This paper reports a triple-layer, polydimethylsiloxane (PDMS)-based lab-on-a-chip platform combining the capture and culture of individual oomycete zoospores with integrated force sensing on germinated hyphae. The platform enables the concurrent study of cell-to-cell variability in hyphal growth and protrusive force generation. To demonstrate the applicability of the platform, individual zoospores of the oomycete Achlya bisexualis were trapped by a constriction structure, cultured on the device and the micro-Newton forces exerted by hyphae measured by tracking the deflection of elastomeric micropillars. The platform provides a new tool to help understand protrusive growth on a single cell level.

KEYWORDS
Lab-on-a-Chip, Single Cell Analysis, Membrane Valve, Force Sensor, Fungi and Oomycetes.

INTRODUCTION
Hyphal microorganisms, such as fungi and oomycetes, can act as pathogens and cause significant threat to both plants and animals [1]. Several species actively impact food security and primary industries by attacking crops [2] and forestry plants [3], both which can lead to major economic losses. The spread of these diseases is further compounded by climate change [4], emergence of drug resistance [5] and world-wide dispersion by the globalization of human activities [6]. Recent years have seen increasing efforts to screen naturally bioactive compounds and develop new biocontrol strategies alternative to agrochemicals [7].

Common to all these activities is the need to screen a large number of potential candidates for their impact on fungal and oomycete growth and penetrative force generation at various stages in their respective life cycles. One such crucial stage is the germination of hyphae from either zoospores or conidia, leading to penetration into and infection of parts of a plant or animal [8]. This capability of tips growing from fungal hyphae to penetrate throughout host tissue is essential for pathogenicity [8]. The process is driven by cytoskeletal changes [9] and the yielding of hyphal tip to the internal hydrostatic pressure (turgor), which manifests externally as the generation of a protrusive force [10].

To better understand protrusive force generation by hyphal organisms, we have demonstrated force sensing technology based on elastomeric micropillars [11]. We used this technology to measure forces exerted by hyphae of the oomycete Achlya bisexualis [12] and the fungi Neurospora crassa [13] growing from mycelia seeded into monolithic polydimethylsiloxane (PDMS)-based devices. In the current work we extend on this by combining our in-channel high-aspect ratio sensing pillars for protrusive force measurements on hyphal organisms [14] with zoospore trapping, on-chip germination and maintenance [15]. This allows for high-throughput compartmentalization of individual zoospores or conidia into separate force measurement channels for parallelized screening, taking into account cellular heterogeneity, nuclear distribution and dynamics at the sub-hyphal level in response to exposure to various biocontrol strategies.

EXPERIMENTAL
Platform design
The monolithic PDMS platform consists of two layers separated by a membrane of 15 µm thickness, as shown in Fig. 1(a). A gas layer on the top provides membrane valve control for zoospore sorting. The fluidic layer on the bottom is composed of parallel measurement channels, each containing a zoospore trap-site with a constriction structure in the entrance, shown in Fig. 1(b), and a freely bending elastomeric micropillar (diameter 7 µm, height 24 µm) as force sensor. Width and height of the measurement channels were 35 and 30 µm, respectively. Zoospores are introduced from the zoospore inlet, flow along the loading channel and are captured in a trap site (Fig. 1(b)). Once all sites are occupied, the gas layer is used to actuate the membrane valve, which closes the channel and prevents trapped zoospores from escaping (Fig. 1(e)). The hyphae then grow through the measurement channels and deflect their respective force...
sensing pillars, as shown in Fig. 1(c) and (d). Meanwhile, media continues to be supplied from the media inlet to the end of the measurement channels, from where nutrients diffuse to the organisms.

**Platform fabrication**

Each PDMS layer was fabricated by replica-molding off photoresist masters and bonded via the central, spin-coated PDMS membrane. A two-layer photoresist master was fabricated for the fluidic layer by a combination of negative photoresist ADEX05 (DJ Microlaminates) and positive photoresist AZ 40XT (M.M.R.C Pty Ltd.), which provides a high-aspect freely-bending pillar inside a channel constriction [14].

For the gas layer master photolithography processing was used. In brief, a 4” chrome-on-glass photomask (Nanofilm) containing the gas layer pattern was prepared (μPG101, Heidelberg Instruments). Negative dry-film photoresist, ADEX 50 (thickness 50 μm, DJ Microlaminates) was laminated on a pre-cleaned 4” Si wafer. This was then exposed using UV lithography (MA-6, Suss) at an exposure dose of 325 mJ/cm² in vacuum contact mode with a filter (PL-360, Chroma). The wafer was developed in cyclohexanone, rinsed with isopropyl alcohol (IPA) and dried by N₂, followed by a ramped post-exposure bake of 5 min at 65 °C and 10 min at 95 °C. The master mold of the gas layer was completed by a hard-bake at 150 °C for 1 h.

Both PDMS fluidic and gas layers were molded into Sylgard 184 silicone elastomer (Dow Corning, 10:1 w/w). The two master molds were treated by vapor-coating with Trichloro (1H,1H,2H,2H-perfluorooctyl)silane (TFOCS, Sigma-Aldrich) to facilitate mold release. Pre-mixed Trichloro (1H,1H,2H,2H-perfluorooctyl)silane (TFOCS, Sigma-Aldrich) to facilitate mold release. Pre-mixed (10:1 w/w) PDMS was poured onto the molds and degassed. After a 2 h bake at 80 °C on a hotplate, the PDMS layers were peeled off carefully and 1 mm holes punched for all the inlets and outlets. Meanwhile, pre-mixed (10:1 w/w) PDMS was spin-coated onto a TFOCS-treated Si wafer (WS-650, Laurell) at 3000 rpm for 30 sec, achieving a thickness of 17 μm. The PDMS membrane was cured via a hard-bake at 150 °C for 1 h.

The fluidic layer of PDMS platform was first filled with the mineral salt solution. Zoospores were then introduced from the zoospore inlet via syringe pump (NE-300, New Era Pump Systems) and hydrodynamically captured in the trap-sites [15]. Valves were then shut off, while PYG broth continued to be supplied from the other side of the measurement channels. During maintenance, both valve and media supply were driven by a pressure controller (OB1, Elveflow). Growth of hyphae from zoospores in the measurement channels and micropillar deflection by the tip of each hypha were recorded using an optical microscope (Eclipse 80i, Nikon). The force generated by individual hyphae was obtained by a combination of image processing in ImageJ (V1.51 h, Fiji) and analysis in MATLAB (2016a, Mathworks) [12, 14].

**RESULTS**

**Lab-on-a-chip device**

As shown in Fig. 2, high-aspect ratio, in-channel force sensing pillars were successfully integrated with hydrodynamic traps on a monolithic PDMS valving platform. The device currently contains six parallel measurement channels, each containing a trap and sensor pillar, while a single large valve closes off the zoospore inlet channel. Device fabrication was complicated by the fact that pillar integration requires channels with rectangular cross-section, while normally-open PDMS valves require rounded channel profiles. The positive resist layer required to integrate the pillars [14] currently precludes the use of resist reflow to form rounded structures for complete valve closure [15], as this would also distort the pillar shape. As such, the current design uses PDMS sieve valves to retain zoospores in the constriction structures.

**Trapping and compartmentalization**

To demonstrate trapping and compartmentalization on...
the platform, polystyrene microspheres (20 μm diameter) suspended in water with red food coloring were first utilized. Figure 3 (a) and (b) show a single microsphere flowing into a measurement channel, closing of the constriction, and thus diverting successive microspheres to the next traps. After becoming trapped, the microsphere is retained in the trap site by closure of the membrane valve, as shown in Fig. 3 (c). DI water, provided to simulate media supply, faded the red color in the channel, but did not dislodge the microsphere (Fig. 3(d)). The same processes were applied to *A. bisexualis* zoospores, which range from 10 to 15 μm in diameter. Figure 3 (e) to (f) show an example of the trapping and successful retention of a zoospore during a media exchange cycle.

Following trapping, the growth of hyphae germinating from zoospores compartmentalized into separate channels was observed using optical microscopy, as shown in Fig. 4. Two types of constrictions were tested, a Type A (8 x 5 μm width x height, additional vertical constriction) and a Type B (10 x 30 μm width x height, no vertical constriction), both based on previous work [15] and shown in Fig. 4(d). As indicated in Fig. 4(c), we observed that the growth success ratio of zoospores captured by the horizontal only constriction Type B was much higher (7 out of 10 zoospores observed) than that of horizontal and vertical constriction Type A (2 out of 12 zoospores observed). This may indicate a yet to be determined mechanosensory response by the zoospores to the trap geometry.

### Integrated force sensing

Using the media exchange described above, trapped zoospores could be maintained on the platform until hyphae had germinated and extended to the force sensing pillars in their respective measurement channels. At this point, as described previously [12, 14], bright-field optical microscopy and computational image processing techniques were used to measure the forces exerted by hyphae on the pillars. Figure 5 illustrates that both magnitude and direction of the force exerted by single hyphae originating from single zoospores could be recorded as function of time. For the example hypha shown in Fig. 5(a), the force in X direction increased to 0.8 μN, while the force in Y direction fluctuated around 0.1 μN. As further illustrated by the plot of force versus time in Fig. 5(b), this significant increase in force perpendicular to the growth direction of this particular hyphae indicates a squeezing type event [12], where the hyphae is deflected into the gap between the pillar and channel wall and exerts force with the hyphal wall while traversing the gap.

Such squeezing type events predominantly occur for thinner hyphae and can be avoided by reducing the pillar-wall gap or growing hyphae to wider diameters [14]. We are currently investigating whether the position of the measurement pillar along the channel length can be used to tune the diameter of the hyphae. Previous work has indicated that, depending on the culture duration, hyphal length can be sensitive to channel length [15], but did not look at any correlation with channel width. In the future we...
will use the platform to characterize the heterogeneity of force generation in populations of zoospores and to screen the effect of biocontrol compounds on the ability of hyphae to project protrusive force.

CONCLUSIONS
We have demonstrated the fabrication, characterization and application of a microfluidic platform combining in-channel, high-aspect ratio sensing pillars for protrusive force measurements on hyphal organisms with zoospore trapping, on-chip germination and organism maintenance. The platform constitutes a new tool to study heterogeneity in force generation by hyphae grown from individual zoospores, as well as screen biocontrol agents and strategies against populations of zoospores.

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