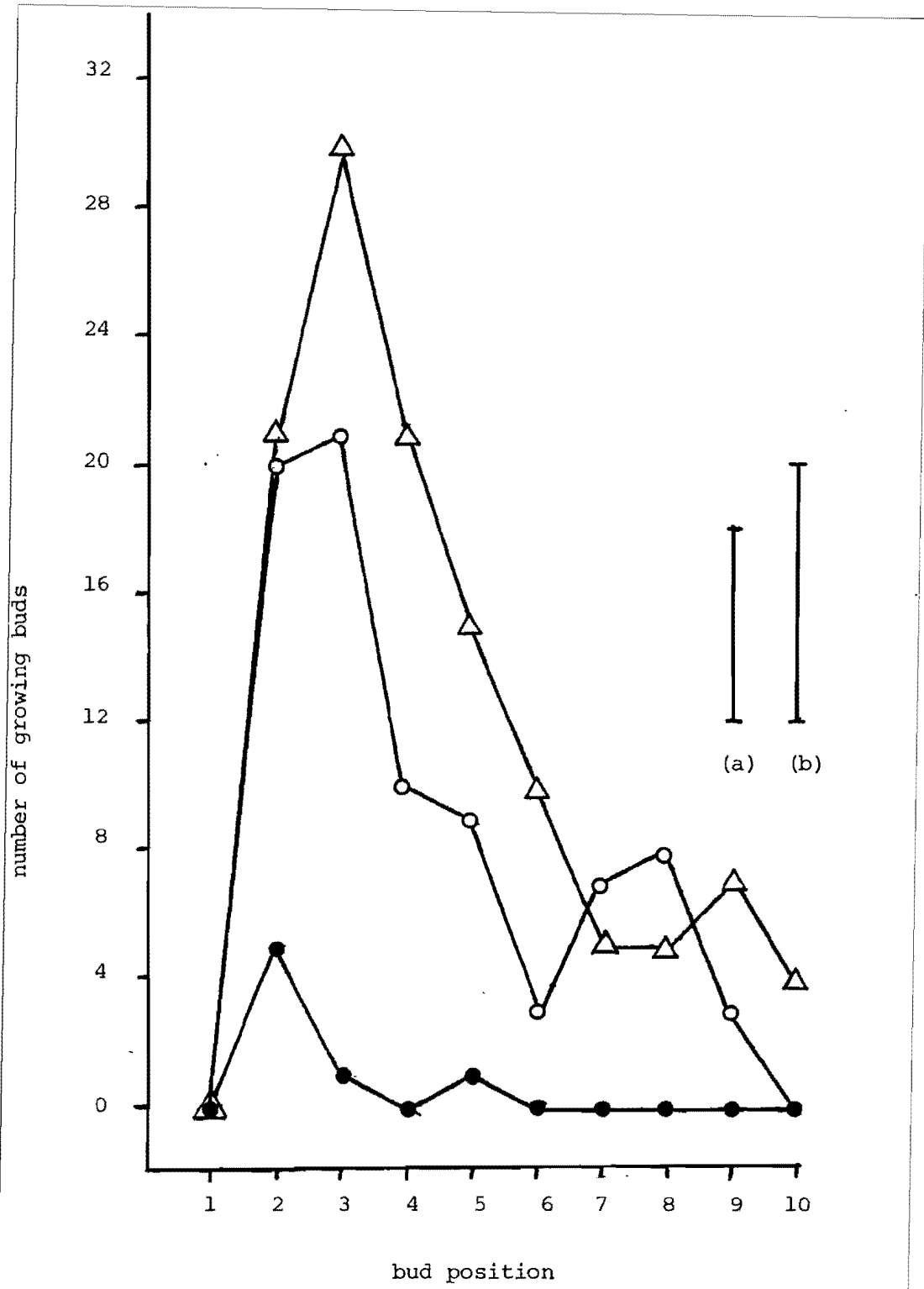
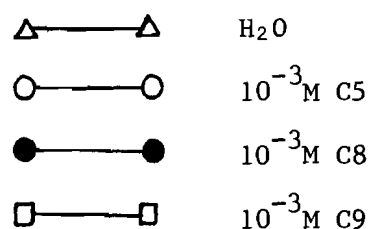


Figure 119. The effect on bud burst of short chain fatty acid application to the base of isolated dormant shoots of *Alnus viridis* (late winter harvest). The values represent percent bud burst as a function of bud position on the stem. Each value is the mean of four replicates, each comprising 10 shoots. Vertical bars indicate the least significant difference at the (a) 5% and (b) 1% level.



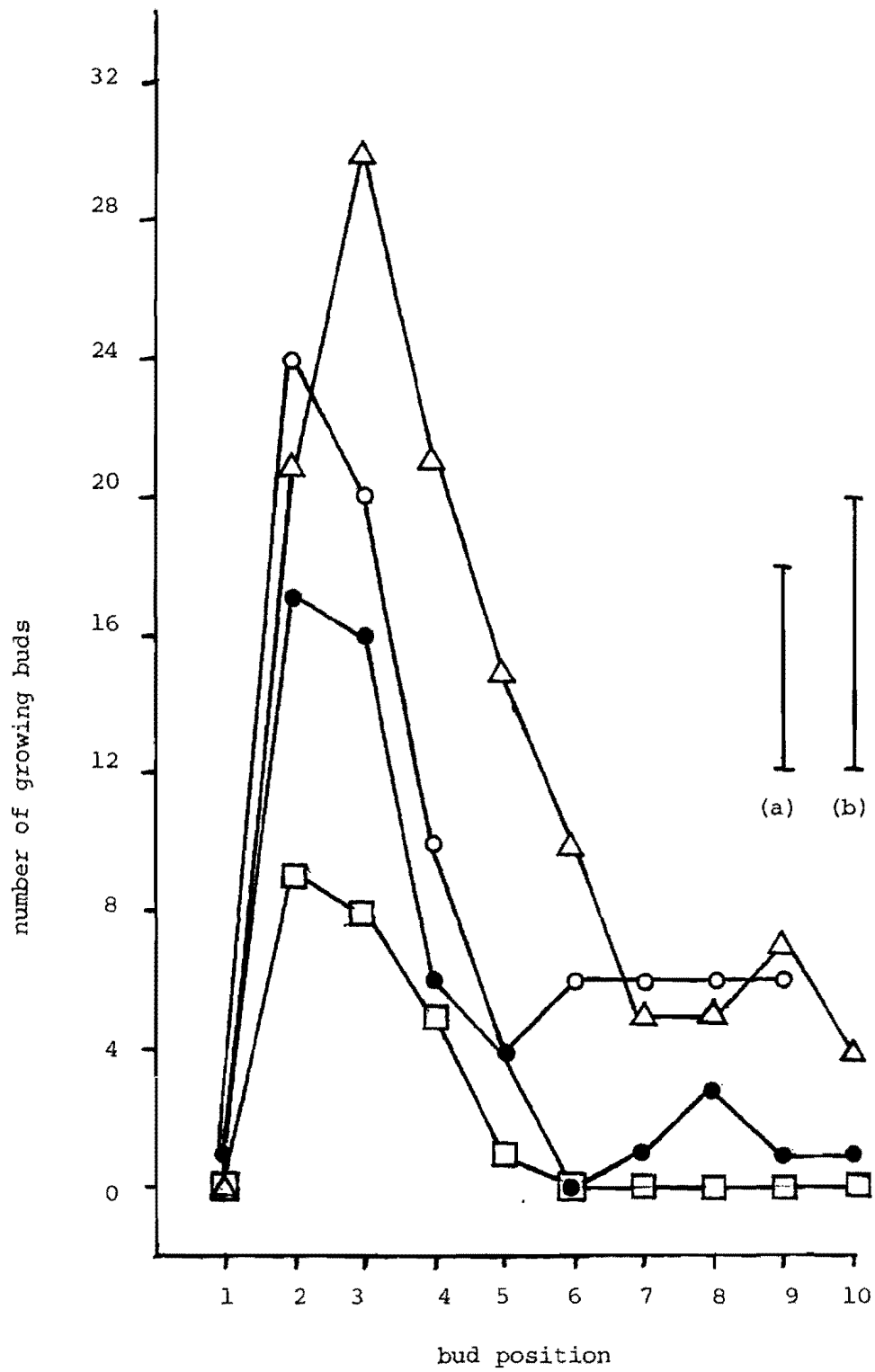
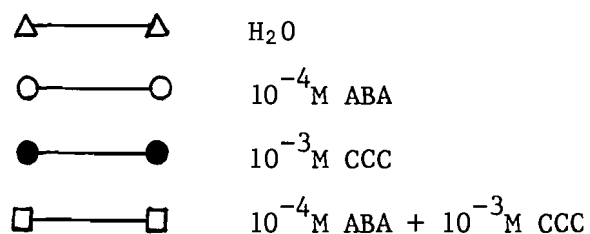


Figure 120. The effect of growth regulator application and (a) the presence and (b) the absence of leaves on bud burst in isolated dormant shoots of *Alnus viridis* (autumn harvest). The values represent the total number of growing buds as a function of position on the stem, with the leaves removed at the beginning of the experiment. Each treatment comprised three replicates, each of 10 shoots. Growth regulators were applied to the base of the shoots. Vertical bars represent the least significant difference at the (a) 5% and (b) 1% level.



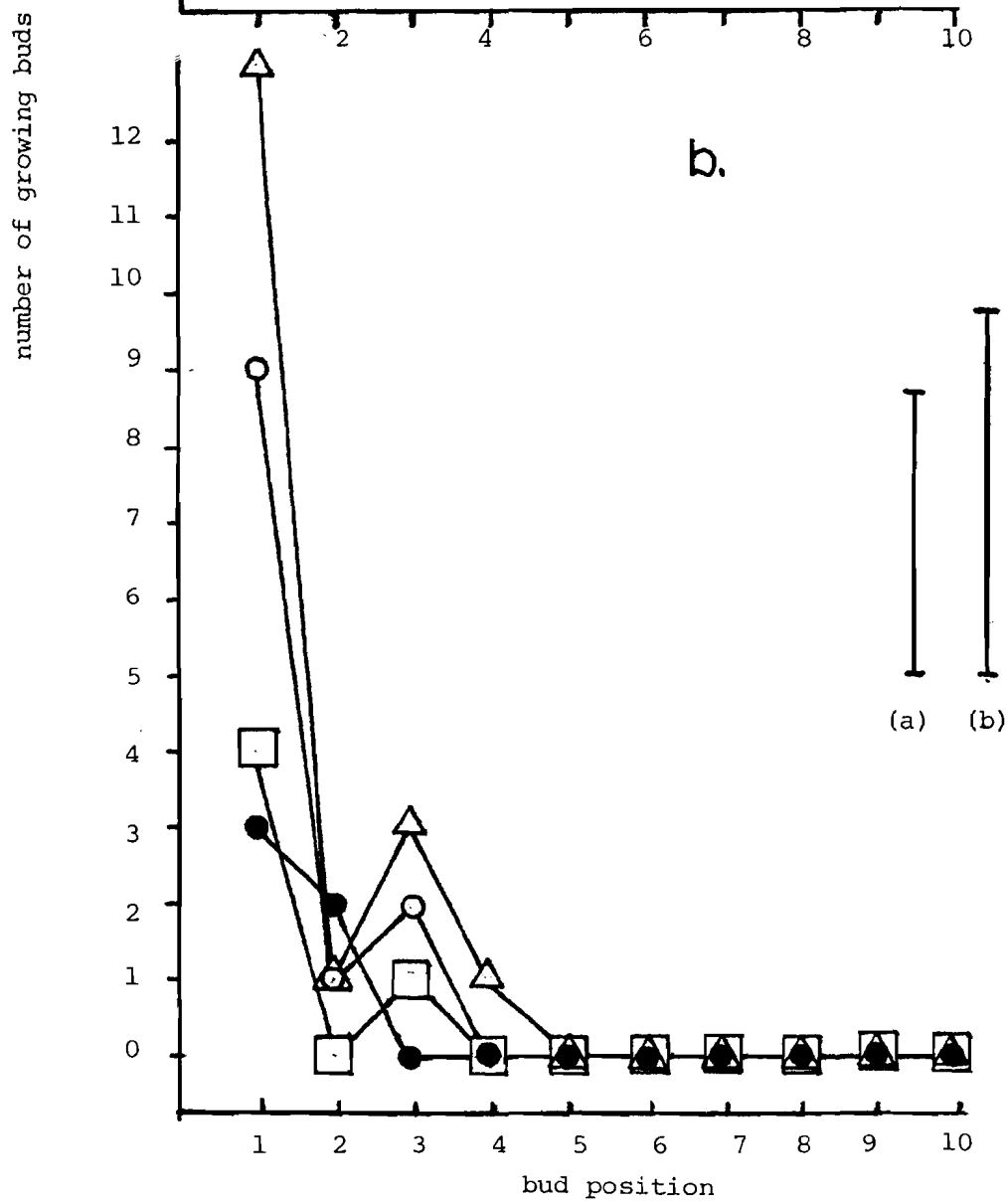
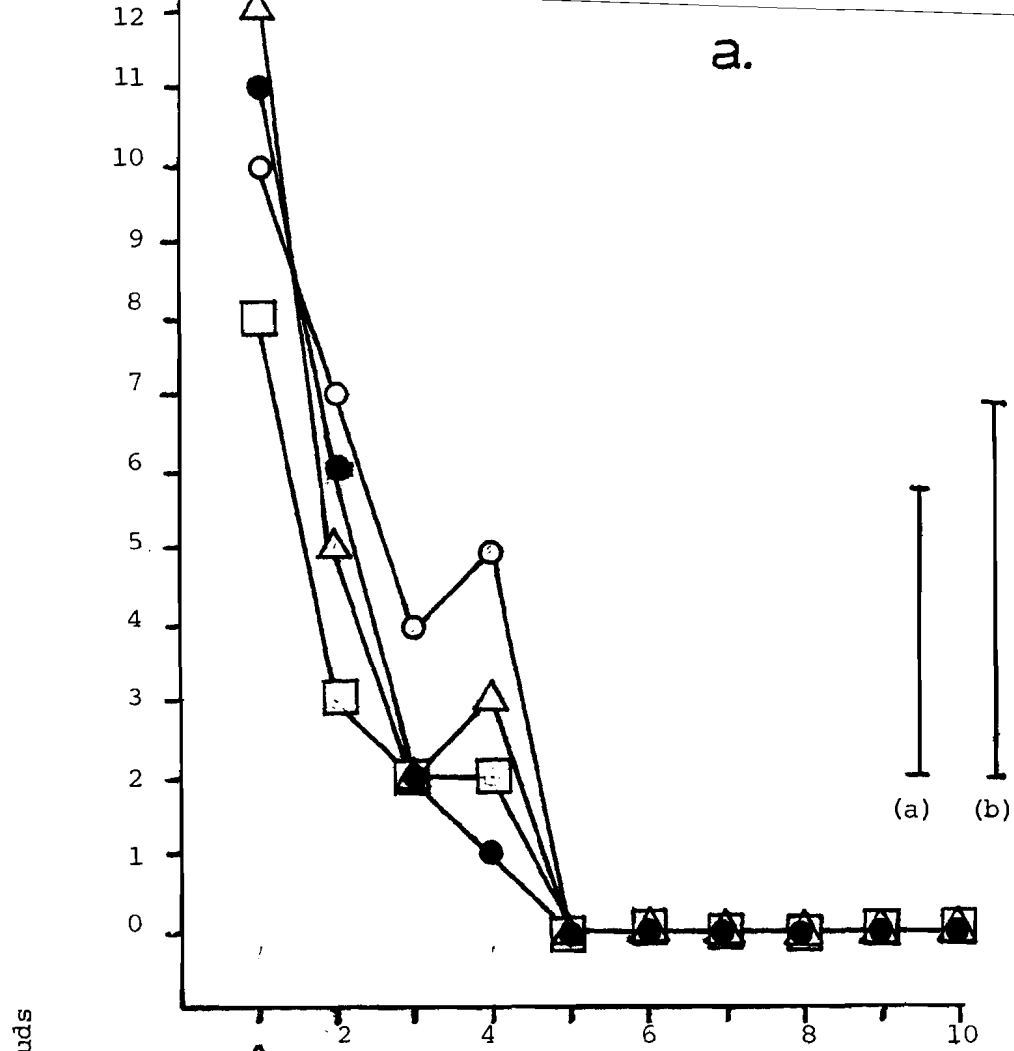
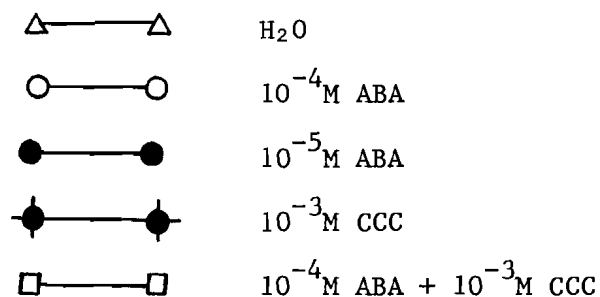


Figure 121. The effect of growth regulator application and (a) the presence and (b) the absence of leaves on bud burst in isolated dormant shoots of *Salix alba/babylonica* (autumn harvest). The values represent the total number of growing buds as a function of position on the stem, with the leaves removed at the beginning of the experiment. Each treatment comprised two replicates, each of 10 shoots. Growth regulator application was to the base of the shoots.



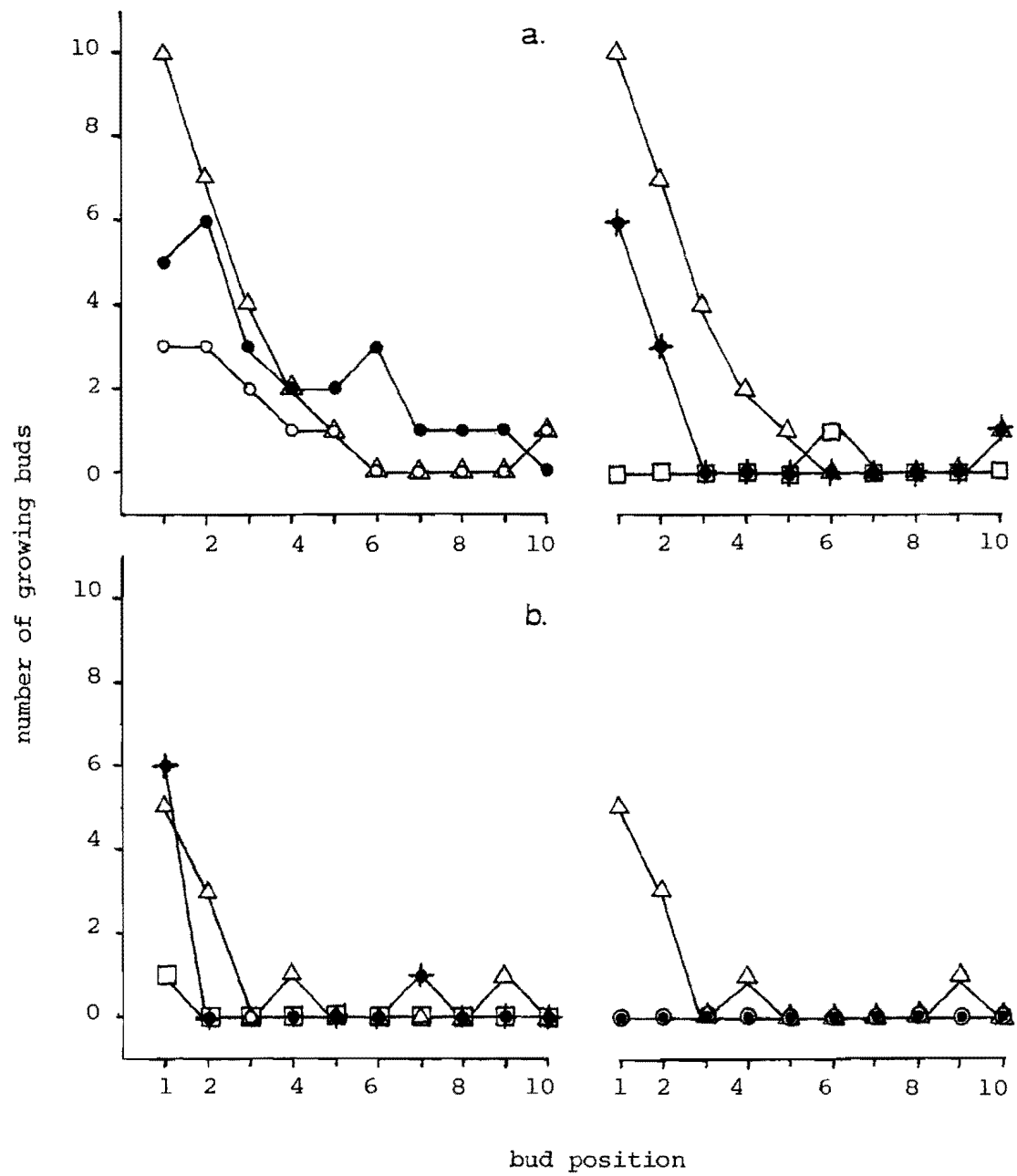
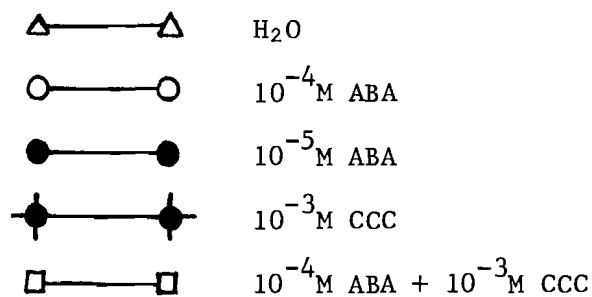


Figure 122. The effect of growth regulator application and the presence of leaves at (a) basal positions and (b) apical positions on bud burst in isolated dormant shoots of *Salix alba/babylonica* (autumn harvest). The values represent the total number of growing buds as a function of position on the stem with the leaves removed at the beginning of the experiment. Growth regulator application was to the base of the shoots. Each treatment comprised two replicates each of 10 shoots.



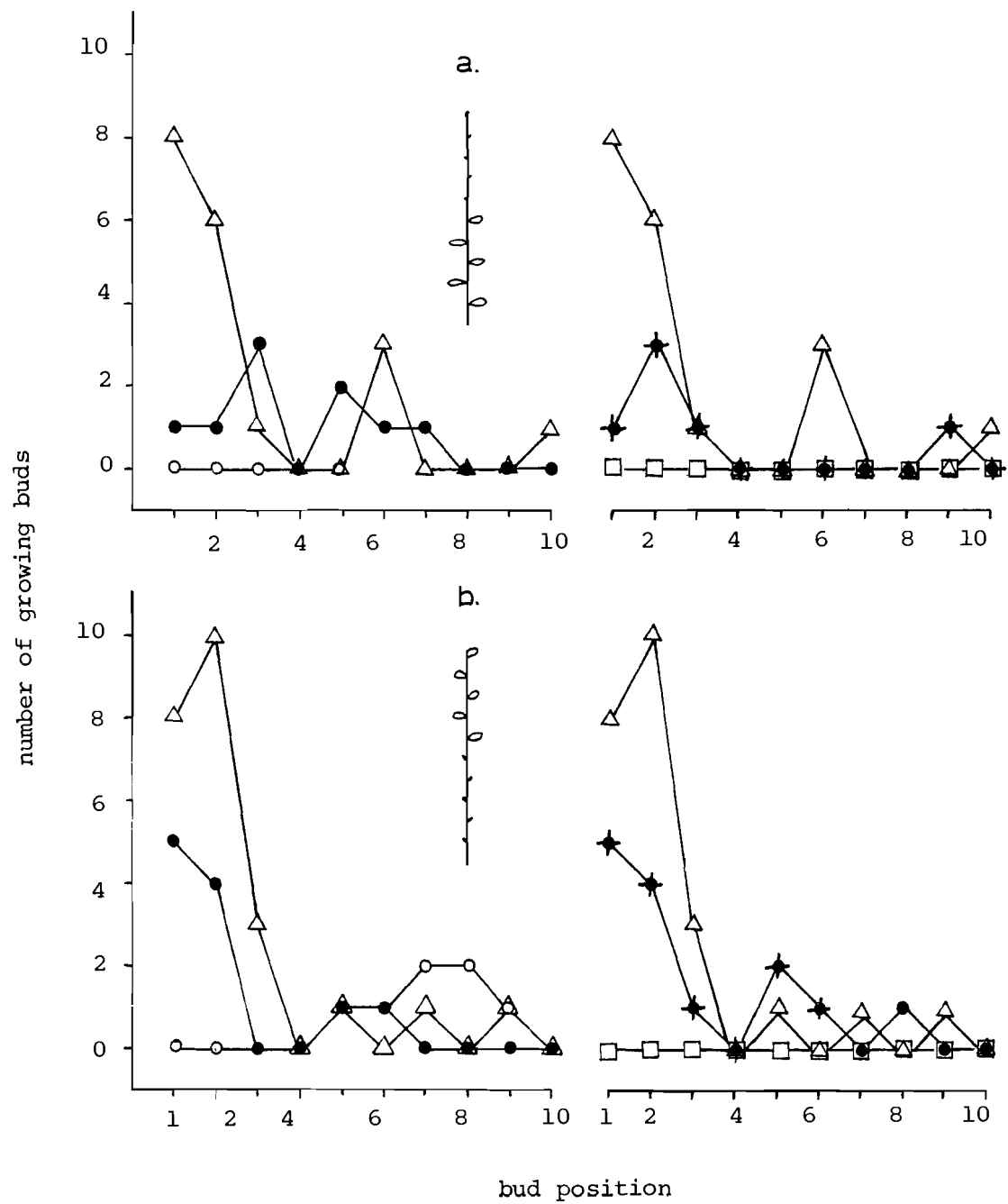
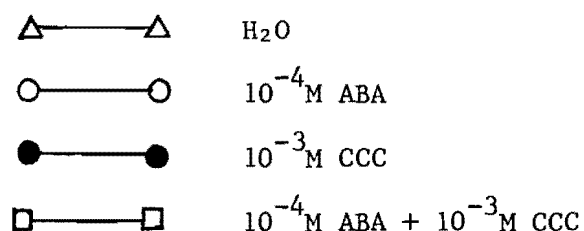


Figure 123. The effect of apical dominance, growth regulator application and (a) the presence and (b) the absence of leaves on bud burst in isolated dormant shoots of *Salix alba/babylonica* (autumn harvest). The values represent the total number of growing buds as a function of position on the stem. Leaves were removed and the apical and basal sections were separated at the beginning of the experiment. Each treatment comprised two replicates, each of 10 shoots. Growth regulator application was to the base of the shoots.



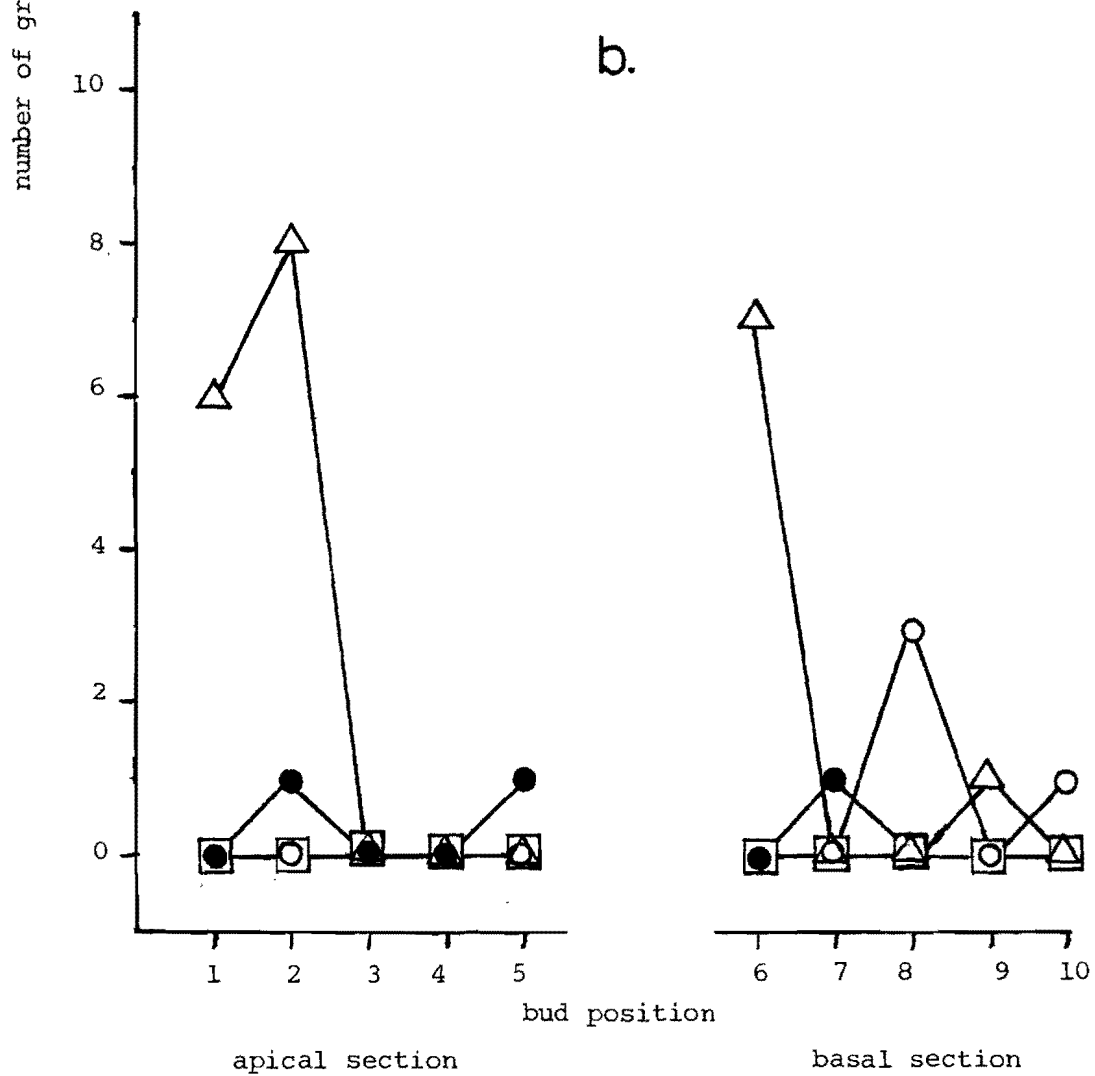
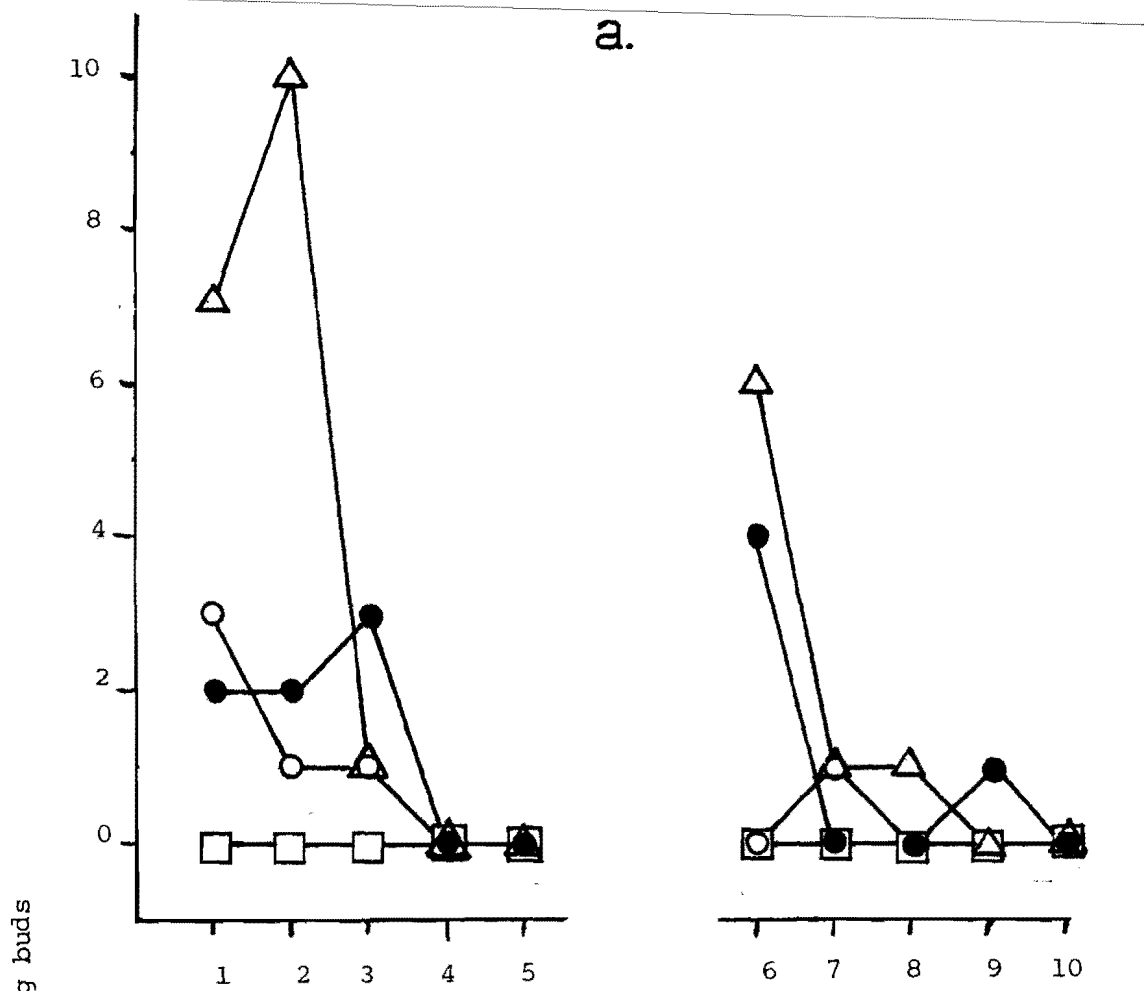
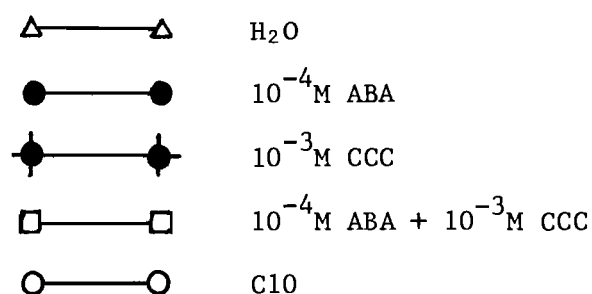


Figure 124. The effect on bud burst of growth regulator application to the base of isolated dormant shoots of *Salix alba/babylonica* (autumn harvest). The values represent the total number of growing buds as a function of position on the stem. Each treatment comprised two replicates, each of 10 shoots. Leaves were removed at the beginning of the experiment.



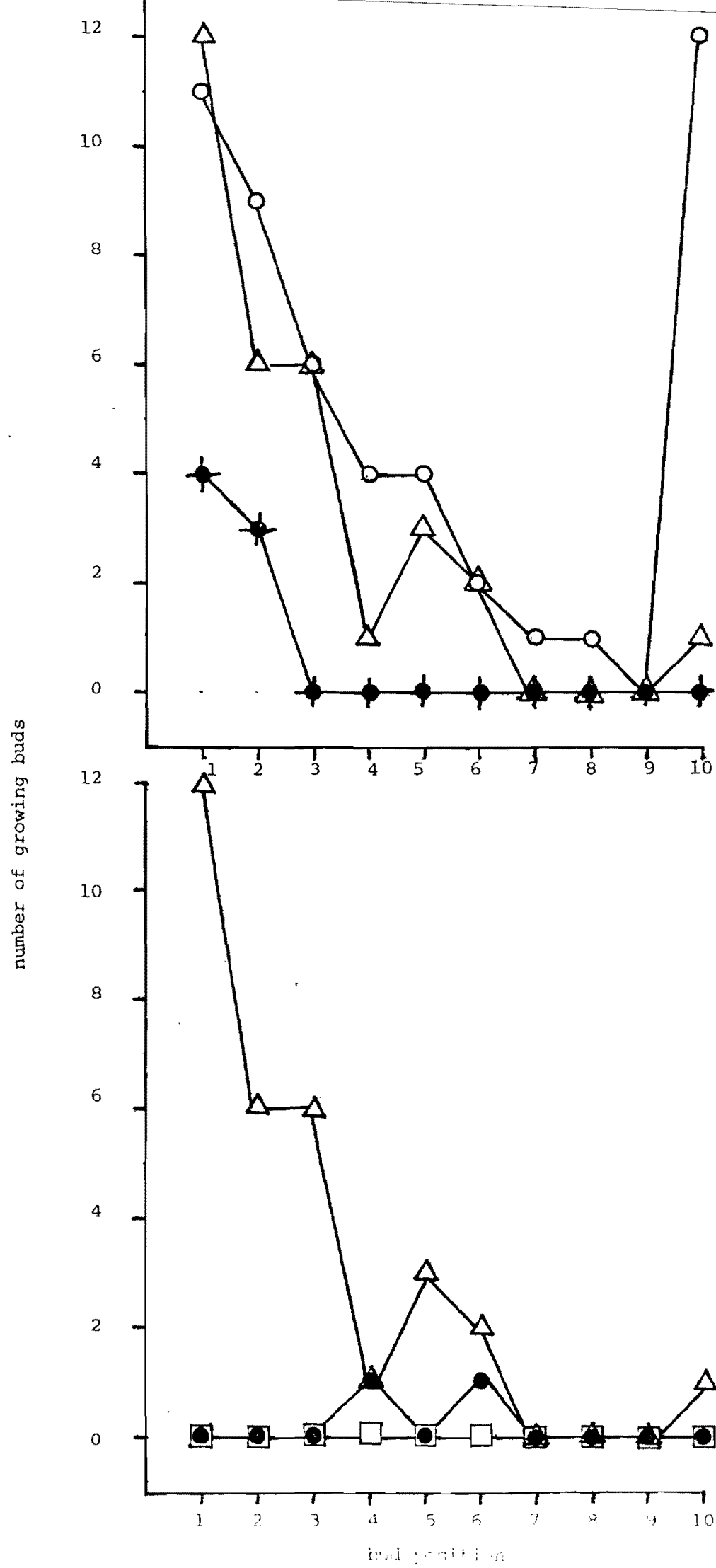


Figure 125. In vitro *Populus yunnanensis* shoot tip assay of the serial dilution of an acidic ether-soluble extract of *Alnus glutinosa* leaves harvested on 4 April 1979 (H3). The equivalent of (a) 0.0, (b) 0.25, (c) 0.5 and (d) 1.0 g DW of leaf material was assayed and the chromatographs were developed in isopropanol:NH₃:H₂O (10:1:1::v:v:v) solvent. Each R_F section was assayed on one shoot tip and each histogram represents the mean \pm S.E. of three replicate chromatographs. The column on the right represents fresh weight of shoots in the absence of a chromatography section. Number in brackets over histogram bars represents one S.E.

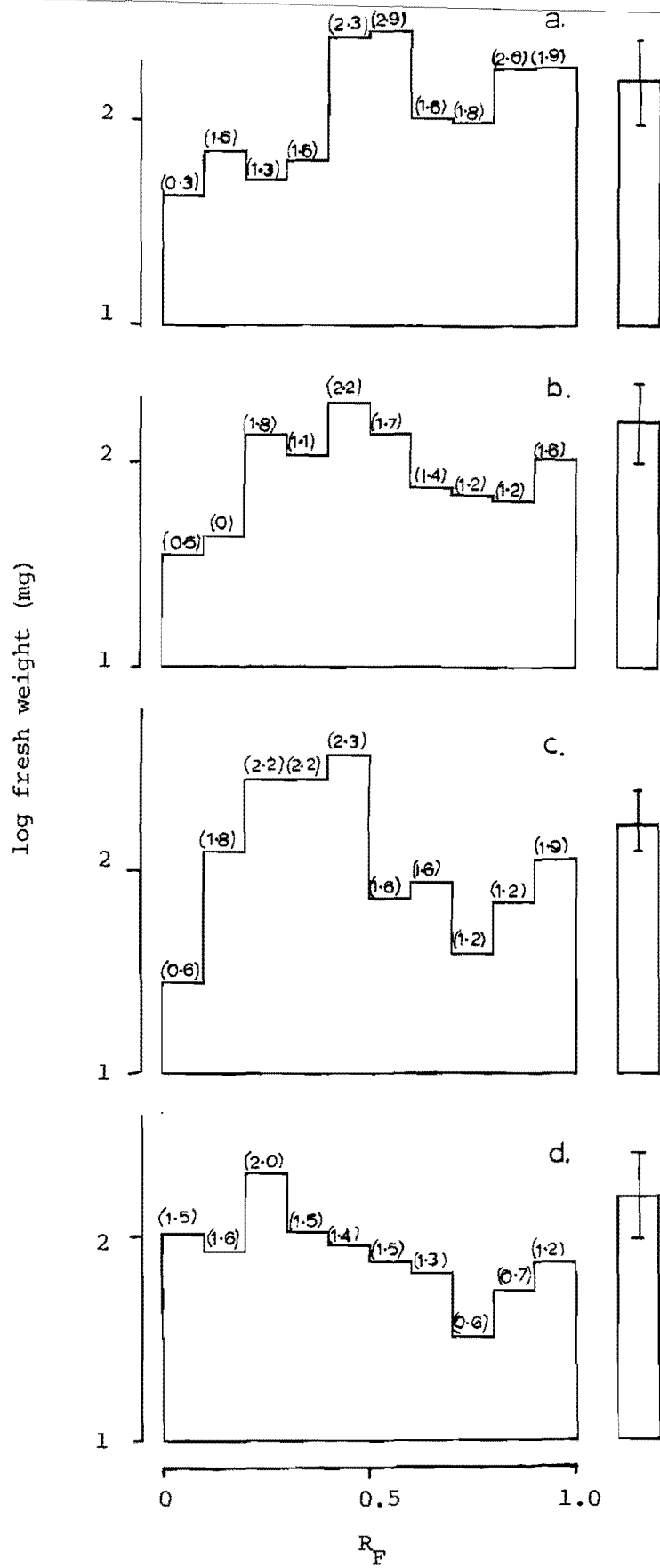


Figure 126. In vitro *Populus yunnanensis* shoot tip assay of the serial dilution of an acidic ether-soluble extract of *Alnus glutinosa* leaves harvested on 4 April 1979 (H3). The equivalent of (a) 0.0, (b) 0.25, (c) 0.5 and (d) 1.0 g DW of leaf material was assayed and the chromatographs were developed in isopropanol:NH₃:H₂O (10:1:1::v:v:v) solvent. Vertical bars represent twice the S.E. of the mean, and the column on the right represents length of shoots in the absence of a chromatography section. Other details as in figure 125.

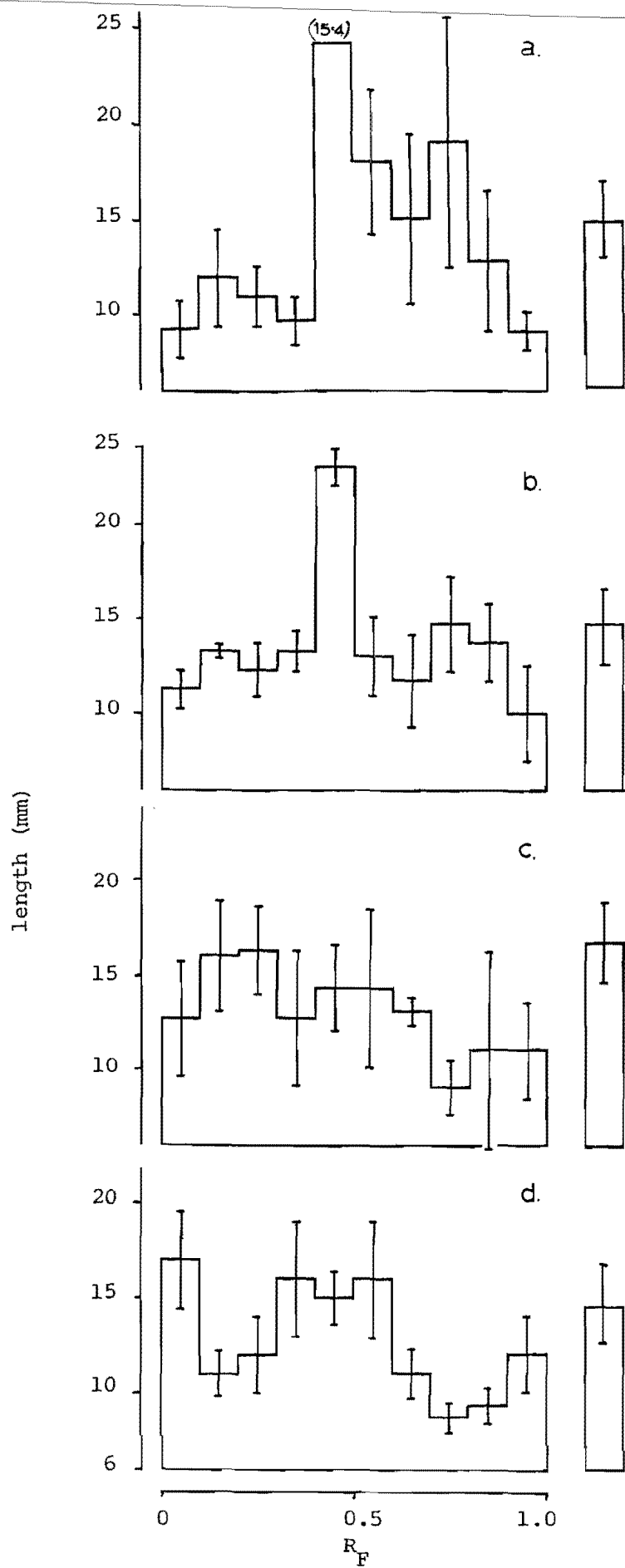


Figure 127. In vitro *Populus yunnanensis* shoot tip assay of the serial dilution of an acidic ether-soluble extract of *Alnus viridis* leaves harvested on day 7 of SD treatment (H2). The equivalent of (a) 0.0, (b) 0.25, (c) 0.5 and (d) 1.0 g DW of leaf material was assayed and the chromatographs were developed in isopropanol:NH₃:H₂O (10:1:1::v:v:v) solvent. Other details as in figure 125.

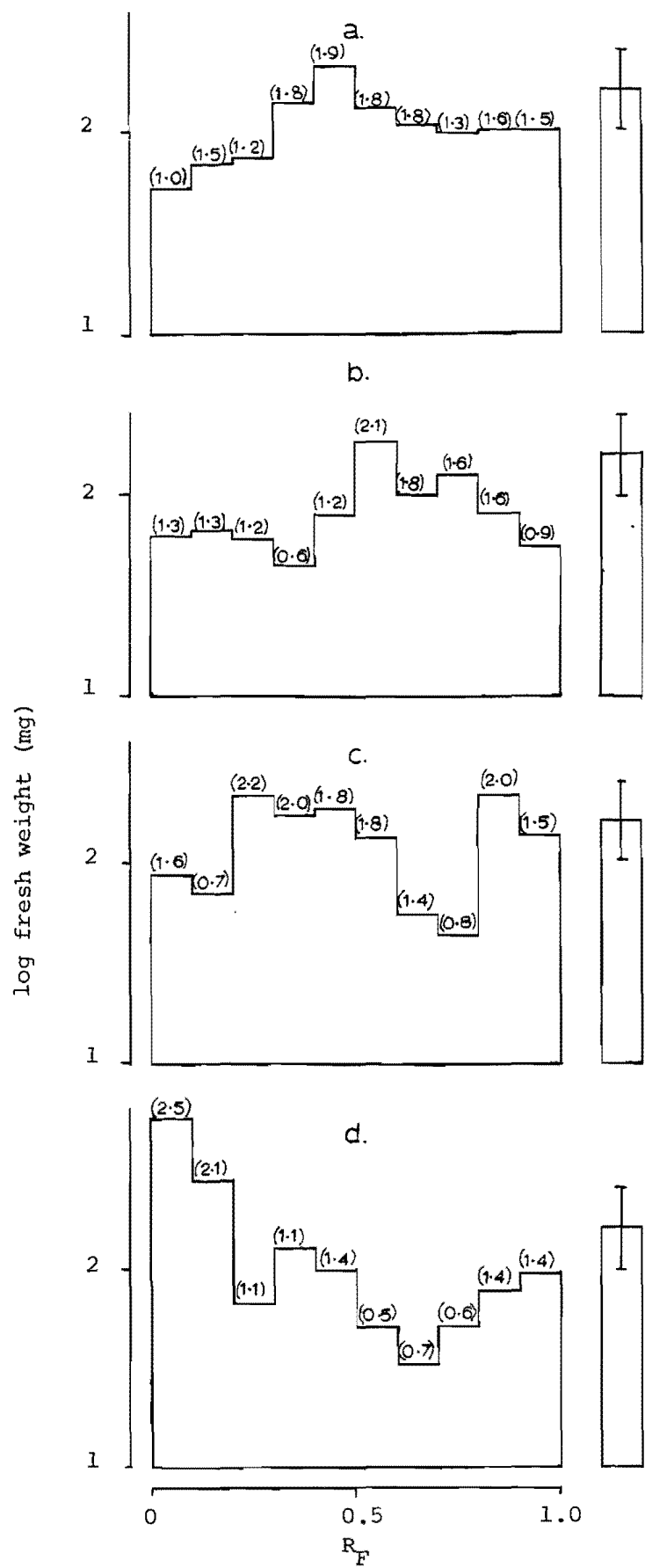


Figure 128. In vitro *Populus yunnanensis* shoot tip assay of the serial dilution of an acidic ether-soluble extract of *Alnus viridis* leaves harvested on day 7 of SD treatment (H2). The equivalent of (a) 0.0, (b) 0.25, (c) 0.5 and (d) 1.0 g DW of leaf material was assayed and the chromatographs were developed in isopropanol:NH₃:H₂O (10:1:1::v:v:v) solvent. Other details as in figure 126.

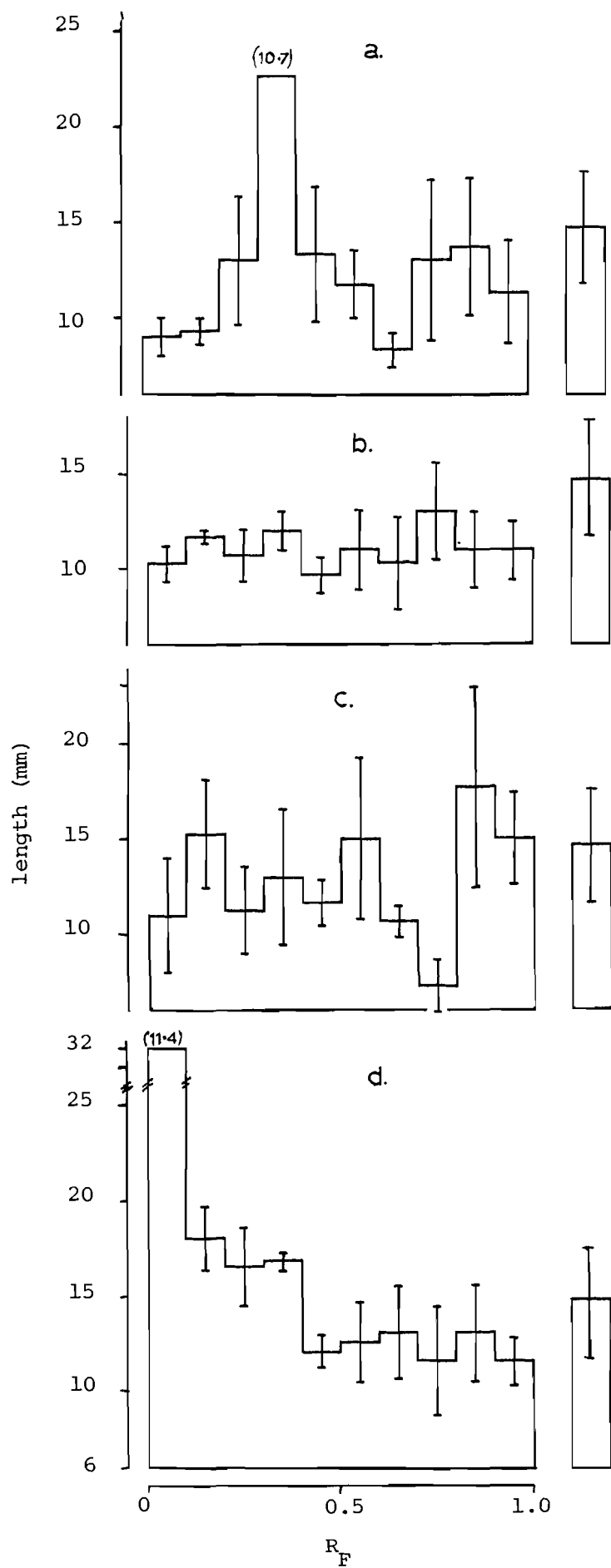


Figure 129. In vitro *Populus yunnanensis* shoot tip assay of the serial dilution of an acidic ether-soluble extract of *Alnus glutinosa* apices harvested on 4 April 1979 (H3). The equivalent of (a) 0.0, (b) 0.25, (c) 0.5 and (d) 1.0 g DW of apical material was assayed and the chromatographs were developed in isopropanol:NH₃:H₂O (10:1:1::v:v:v) solvent. Other details as in figure 125.

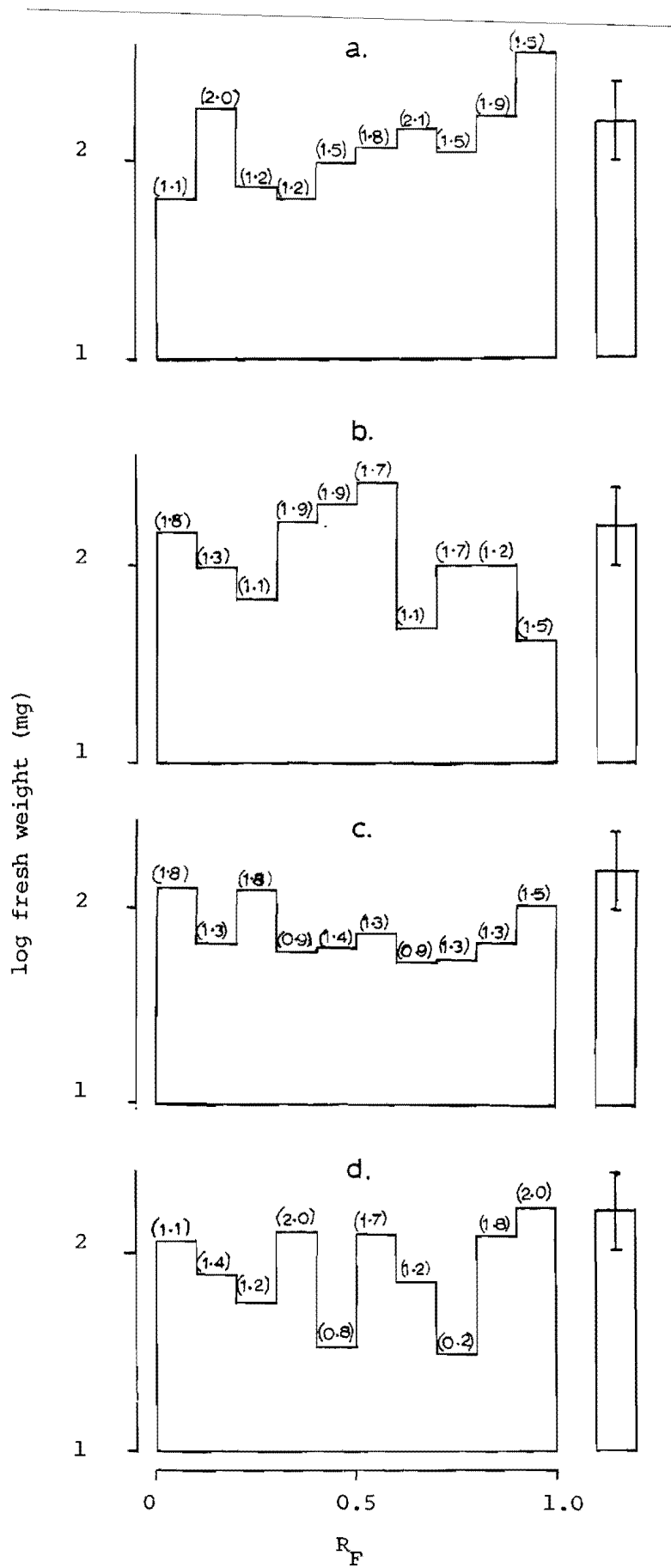


Figure 130. In vitro *Populus yunnanensis* shoot tip assay of the serial dilution of an acidic ether-soluble extract of *Alnus glutinosa* apices harvested on 4 April 1979 (H3). The equivalent of (a) 0.0, (b) 0.25, (c) 0.5 and (d) 1.0 g DW of apical material was assayed and the chromatographs were developed in isopropanol:NH₃:H₂O (10:1:1::v:v:v) solvent. Other details as in figure 126.

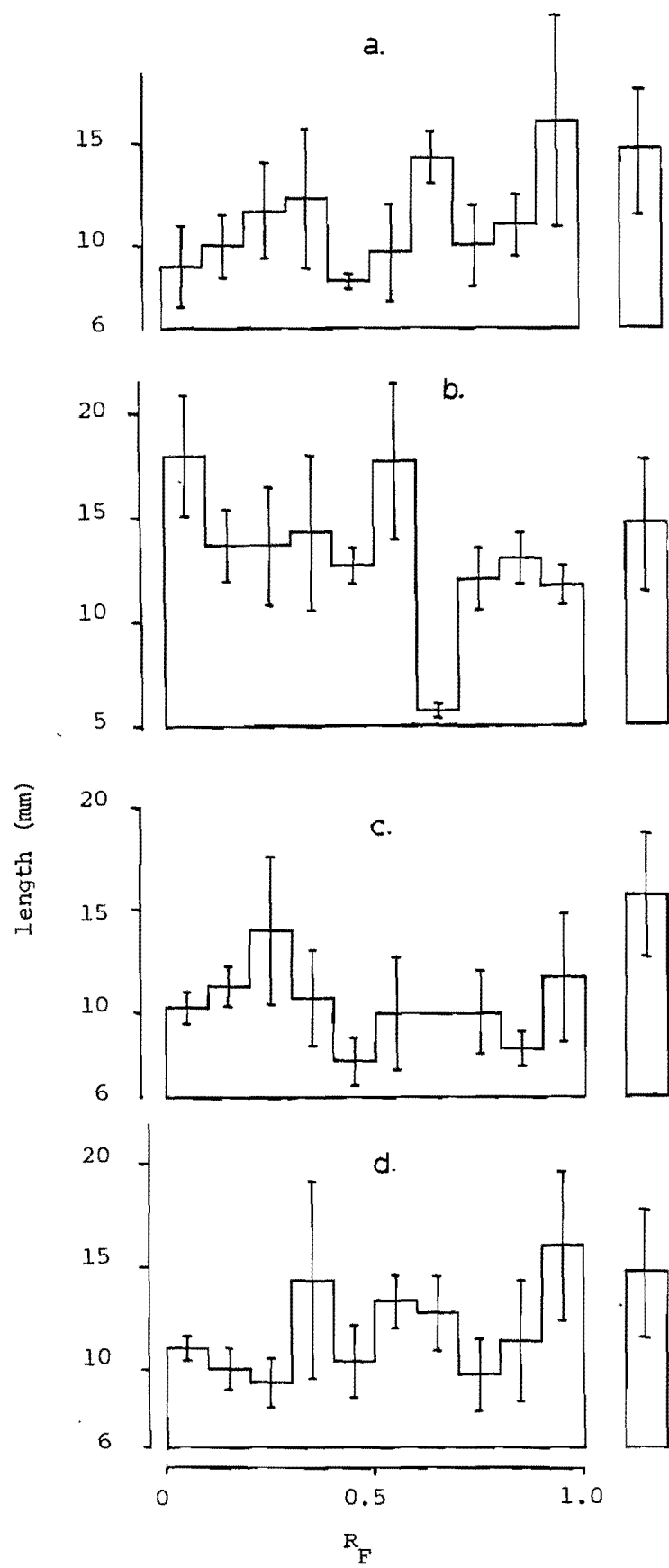


Figure 131. Effect of serial dilutions of crude extract on (a) fresh weight and (b) length of *Populus yunnanensis* shoots culture in vitro. Crude extract was obtained from the aqueous fraction of an 80% MeOH extract of *P. yunnanensis* shoots subjected to SD treatment. Apparently single shoots were cultured on medium 2 containing extract and were maintained under LD photoperiods at 25⁰C for 4 weeks. Each dilution was tested on twenty shoots and the vertical bars represent twice the S.E. of the mean.

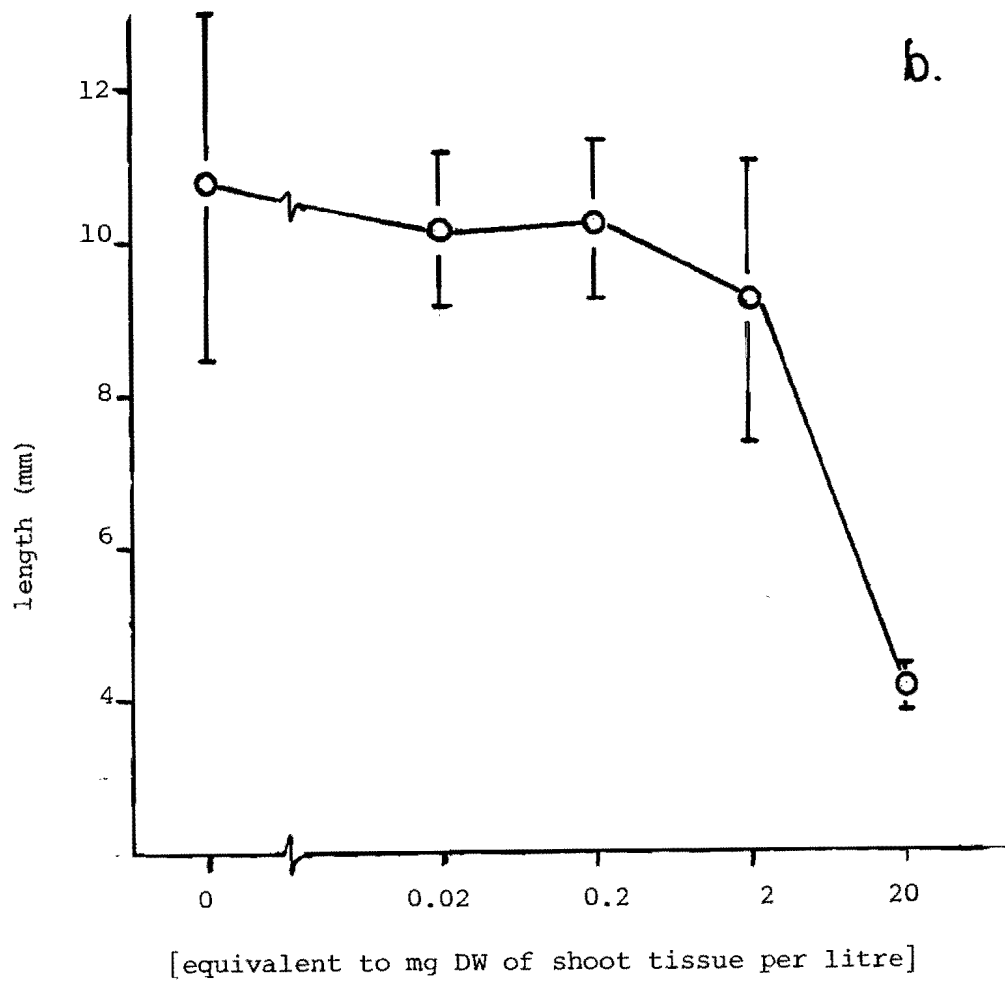
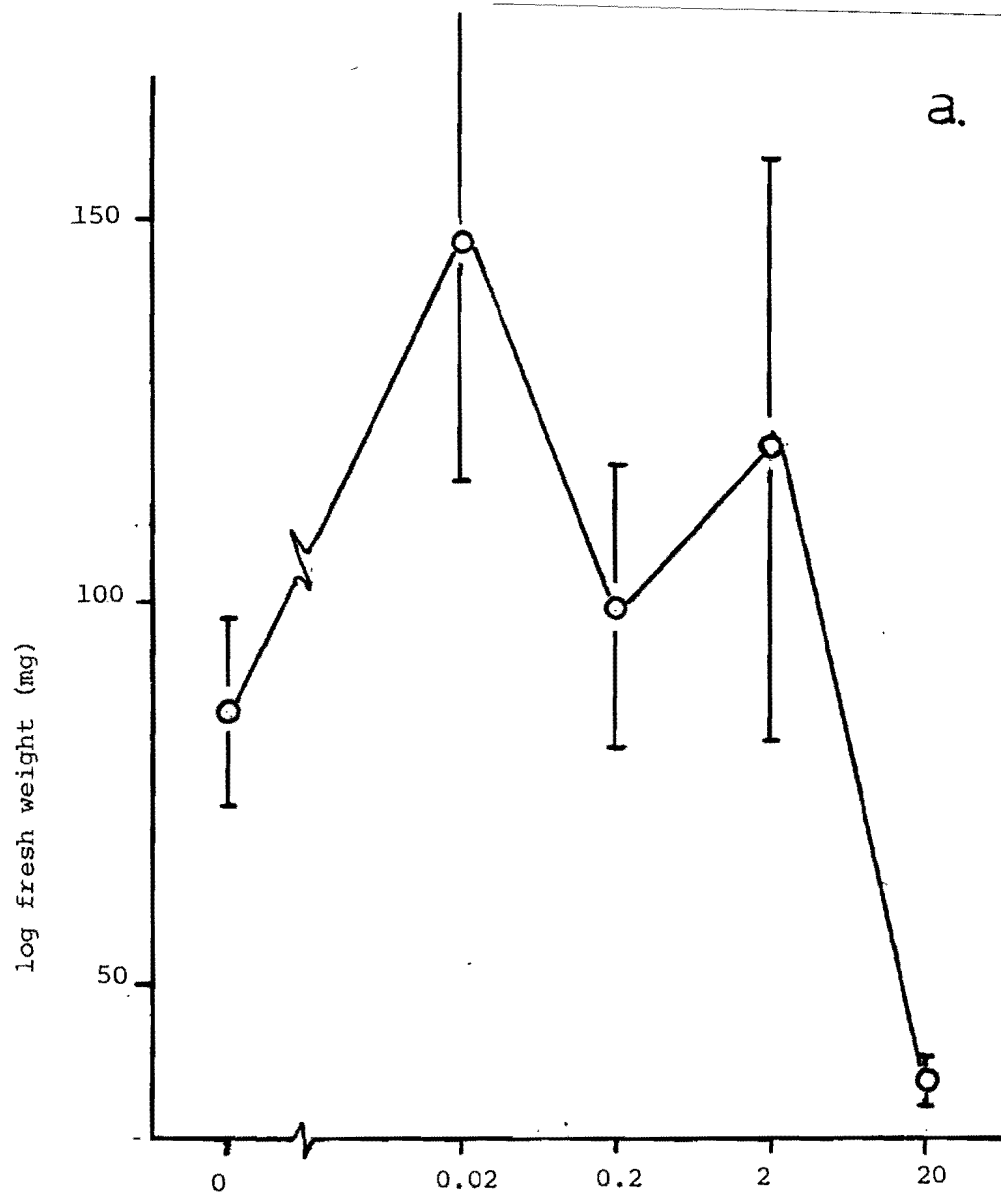


Figure 132. Effect of serial concentrations of ABA on (a) fresh weight and (b) length of *Populus yunnanensis* shoots cultured in vitro. Apparently single shoots were cultured on medium 1 (●) or medium 2 (○, △, □, ▲, ■) containing ABA and maintained under LD (16 hour) photoperiods at 20°C for 4 weeks. The experiment was repeated five times using medium 2 and each value represents the mean \pm S.E. of twenty shoots. The vertical bars show one S.E. above and/or below the lines.

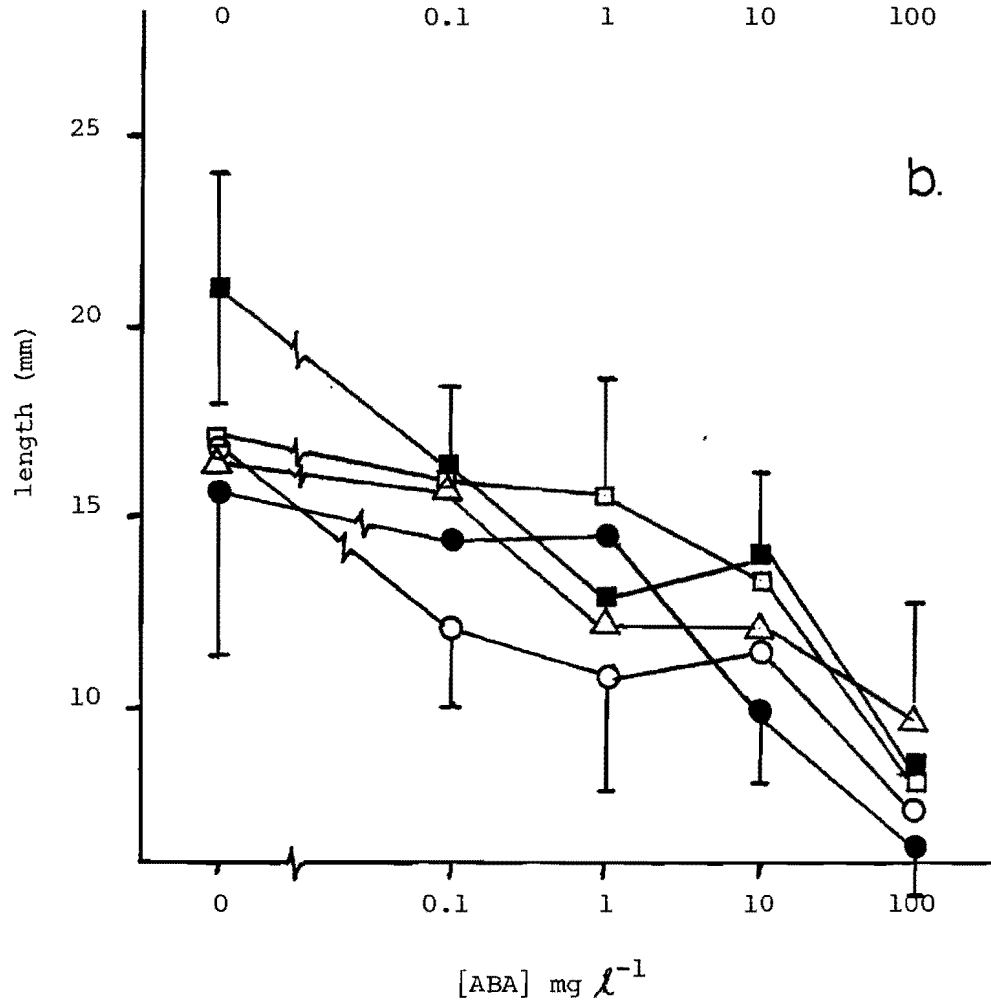
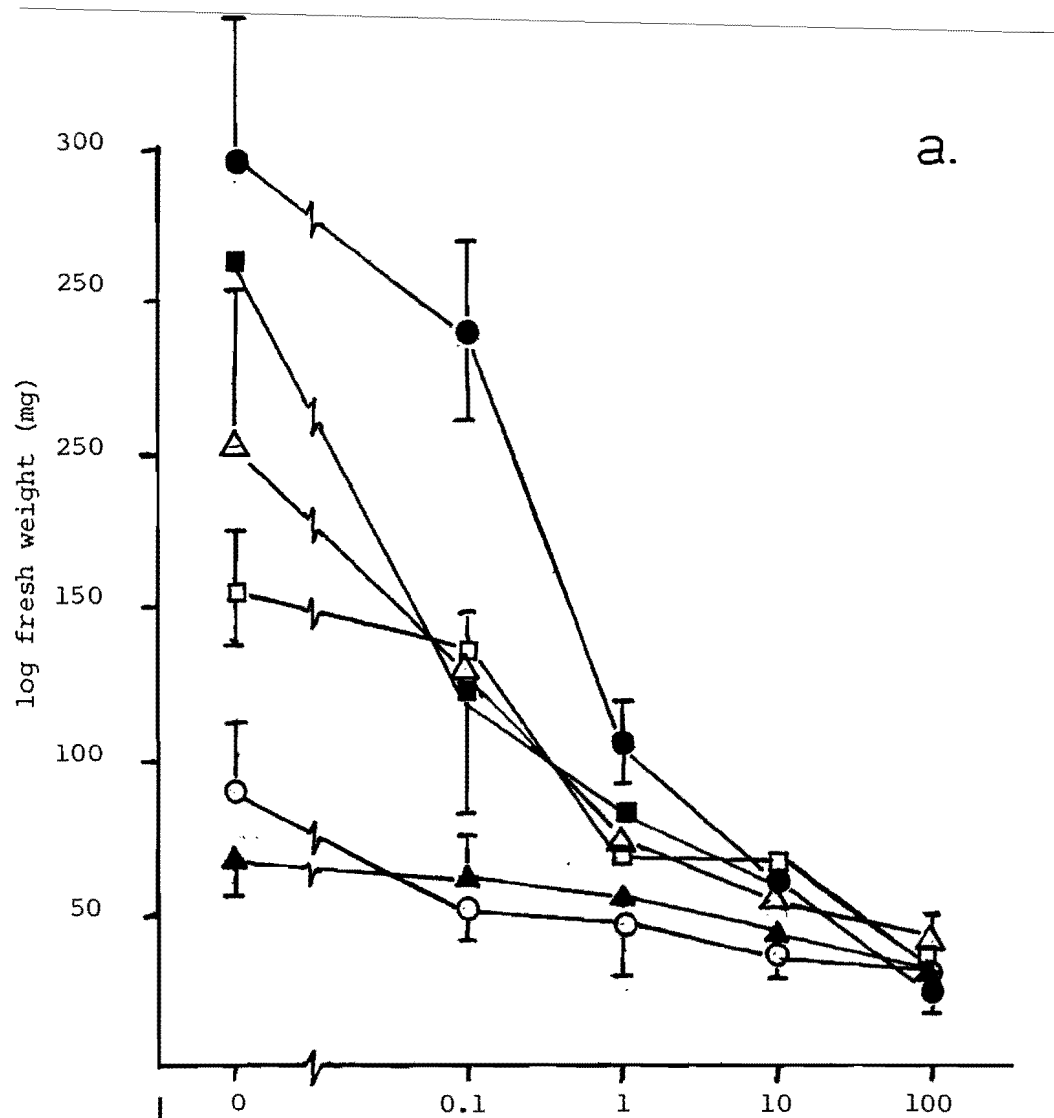


Figure 133. U.V. spectra of (a) abscisic acid in EtoAc and (b) aqueous ABA before and after autoclaving. The mixed isomers of authentic ABA were added to the solution containing inorganic and organic nutrients and re-extracted into EtoAc at pH 3. The aqueous solution was made up with the mixed isomers of ABA. λ_{max} was 248 nm.

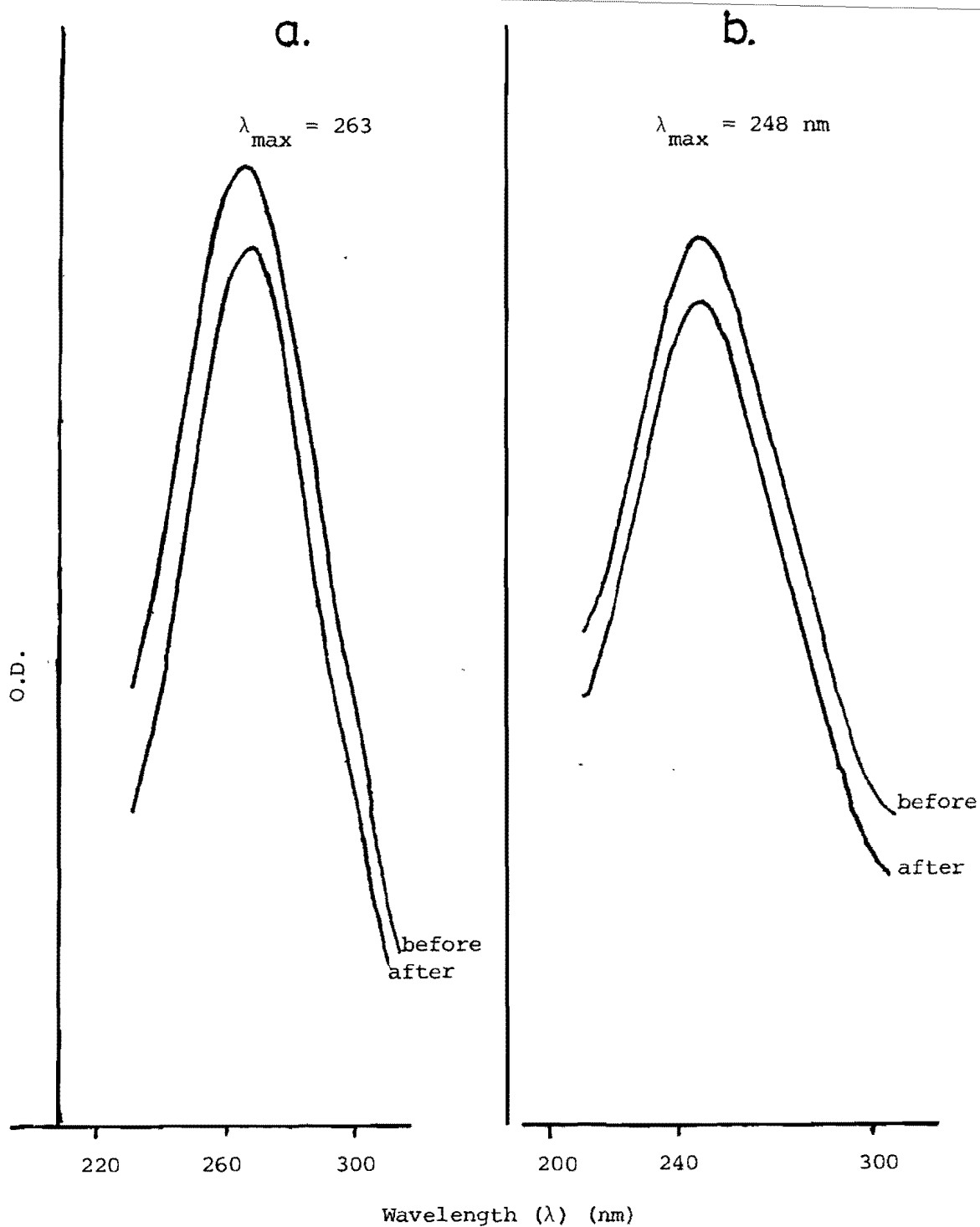


Figure 134. Effect of serial concentrations of AMO 1618 on (a) fresh weight and (b) length of *Populus yunnanensis* shoots cultured in vitro. Apparently single shoots were cultured on medium 2 containing AMO 1618 and maintained under LD photoperiods at 25°C for 4 weeks. The experiment was repeated twice and each value represents the mean \pm S.E. of twenty shoots. The vertical bars show one S.E. above and/or below the lines.

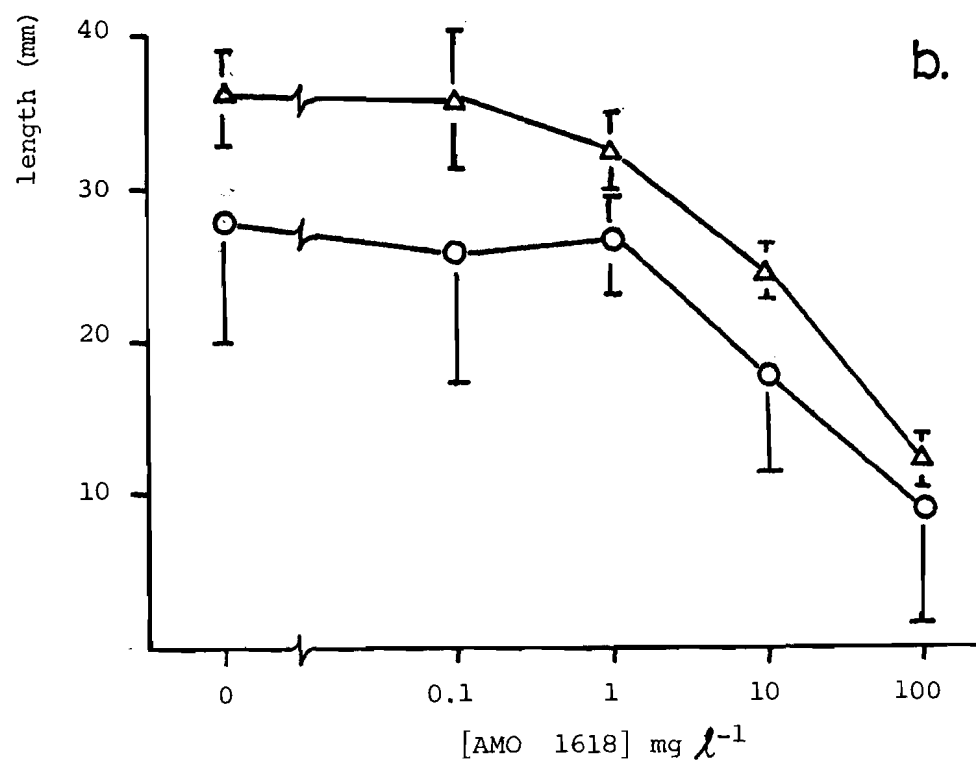
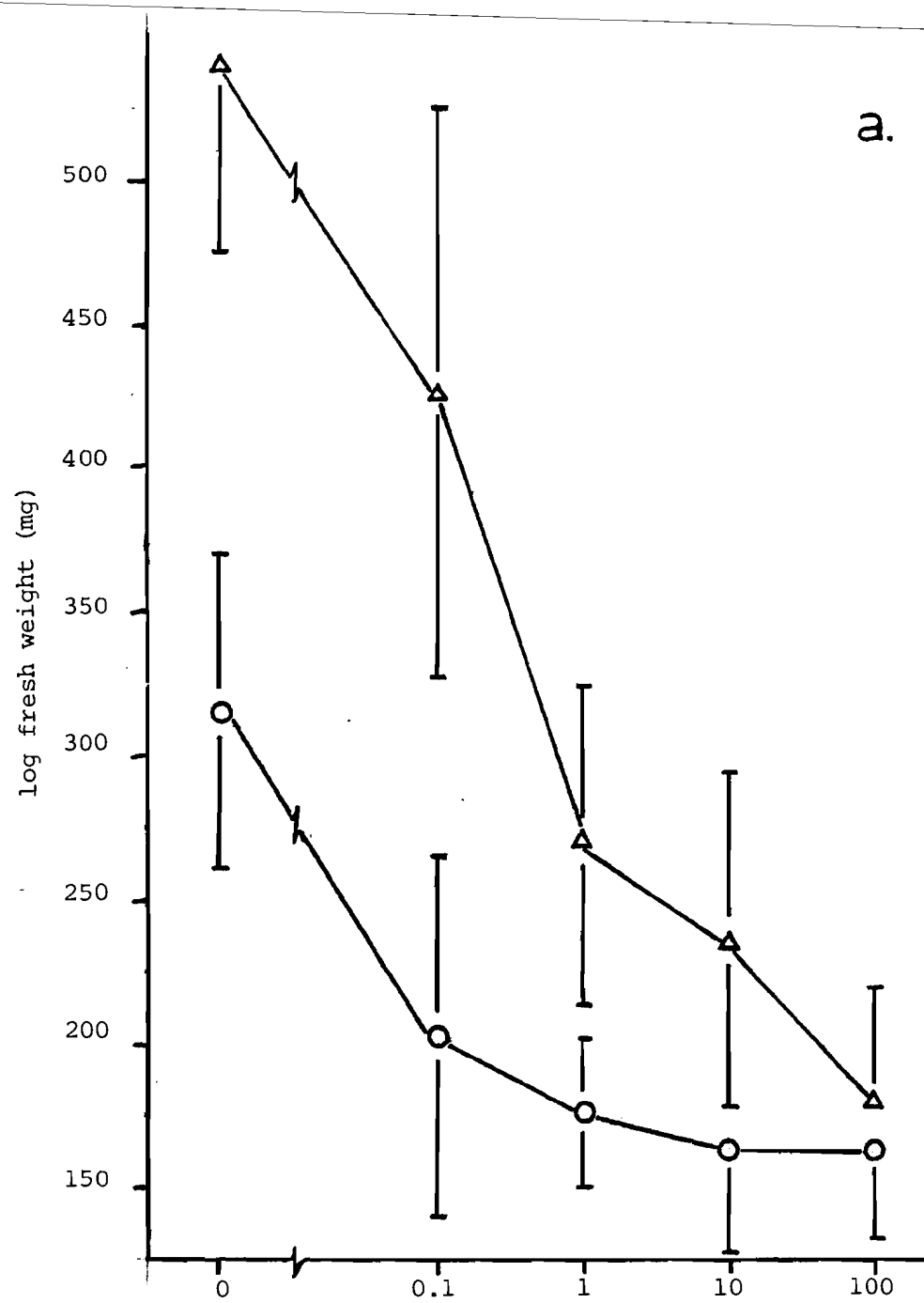


Figure 135. Effect of serial concentrations of ABA + AMO 1618 on (a) fresh weight and (b) length of *Populus yunnanensis* shoots cultured in vitro. Apparently single shoots were cultured on medium 2 containing ABA + AMO 1618 and maintained under LD photoperiods at 25°C for 4 weeks. The experiment was repeated twice and each value represents the mean \pm S.E. of twenty shoots. The vertical bars show one S.E. above and/or below the lines.

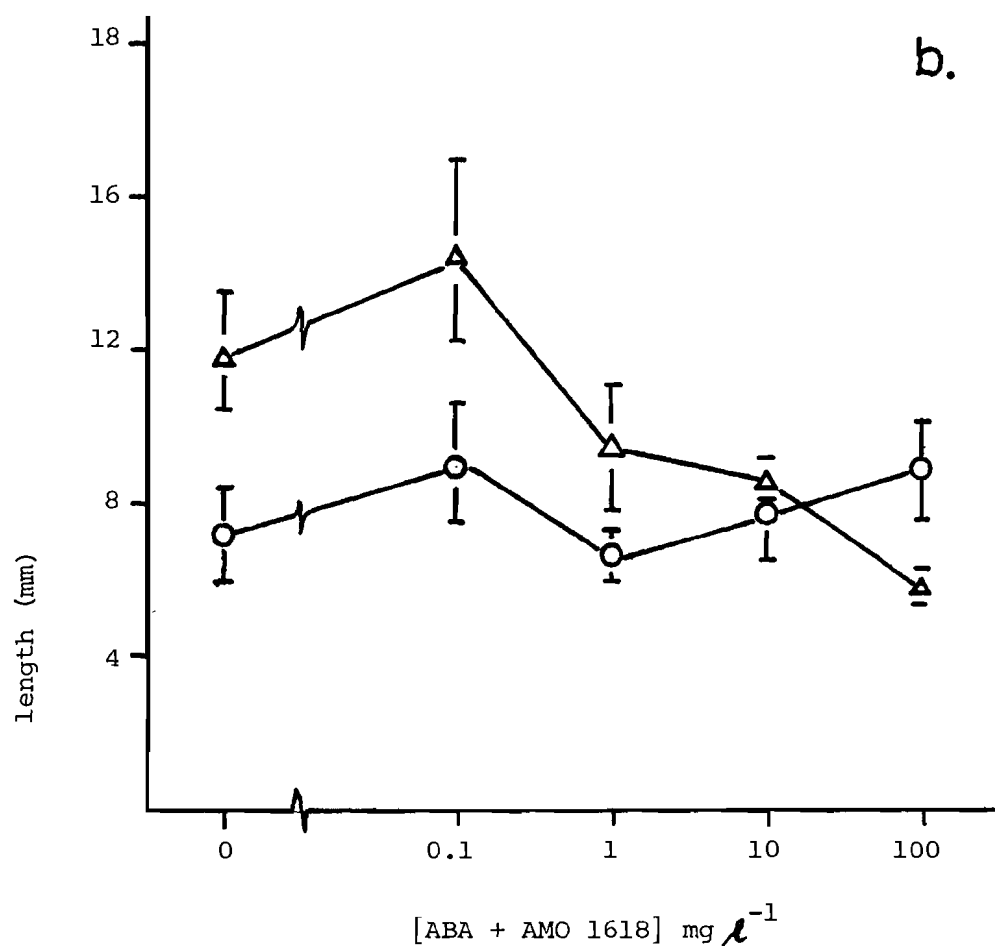
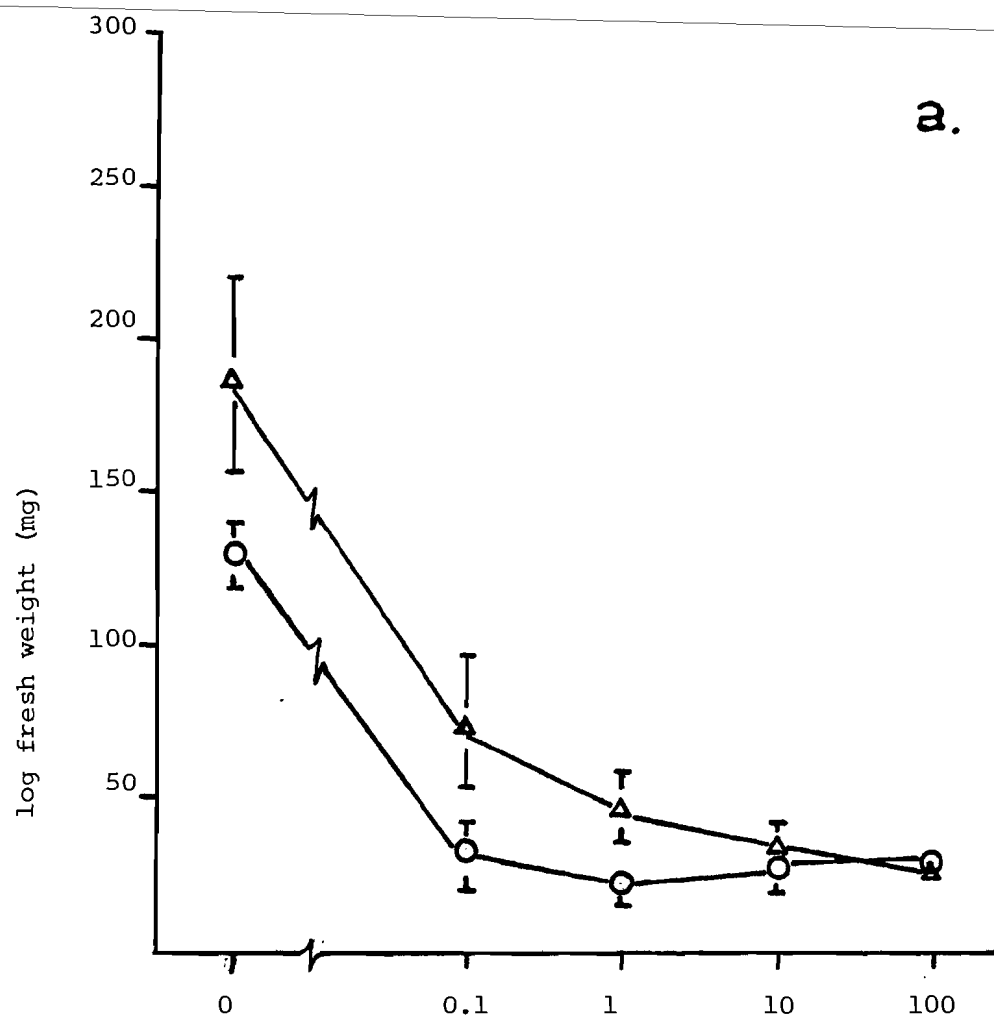


Figure 136. Effect of serial concentrations of GA₃ on (a) fresh weight and (b) length of *Populus yunnanensis* shoots cultured in vitro. Apparently single shoots were cultured on medium 2 containing GA₃ and maintained under LD photo-periods at 25°C for 4 weeks. The experiment was repeated twice and each value represents the mean \pm S.E. of twenty shoots. The vertical bars show one S.E. above and/or below the lines.

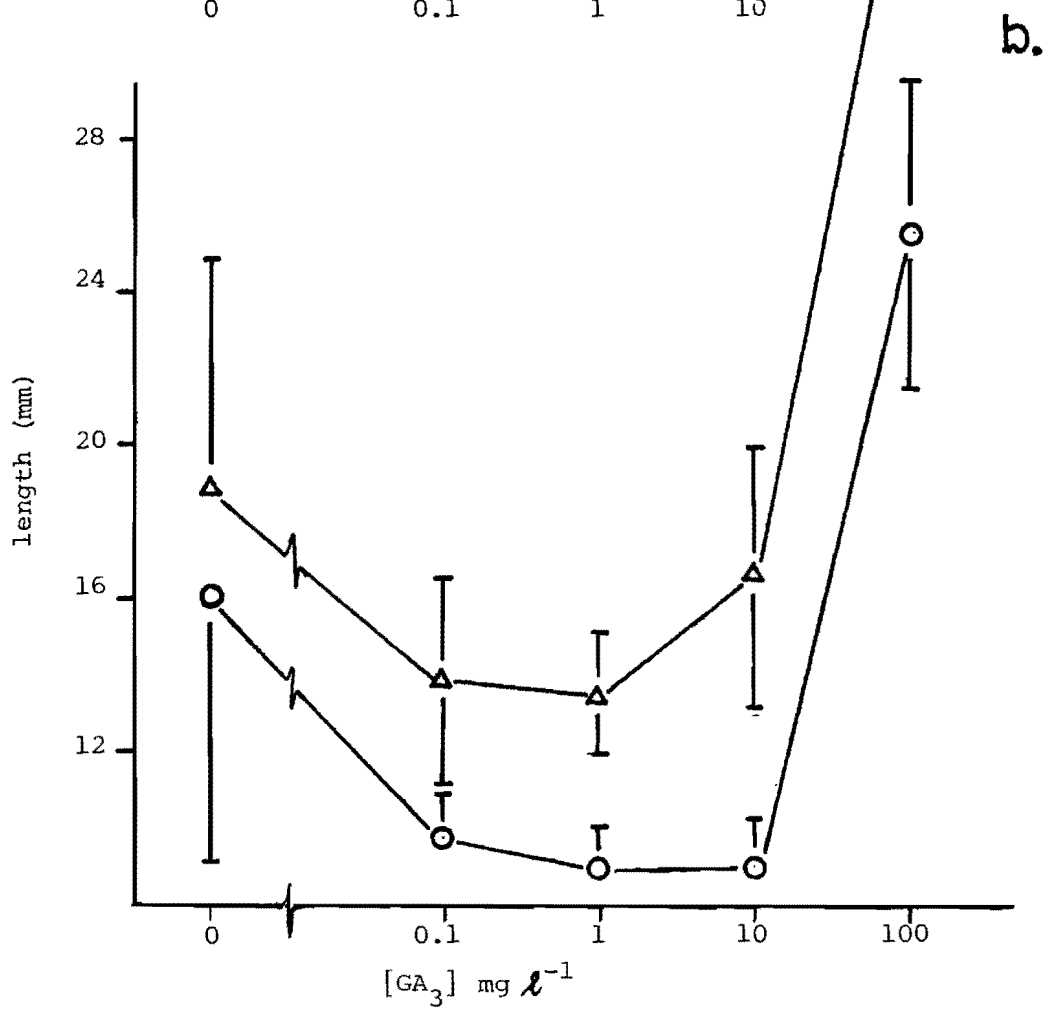
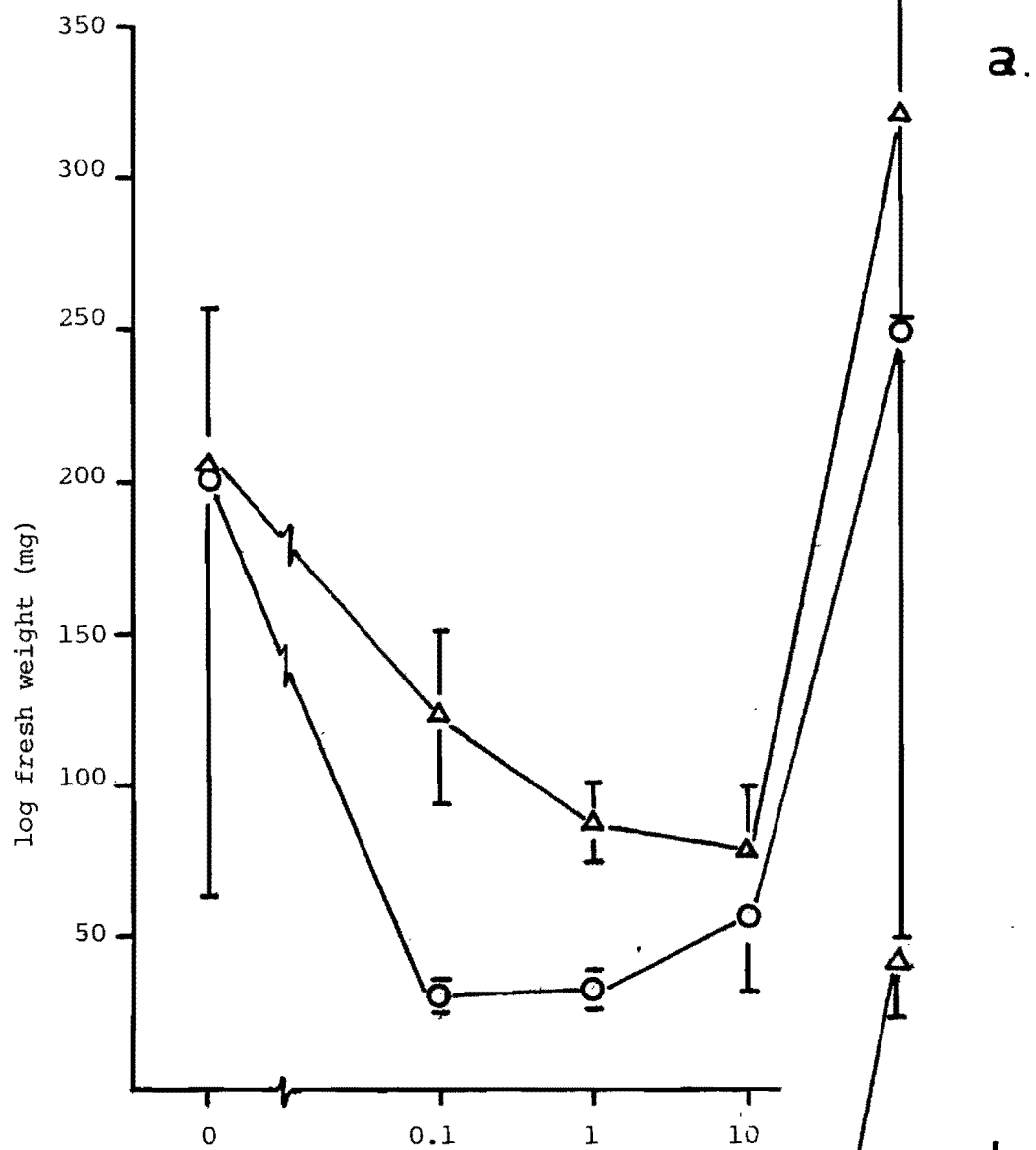


Figure 137. Effect of serial concentrations of C5 on (a) fresh weight and (b) length of *Populus yunnanensis* shoots cultured in vitro. Apparently single shoots were cultured on medium 2 containing C5 and maintained under LD photoperiods at 25°C for 4 weeks. The experiment was repeated twice and each value represents the mean \pm S.E. of twenty shoots. The vertical bars show one S.E. above and/or below the lines.

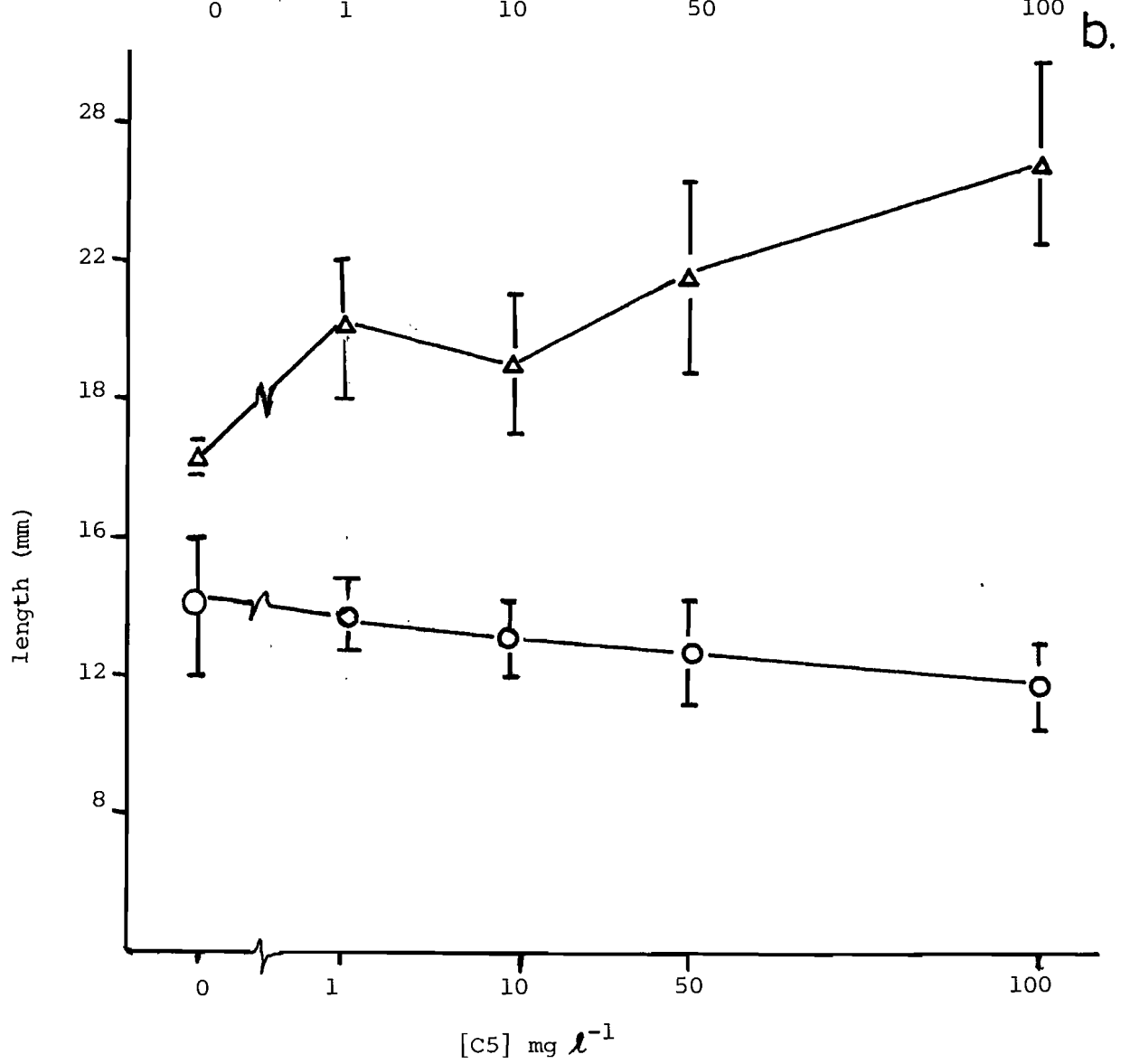
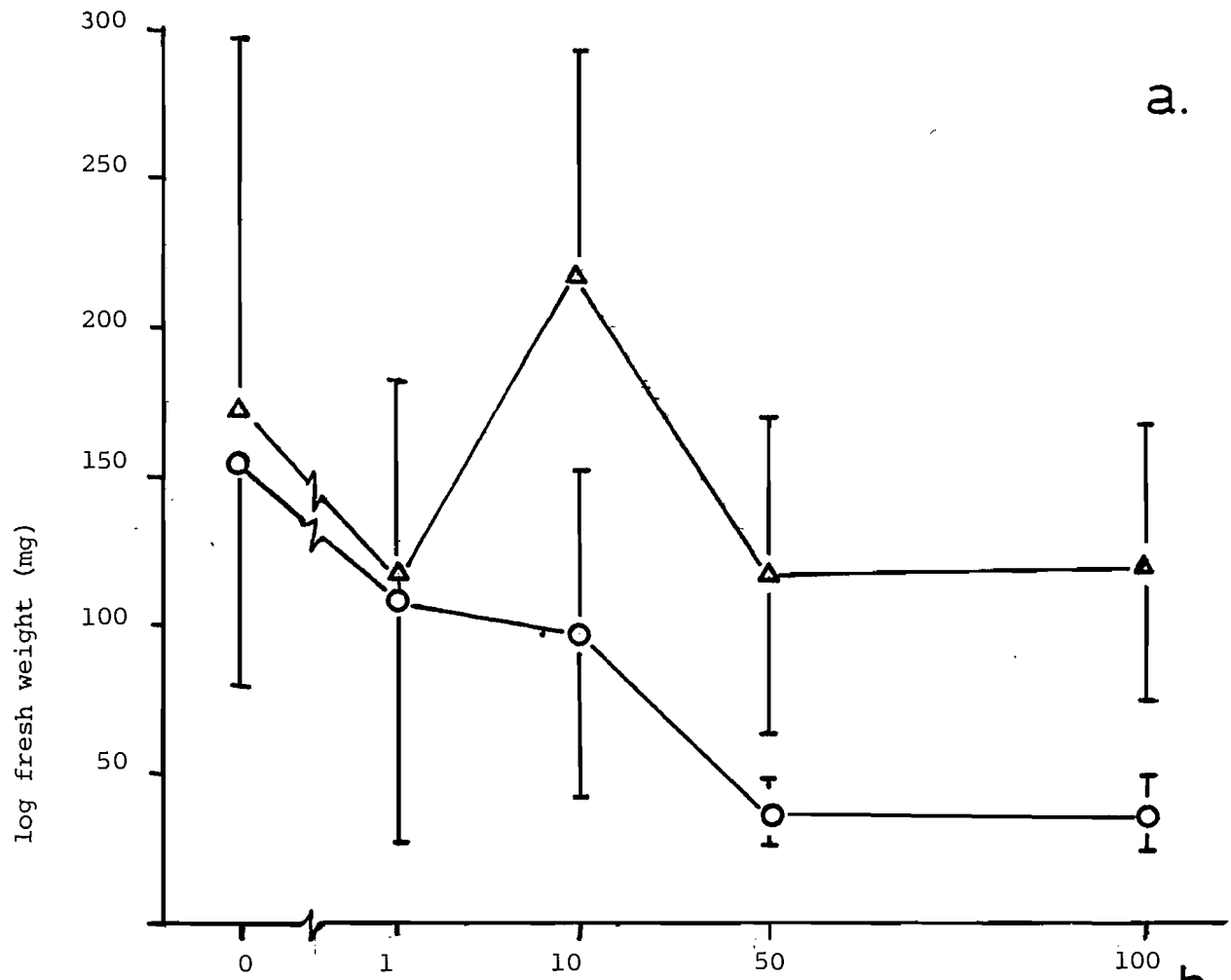


Figure 138. Effect of serial concentrations of C10 on (a) fresh weight and (b) length of *Populus yunnanensis* shoots cultured in vitro. Apparently single shoots were cultured on medium 2 containing C10 and maintained under LD photo-periods at 20°C for 4 weeks. The experiment was repeated twice and each value represents the mean \pm S.E. of twenty shoots. The vertical bars show one S.E. above and/or below the lines.

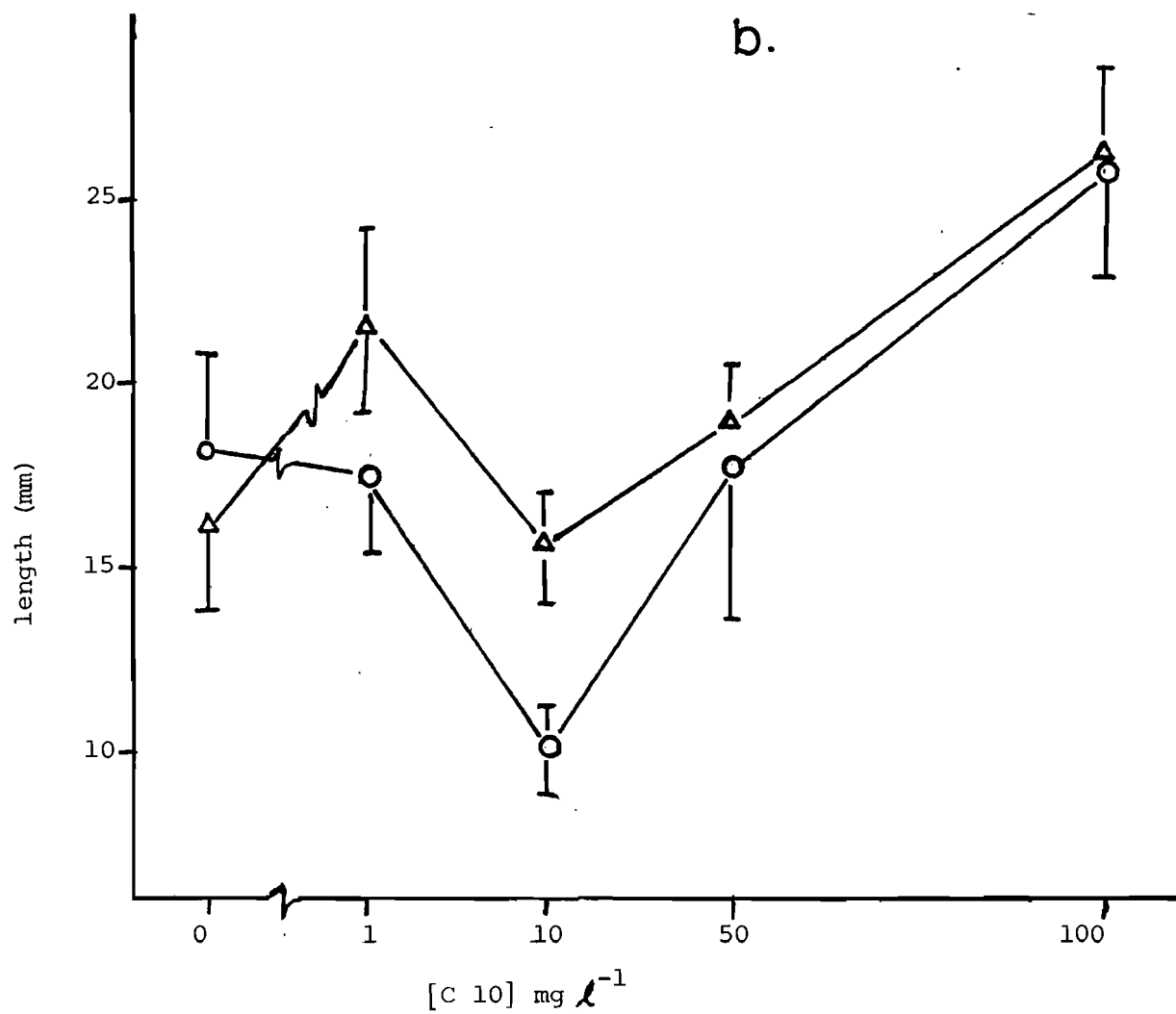
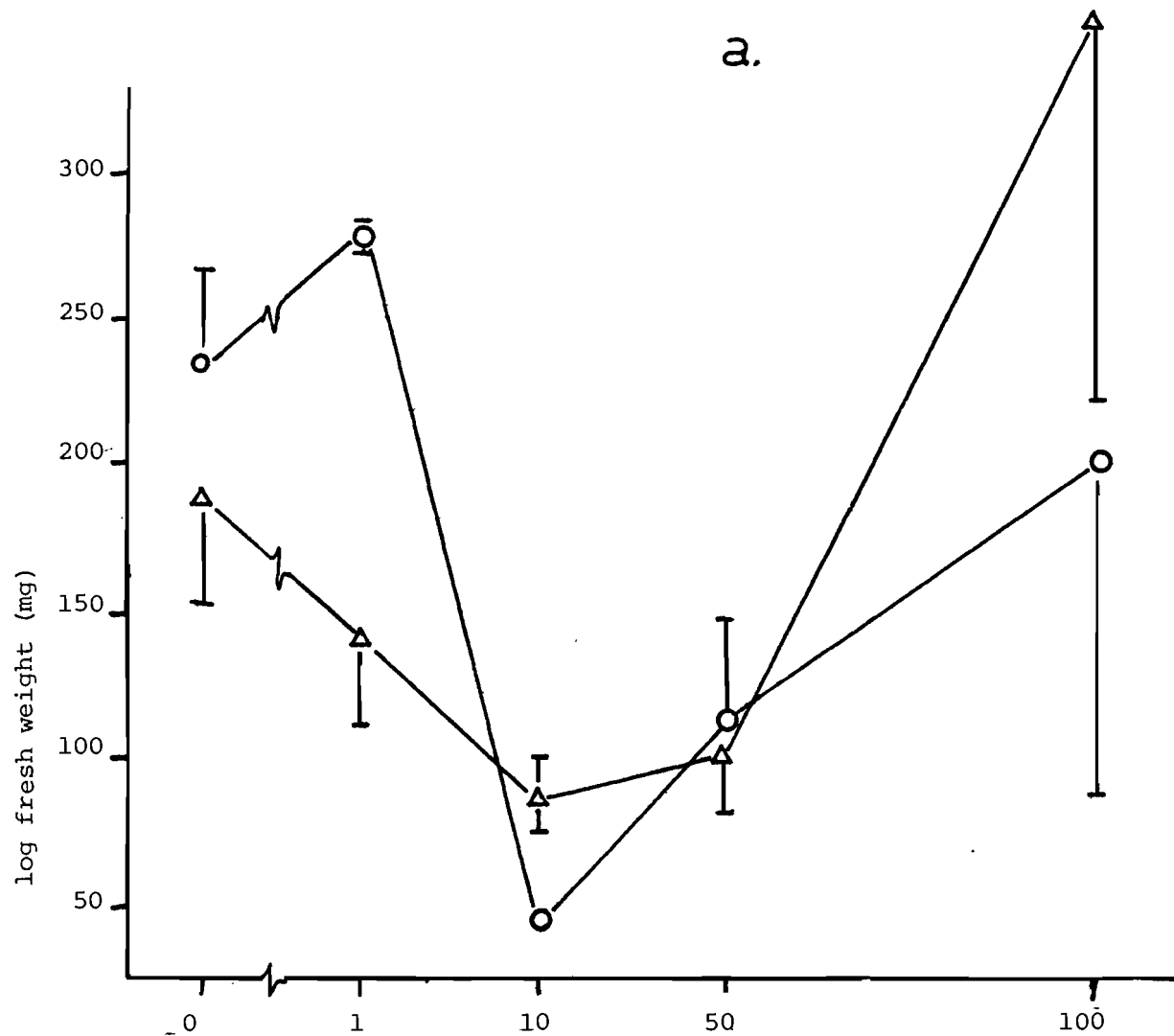


Plate 1. Growth of *Populus yunnanensis* shoots in vitro.

Shoots were cultured on medium 2 and maintained under LD's (16 h photoperiod) at 25°C.

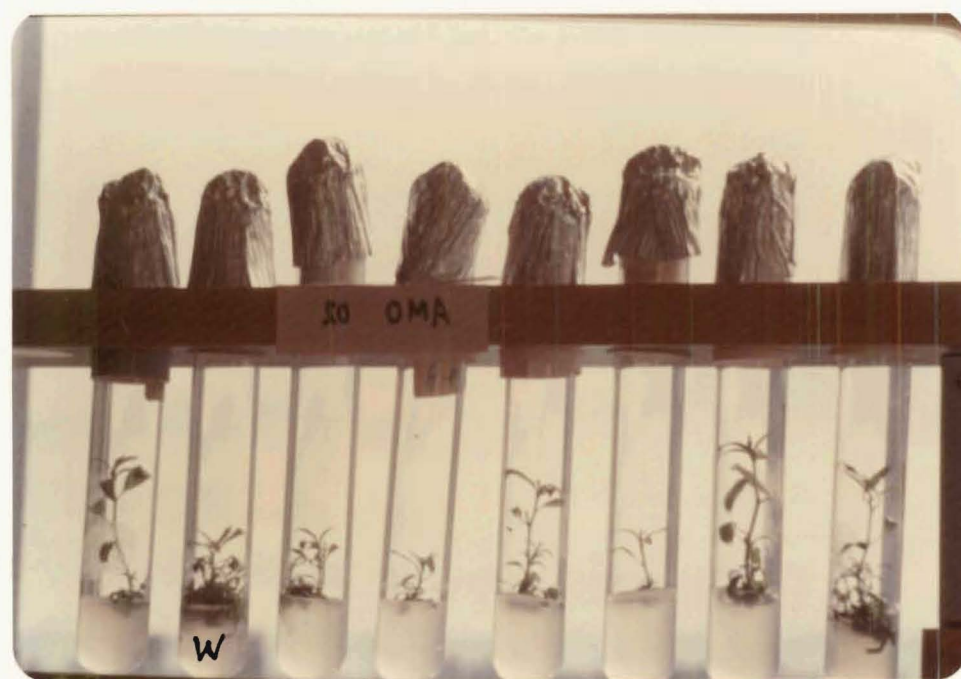
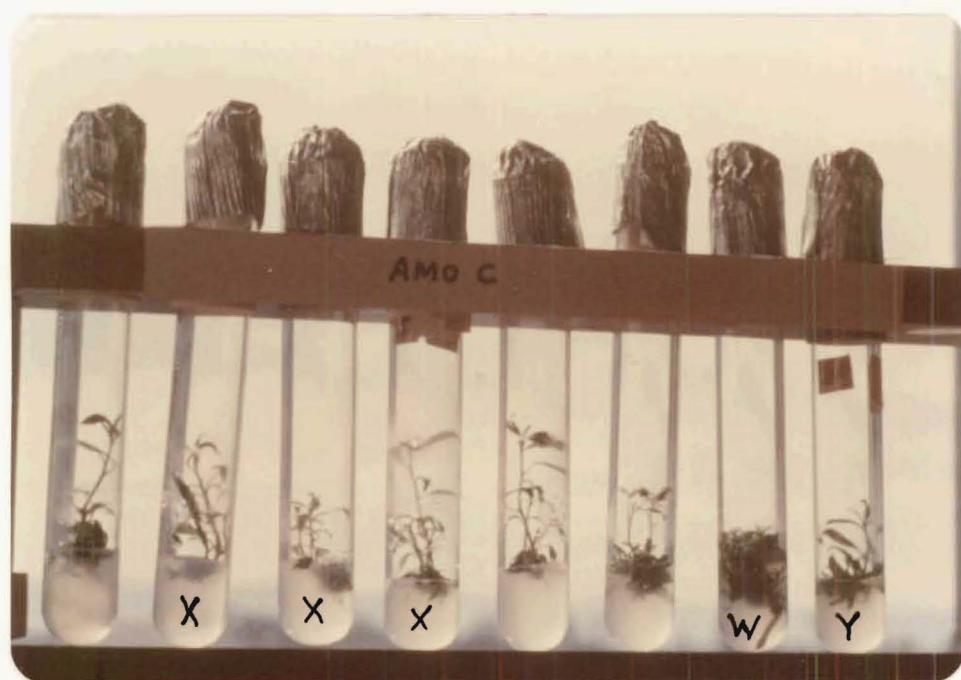


Plate 2A. Apical region of a *Populus yunnanensis* shoot cultured in vitro and maintained under LD's at 20°C. Note the new emerging leaf at the apex.

Plate 2B. Apical region of a *Populus yunnanensis* shoot cultured in vitro after 21 days of transfer from LD's to SD's. Note the cessation of extension growth resulting in the suppression of internodes and the absence of new leaves at the apex.



Plate 3. Effect of serial concentrations of ABA on the growth of *Populus yunnanensis* shoots cultured in vitro. Shoots were cultured on medium 2 plus ABA and maintained under LD's at 25°C.

Plate 4. Effect of serial concentrations of AMO 1618 on the growth of *Populus yunnanensis* shoots cultured in vitro.



conc.
mg l⁻¹

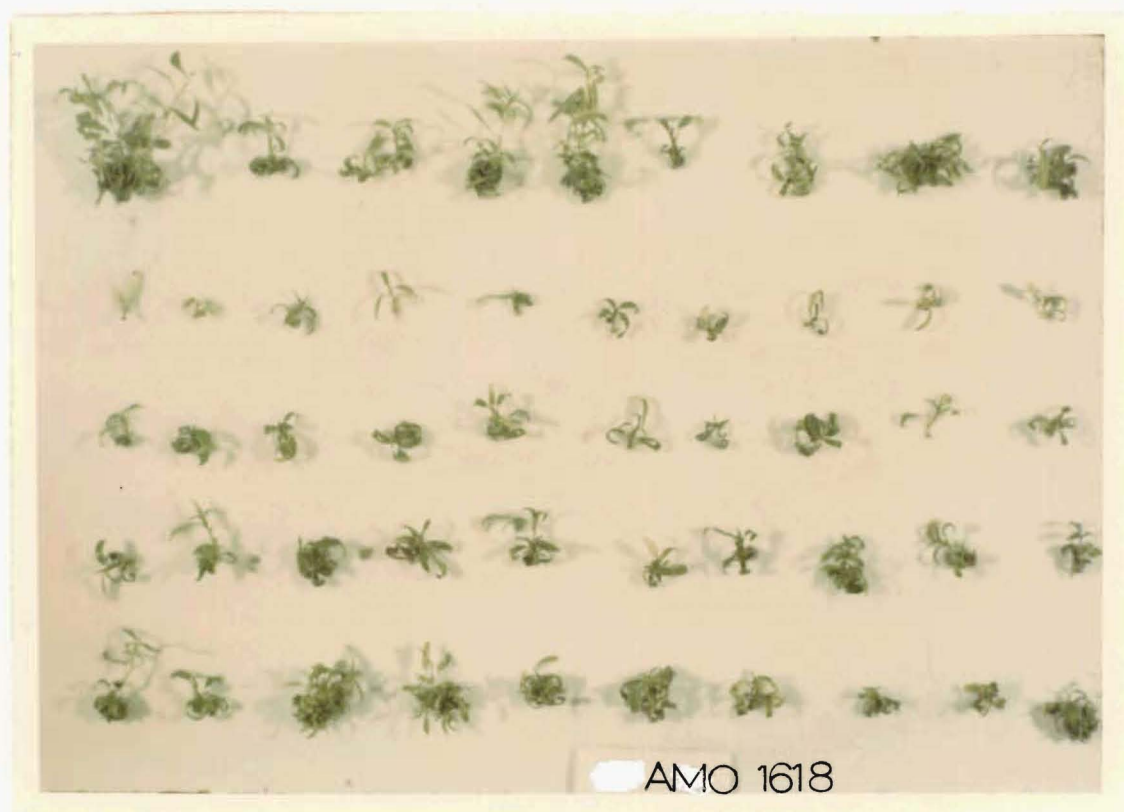
0

0.01

0.1

1

10



conc.
mg l⁻¹

0

0.01

0.1

1

10

Plate 5. Effect of serial concentrations of ABA + AMO 1618[®] on the growth of *Populus yunnanensis* shoots cultured in vitro.

Plate 6. Effect of serial concentrations of GA₃ on the growth of *Populus yunnanensis* shoots cultured in vitro.



conc.
mg l⁻¹

0

10

1

0.1

0.01



conc.
mg l⁻¹

0

0.01

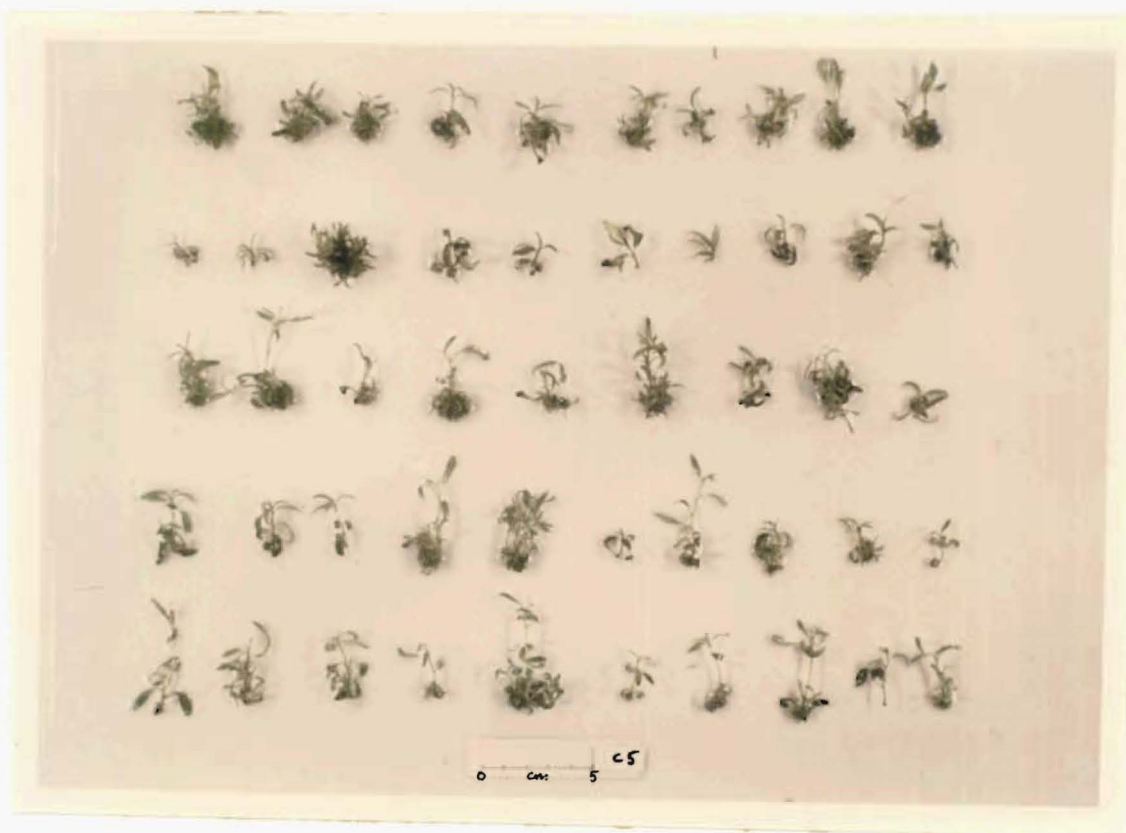
0.1

1

10

Plate 7. Effect of serial concentrations of C5 on the growth of *Populus yunnanensis* shoots cultured in vitro.

Plate 8. Effect of serial concentrations of C10 on the growth of *Populus yunnanensis* shoots cultured in vitro.



conc.
mg l⁻¹
0

100

50

10

1



conc.
mg l⁻¹
0

1

10

50

100